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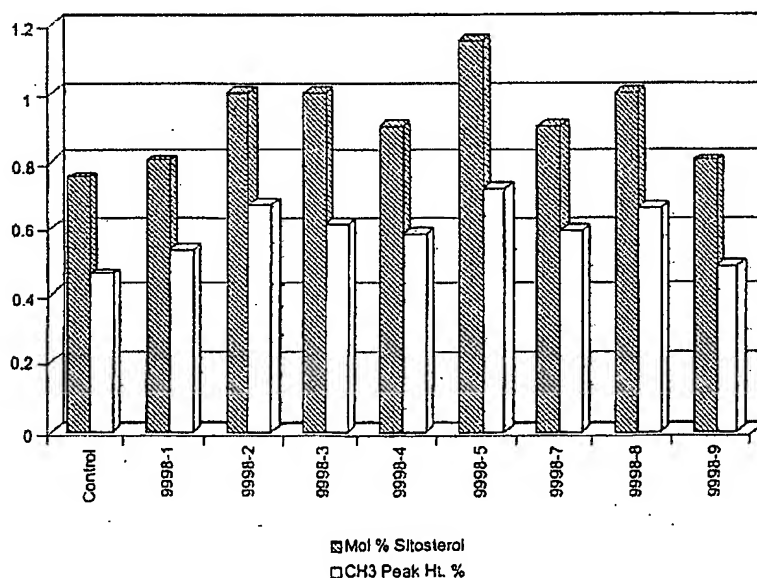
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(54) Title: PLANT STEROL ACYLTRANSFERASES



(57) Abstract: The present invention is directed to lecithin: cholesterol acyltransferase-like polypeptides (LCAT) and acyl CoA: cholesterol acyltransferases-like polypeptides (ACAT). The invention provides polynucleotides encoding such cholesterol: acyl-transferases-like polypeptides; polypeptides encoded by such polynucleotides, and the use of such polynucleotides to alter sterol composition and oil production in plants and host cells. Also provided are oils produced by the plants and host cells containing the polynucleotides and food products, nutritional supplements, and pharmaceutical composition containing plants or oils of the present invention. The polynucleotides of the present invention include those derived from plant sources.

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## PLANT STEROL ACYLTRANSFERASES

### CROSS-REFERENCE TO RELATED APPLICATIONS

- 5           This application claims priority to U.S. provisional application Serial No. 60/152,493, filed August 30, 1999 and herein incorporated by reference in its entirety for all purposes.

### BACKGROUND

10   Technical Field

The present invention is directed to plant acyltransferase-like nucleic acid and amino acid sequences and constructs, and methods related to their use in altering sterol composition and/or content, and oil composition and/or content in host cells and plants.

15   Related Art

- Through the development of plant genetic engineering techniques, it is now possible to produce transgenic varieties of plant species to provide plants which have novel and desirable characteristics. For example, it is now possible to genetically engineer plants for tolerance to environmental stresses, such as resistance to pathogens and tolerance to
- 20   herbicides. It is also possible to improve the nutritional characteristics of the plant, for example to provide improved fatty acid, carotenoid, sterol and tocopherol compositions. However, the number of useful nucleotide sequences for the engineering of such characteristics is thus far limited.

- There is a need for improved means to obtain or manipulate compositions of sterols
- 25   from biosynthetic or natural plant sources. The ability to increase sterol production or alter the sterol compositions in plants may provide for novel sources of sterols for use in human and animal nutrition.

- Sterol biosynthesis branches from the farnesyl diphosphate intermediate in the isoprenoid pathway. Sterol biosynthesis occurs via a mevalonate dependent pathway in
- 30   mammals and higher plants (Goodwin, (1981) *Biosynthesis of Isoprenoid Compounds*, vol 1 (Porter, J.W. & Spurgeon, S.L., eds) pp.443-480, John Wiley and Sons, New York), while in green algae sterol biosynthesis is thought to occur via a mevalonate independent pathway (Schwender, *et al.* (1997) *Physiology, Biochemistry, and Molecular Biology of*

*Plant Lipids*, (Williams, J.P., Khan, M.U., and Lem, N.W., eds) pp. 180-182, Kluwer Academic Publishers, Norwell, MA).

The solubility characteristics of sterol esters suggests that this is the storage form of sterols (Chang, *et al.*, (1997) *Annu. Rev. Biochem.*, 66:613-638). Sterol O-acyltransferase enzymes such as acyl CoA:cholesterol acyltransferase (ACAT) and lecithin:cholesterol acyltransferase (LCAT) catalyze the formation of cholesterol esters, and thus are key to controlling the intracellular cholesterol storage. In yeast, it has been reported that overexpression of *LROI*, a homolog of human LCAT, and phospholipid:diacylglycerol acyltransferase increased lipid synthesis (Oelkers *et al.*, (2000) *J. Biol. Chem.*, 26:15609-15612; Dahlqvist *et al.*, (2000) *Proc. Natl. Acad. Sci. USA*, 97:6487-6492).

The characterization of various acyltransferase proteins is useful for the further study of plant sterol synthesis systems and for the development of novel and/or alternative sterol sources. Studies of plant mechanisms may provide means to further enhance, control, modify, or otherwise alter the sterol composition of plant cells. Furthermore, such alterations in sterol content and/or composition may provide a means for obtaining tolerance to stress and insect damage. Of particular interest are the nucleic acid sequences of genes encoding proteins which may be useful for applications in genetic engineering.

### SUMMARY OF THE INVENTION

The present invention is directed to lecithin:cholesterol acyltransferase-like polypeptides (also referred to herein as LCAT) and acyl CoA:cholesterol acyltransferase-like polypeptides (also referred to herein as ACAT). In particular the invention is related to polynucleotides encoding such sterol:acyltransferases, polypeptides encoded by such polynucleotides, and the use of such polynucleotides to alter sterol composition and oil production. The polynucleotides of the present invention include those derived from plant sources.

One aspect of the invention, therefore, is an isolated nucleic acid sequence encoding a plant lecithin:cholesterol acyltransferase-like polypeptide, a fragment of a plant lecithin:cholesterol acyltransferase-like polypeptide, a plant acyl CoA:cholesterol acyltransferase-like polypeptide or a fragment of a plant acyl CoA:cholesterol acyltransferase-like polypeptide.

Another aspect provides an isolated nucleic acid sequence consisting essentially of SEQ ID NO: 2, 4, 6, 8, 10-29, 43-51, 73 or 75. Also provided is an isolated nucleic acid sequence consisting of SEQ ID NO: 2, 4, 6, 8, 10-29, 43-51, 73 or 75.

Still another aspect provides an isolated nucleic acid sequence comprising a polynucleotide selected from the group consisting of an isolated polynucleotide encoding a polypeptide of SEQ ID NO: 3 or SEQ ID NO: 3 with at least one conservative amino acid substitution; SEQ ID NO: 2; an isolated polynucleotide that has at least 70%, 80%, 90%,  
5 or 95% sequence identity with SEQ ID NO: 2; an isolated polynucleotide of at least 10 amino acids that hybridizes under stringent conditions to SEQ ID NO: 2; an isolated polynucleotide complementary to any of the foregoing; and an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 2 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.

10 Still another aspect provides an isolated nucleic acid sequence consisting essentially of a polynucleotide of the formula  $5' X-(R_1)_n-(R_2)_n-(R_3)_n-Y 3'$  where X is a hydrogen, Y is a hydrogen or a metal,  $R_1$  and  $R_2$  are any nucleic acid, n is an integer between 0-3000, and  $R_3$  is selected from the group consisting of an isolated polynucleotide encoding a polypeptide of SEQ ID NO: 3 or SEQ ID NO: 3 with at least one conservative  
15 amino acid substitution; SEQ ID NO: 2; an isolated polynucleotide that has at least 70%, 80%, 90%, or 95% sequence identity with SEQ ID NO: 2; an isolated polynucleotide of at least 10 amino acids that hybridizes under stringent conditions to SEQ ID NO: 2; an isolated polynucleotide complementary to any of the foregoing; and an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 2 and encodes a  
20 plant lecithin:cholesterol acyltransferase-like polypeptide.

Another aspect provides an isolated nucleic acid sequence comprising a polynucleotide selected from the group consisting of an isolated polynucleotide encoding a polypeptide of SEQ ID NO: 5 or SEQ ID NO: 5 with at least one conservative amino acid substitution; SEQ ID NO: 4; an isolated polynucleotide that has at least 70%, 80%, 90%,  
25 or 95% sequence identity with SEQ ID NO: 4; an isolated polynucleotide of at least 10 amino acids that hybridizes under stringent conditions to SEQ ID NO: 4; an isolated polynucleotide complementary to any of the foregoing; and an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 4 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.

30 Another aspect provides an isolated nucleic acid sequence consisting essentially of a polynucleotide of the formula  $5' X-(R_1)_n-(R_2)_n-(R_3)_n-Y 3'$  where X is a hydrogen, Y is a hydrogen or a metal,  $R_1$  and  $R_2$  are any nucleic acid, n is an integer between 0-3000, and  $R_3$  is selected from the group consisting of an isolated polynucleotide encoding a polypeptide of SEQ ID NO: 5 or SEQ ID NO: 5 with at least one conservative amino acid



substitution; SEQ ID NO: 4; an isolated polynucleotide that has at least 70%, 80%, 90%, or 95% sequence identity with SEQ ID NO: 4; an isolated polynucleotide of at least 10 amino acids that hybridizes under stringent conditions to SEQ ID NO: 4; an isolated polynucleotide complementary to any of the foregoing; and an isolated polynucleotide that  
5 hybridizes under stringent conditions to SEQ ID NO: 4 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.

Another aspect provides an isolated nucleic acid sequence comprising a polynucleotide selected from the group consisting of an isolated polynucleotide encoding a polypeptide of SEQ ID NO:7 or SEQ ID NO: 7 with at least one conservative amino acid  
10 substitution; SEQ ID NO: 6; an isolated polynucleotide that has at least 70%, 80%, 90%, or 95% sequence identity with SEQ ID NO: 6; an isolated polynucleotide of at least 10 amino acids that hybridizes under stringent conditions to SEQ ID NO: 6; an isolated polynucleotide complementary to any of the foregoing; and an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 6 and encodes a plant  
15 lecithin:cholesterol acyltransferase-like polypeptide.

Another aspect provides an isolated nucleic acid sequence consisting essentially of a polynucleotide of the formula 5' X-(R<sub>1</sub>)<sub>n</sub>-(R<sub>2</sub>)<sub>n</sub>-(R<sub>3</sub>)<sub>n</sub>-Y 3' where X is a hydrogen, Y is a hydrogen or a metal, R<sub>1</sub> and R<sub>2</sub> are any nucleic acid, n is an integer between 0,3000, and R<sub>2</sub> is selected from the group consisting of an isolated polynucleotide encoding a  
20 polypeptide of SEQ ID NO: 7 or SEQ ID NO: 7 with at least one conservative amino acid substitution; SEQ ID NO: 6; an isolated polynucleotide that has at least 70%, 80%, 90%, or 95% sequence identity with SEQ ID NO: 6; an isolated polynucleotide of at least 10 amino acids that hybridizes under stringent conditions to SEQ ID NO: 6; an isolated polynucleotide complementary to any of the foregoing; and an isolated polynucleotide that  
25 hybridizes under stringent conditions to SEQ ID NO: 6 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.

Another aspect provides an isolated nucleic acid sequence comprising a polynucleotide selected from the group consisting of an isolated polynucleotide encoding a polypeptide of SEQ ID NO:9 or SEQ ID NO: 9 with at least one conservative amino acid  
30 substitution; SEQ ID NO: 8; an isolated polynucleotide that has at least 70%, 80%, 90%, or 95% sequence identity with SEQ ID NO: 8; an isolated polynucleotide of at least 10 amino acids that hybridizes under stringent conditions to SEQ ID NO: 8; an isolated polynucleotide complementary to any of the foregoing; and an isolated polynucleotide that

hybridizes under stringent conditions to SEQ ID NO: 8 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.

Another aspect provides an isolated nucleic acid sequence consisting essentially of a polynucleotide of the formula 5' X-(R<sub>1</sub>)<sub>n</sub>-(R<sub>2</sub>)<sub>n</sub>-(R<sub>3</sub>)<sub>n</sub>-Y 3' where X is a hydrogen, Y is a hydrogen or a metal, R<sub>1</sub> and R<sub>2</sub> are any nucleic acid, n is an integer between 0-3000, and R<sub>3</sub> is selected from the group consisting of an isolated polynucleotide encoding a polypeptide of SEQ ID NO: 9 or SEQ ID NO: 9 with at least one conservative amino acid substitution; SEQ ID NO: 8; an isolated polynucleotide that has at least 70%, 80%, 90%, or 95% sequence identity with SEQ ID NO: 8; an isolated polynucleotide of at least 10 amino acids that hybridizes under stringent conditions to SEQ ID NO: 8; an isolated polynucleotide complementary to any of the foregoing; and an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 8 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.

Another aspect provides an isolated nucleic acid sequence comprising a polynucleotide selected from the group consisting of an isolated polynucleotide encoding a polypeptide of SEQ ID NO: 74 or SEQ ID NO: 74 with at least one conservative amino acid substitution; SEQ ID NO: 73; an isolated polynucleotide that has at least 70%, 80%, 90%, or 95% sequence identity with SEQ ID NO: 73; an isolated polynucleotide of at least 10 amino acids that hybridizes under stringent conditions to SEQ ID NO: 73; an isolated polynucleotide complementary to any of the foregoing; and an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 73 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.

Another aspect provides an isolated nucleic acid sequence consisting essentially of a polynucleotide of the formula 5' X-(R<sub>1</sub>)<sub>n</sub>-(R<sub>2</sub>)<sub>n</sub>-(R<sub>3</sub>)<sub>n</sub>-Y 3' where X is a hydrogen, Y is a hydrogen or a metal, R<sub>1</sub> and R<sub>2</sub> are any nucleic acid, n is an integer between 0-3000, and R<sub>3</sub> is selected from the group consisting of an isolated polynucleotide encoding a polypeptide of SEQ ID NO: 74 or SEQ ID NO: 74 with at least one conservative amino acid substitution; SEQ ID NO: 73; an isolated polynucleotide that has at least 70%, 80%, 90%, or 95% sequence identity with SEQ ID NO: 73; an isolated polynucleotide of at least 10 amino acids that hybridizes under stringent conditions to SEQ ID NO: 73; an isolated polynucleotide complementary to any of the foregoing; and an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 73 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.

Another aspect provides an isolated nucleic acid sequence comprising a polynucleotide selected from the group consisting of an isolated polynucleotide encoding a polypeptide of SEQ ID NO: 76 or SEQ ID NO: 76 with at least one conservative amino acid substitution; SEQ ID NO: 75; an isolated polynucleotide that has at least 70%, 80%,  
5 90%, or 95% sequence identity with SEQ ID NO: 75; an isolated polynucleotide of at least 10 amino acids that hybridizes under stringent conditions to SEQ ID NO: 75; an isolated polynucleotide complementary to any of the foregoing; and an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 75 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.

10 Another aspect provides an isolated nucleic acid sequence consisting essentially of a polynucleotide of the formula 5' X-(R<sub>1</sub>)<sub>n</sub>-(R<sub>2</sub>)<sub>n</sub>-(R<sub>3</sub>)<sub>n</sub>-Y 3' where X is a hydrogen, Y is a hydrogen or a metal, R<sub>1</sub> and R<sub>2</sub> are any nucleic acid, n is an integer between 0-3000, and R<sub>3</sub> is selected from the group consisting of an isolated polynucleotide encoding a polypeptide of SEQ ID NO: 76 or SEQ ID NO: 76 with at least one conservative amino  
15 acid substitution; SEQ ID NO: 75; an isolated polynucleotide that has at least 70%, 80%, 90%, or 95% sequence identity with SEQ ID NO: 75; an isolated polynucleotide of at least 10 amino acids that hybridizes under stringent conditions to SEQ ID NO: 75; an isolated polynucleotide complementary to any of the foregoing; and an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 75 and encodes a plant  
20 lecithin:cholesterol acyltransferase-like polypeptide.

Another aspect provides an isolated nucleic acid sequence comprising a polynucleotide selected from the group consisting of SEQ ID NO: 42 or a degenerate variant thereof; an isolated polynucleotide that has at least 70%, 80%, 90%, or 95% sequence identity with SEQ ID NO: 42; an isolated polynucleotide of at least 10 amino  
25 acids that hybridizes under stringent conditions to SEQ ID NO: 42; an isolated polynucleotide complementary to any of the foregoing; and an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 42 and encodes an acyl CoA:cholesterol acyltransferase-like polypeptide.

Another aspect provides an isolated nucleic acid sequence consisting essentially of  
30 a polynucleotide of the formula 5' X-(R<sub>1</sub>)<sub>n</sub>-(R<sub>2</sub>)<sub>n</sub>-(R<sub>3</sub>)<sub>n</sub>-Y 3' where X is a hydrogen, Y is a hydrogen or a metal, R<sub>1</sub> and R<sub>2</sub> are any nucleic acid, n is an integer between 0-3000, and R<sub>3</sub> is selected from the group consisting of SEQ ID NO: 42 or a degenerate variant thereof; an isolated polynucleotide that has at least 70%, 80%, 90%, or 95% sequence identity with SEQ ID NO: 42; an isolated polynucleotide of at least 10 amino acids that hybridizes

under stringent conditions to SEQ ID NO: 42; an isolated polynucleotide complementary to any of the foregoing; and an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 42 and encodes a acyl CoA:cholesterol acyltransferase-like polypeptide.

5 Also provided is a recombinant nucleic acid construct comprising a regulatory sequence operably linked to a polynucleotide encoding a lecithin:cholesterol acyltransferase-like polypeptide and/or an acyl CoA:cholesterol acyltransferase-like polypeptide. In one embodiment, the sterol acyl transferases are plant sterol acyl transferases. In another embodiment, the recombinant nucleic acid constructs further  
10 comprises a termination sequence. The regulatory sequence can be a constitutive promoter, an inducible promoter, a developmentally regulated promoter, a tissue specific promoter, an organelle specific promoter, a seed specific promoter or a combination of any of the foregoing. Also provided is a plant containing this recombinant nucleic acid construct and the seed and progeny from such a plant. This recombinant nucleic acid  
15 construct can also be introduced into a suitable host cell to provide yet another aspect of the invention. If the host cell is a plant host cell, the cell can be used to generate a plant to provide another aspect of the invention. Further aspects include seed and progeny from such a plant.

Yet another aspect is a purified polypeptide comprising, SEQ ID NO: 3, SEQ ID  
20 NO: 5, SEQ ID NO: 7, SEQ ID NO: 9, SEQ ID NO: 74, SEQ ID NO: 76, or any of the preceding sequences with at least one conservative amino acid substitution.

Still another aspect provides a purified immunogenic polypeptide comprising at least 10 consecutive amino acids from an amino acid sequence selected from the group consisting of SEQ ID NO: 3, 5, 7, 9, 74, 76 and any of the preceding sequences containing  
25 at least one conservative amino acid substitution. Also provided are antibodies, either polyclonal or monoclonal, that specifically bind the preceding immunogenic polypeptides.

One aspect provides a method for producing a lecithin:cholesterol acyltransferase-like polypeptide or an acyl CoA:cholesterol acyltransferase-like polypeptide comprising culturing a host cell containing any recombinant nucleic acid construct of the present  
30 invention under condition permitting expression of said lecithin:cholesterol acyltransferase-like polypeptide or acyl CoA:cholesterol acyltransferase-like polypeptide.

Another aspect provides a method for modifying the sterol content of a host cell, comprising transforming a host cell with a recombinant construct containing a regulatory sequence operably linked to a polynucleotide encoding a lecithin:cholesterol

acyltransferase-like polypeptide and culturing said host cell under conditions wherein said host cell expresses a lecithin:cholesterol acyltransferase-like polypeptide such that said host cell has a modified sterol composition as compared to host cells without the recombinant construct.

5           An additional aspect is a method for modifying the sterol content of a host cell comprising transforming a host cell with a recombinant construct containing a regulatory sequence operably linked to a polynucleotide encoding an acyl CoA:cholesterol acyltransferase-like polypeptide and culturing said host cell under conditions wherein said host cell expresses an acyl CoA:cholesterol acyltransferase-like polypeptide such that said  
10   host cell has a modified sterol composition as compared to host cells without the recombinant construct.

          A further aspect is a plant comprising a recombinant construct containing a regulatory sequence operably linked to a polynucleotide encoding a lecithin:cholesterol acyltransferase-like polypeptide wherein expression of said recombinant construct results  
15   in modified sterol composition of said plant as compared to the same plant without said recombinant construct.

          Another aspect provides a plant comprising a recombinant construct containing a regulatory sequence operably linked to a polynucleotide encoding an acyl CoA:cholesterol acyltransferase-like polypeptide wherein expression of said recombinant construct results  
20   in modified sterol composition of said plant as compared to the same plant without said recombinant construct.

          In a further aspect is provided an oil obtained from any of the plants or host cells of the present invention.

          In still another aspect is provided a method for producing an oil with a modified  
25   sterol composition comprising providing any of the plants or host cells of the present invention and extracting oil from the plant by any known method. Also provided is an oil produced by the preceding method.

          Still another aspect provides a method for altering oil production by a host cell comprising, transforming a host cell with a recombinant construct containing a regulatory  
30   sequence operably linked to a polynucleotide encoding a lecithin:cholesterol acyltransferase-like polypeptide and culturing the host cell under conditions wherein the host cell expresses a lecithin:cholesterol acyltransferase-like polypeptide such that the host cell has an altered oil production as compared to host cells without the recombinant construct.

Another aspect provides a method for altering oil production by a host cell comprising, transforming a host cell with a recombinant construct containing a regulatory sequence operably linked to a polynucleotide encoding an acyl CoA:cholesterol acyltransferase-like polypeptide and culturing the host cell under conditions wherein the host cell expresses an acyl CoA:cholesterol acyltransferase-like polypeptide such that the host cell has an altered oil production as compared to host cells without the recombinant construct.

Also provided is a plant comprising a recombinant construct containing a regulatory sequence operably linked to a polynucleotide encoding a lecithin:cholesterol acyltransferase-like polypeptide wherein expression of said recombinant construct results in an altered production of oil by said plant as compared to the same plant without said recombinant construct.

In a further aspect is provided a plant comprising a recombinant construct containing a regulatory sequence operably linked to a polynucleotide encoding an acyl CoA:cholesterol acyltransferase-like polypeptide wherein expression of said recombinant construct results in an altered production of oil by said plant as compared to the same plant without said recombinant construct.

Additional aspects provide a food, food ingredient or food product comprising any oil, plant or host cell of the present invention; a nutritional or dietary supplement comprising any oil, plant or host cell of the present invention; and a pharmaceutical composition comprising any oil, plant or host cell of the present invention along with a suitable diluent, carrier or excipient.

Additional aspects will be apparent from the descriptions and examples that follow.

## BRIEF DESCRIPTION OF THE DRAWINGS

These and other features, aspects, and advantages of the present invention will become better understood with regard to the following description, appended claims and accompanying figures where:

Figure 1 shows an alignment of yeast, human and rat lecithin:cholesterol acyltransferase protein sequences with *Arabidopsis* LCAT1, LCAT2, LCAT3, and LCAT4 deduced amino acid sequences.

Figure 2 shows the results of NMR sterol ester analysis on T2 seed from plant expressing LCAT4 under the control of the napin promoter (pCGN9998).

Figure 3 shows the results of HPLC/MS sterol analysis on oil extracted from T2 seed from control lines (pCGN8640) and lines expressing LCAT3 (pCGN9968) under the control of the napin promoter.

Figure 4 shows the results of HPLC/MS sterol analysis on oil extracted from T2 seed from control lines (pCGN8640), and plant line expressing LCAT1 (pCGN9962), LCAT2 (pCGN9983), LCAT3 (pCGN9968), and LCAT4 (pCGN9998) under the control of the napin promoter. Additionally, data from 3 lines expressing LCAT4 under the control of the 35S promoter (pCGN9996) are shown.

Figure 5 shows the results of Nir analysis of the oil content of T2 seed from control lines (pCGN8640), and plant lines expressing LCAT1 (pCGN9962), LCAT2 (pCGN9983), and LCAT3 (pCGN9968) under the control of the napin promoter. Additionally, data from 16 lines expressing LCAT2 under the control of the 35S promoter (pCGN9981) are shown.

#### DETAILED DESCRIPTION

The following detailed description is provided to aid those skilled in the art in practicing the present invention. Even so, this detailed description should not be construed to unduly limit the present invention as modifications and variations in the embodiments discussed herein can be made by those of ordinary skill in the art without departing from the spirit or scope of the present inventive discovery.

All publications, patents, patent applications and other references cited in this application are herein incorporated by reference in their entirety as if each individual publication, patent, patent application or other reference were specifically and individually indicated to be incorporated by reference.

The present invention relates to lecithin:cholesterol acyltransferase, particularly the isolated nucleic acid sequences encoding lecithin:cholesterol-like polypeptides (LCAT) from plant sources and acyl CoA:cholesterol:acyltransferase, particularly the isolated nucleic acid sequences encoding acyl CoA:cholesterol acyltransferase-like polypeptides (ACAT) from plant sources. Lecithin:cholesterol acyltransferase-like as used herein includes any nucleic acid sequence encoding an amino acid sequence from a plant source, such as a protein, polypeptide or peptide, obtainable from a cell source, which demonstrates the ability to utilize lecithin (phosphatidyl choline) as an acyl donor for acylation of sterols or glycerides to form esters under enzyme reactive conditions along with such proteins polypeptides and peptides. Acyl CoA:cholesterol acyltransferase-like

as used herein includes any nucleic acid sequence encoding an amino acid sequence from a plant source, such as a protein, polypeptide or peptide, obtainable from a cell source, which demonstrates the ability to utilize acyl CoA as an acyl donor for acylation of sterols or glycerides to form esters under enzyme reactive conditions along with such proteins  
5 polypeptides and peptides. By "enzyme reactive conditions" is meant that any necessary conditions are available in an environment (i.e., such factors as temperature, pH, lack of inhibiting substances) which will permit the enzyme to function.

The term "sterol" as applied to plants refers to any chiral tetracyclic isopentenoid which may be formed by cyclization of squalene oxide through the transition state  
10 possessing stereochemistry similar to the *trans-syn-trans-anti-trans-anti* configuration, for example, protosteroid cation, and which retains a polar group at C-3 (hydroxyl or keto), an *all-trans-anti* stereochemistry in the ring system, and a side-chain 20R-configuration (Parker, *et al.* (1992) In Nes, *et al.*, Eds., *Regulation of Isopentenoid Metabolism*, ACS Symposium Series No. 497, p. 110; American Chemical Society, Washington, D.C.).

15 Sterols may or may not contain a C-5-C-6 double bond, as this is a feature introduced late in the biosynthetic pathway. Sterols contain a C<sub>8</sub>-C<sub>10</sub> side chain at the C-17 position.

The term "phytosterol," which applies to sterols found uniquely in plants, refers to a sterol containing a C-5, and in some cases a C-22, double bond. Phytosterols are further  
20 characterized by alkylation of the C-17 side-chain with a methyl or ethyl substituent at the C-24 position. Major phytosterols include, but are not limited to, sitosterol, stigmasterol, campesterol, brassicasterol, etc. Cholesterol, which lacks a C-24 methyl or ethyl side-chain, is found in plants, but is not unique thereto, and is not a "phytosterol."

"Phytostanols" are saturated forms of phytosterols wherein the C-5 and, when  
25 present, C-22 double bond(s) is (are) reduced, and include, but are not limited to, sitostanol, campestanol, and 22-dihydrobrassicastanol.

"Sterol esters" are further characterized by the presence of a fatty acid or phenolic acid moiety rather than a hydroxyl group at the C-3 position.

The term "sterol" includes sterols, phytosterols, phytosterol esters, phytostanols,  
30 and phytostanol esters.

The term "sterol compounds" includes sterols, phytosterols, phytosterol esters, phytostanols, and phytostanol esters.

The term "phytosterol compound" refers to at least one phytosterol, at least one phytosterol ester, or a mixture thereof.



The term "phytostanol compound" refers to at least one phytostanol, at least one phytostanol ester, or a mixture thereof.

The term "glyceride" refers to a fatty acid ester of glycerol and includes mono-, di-, and tri- acylglycerols.

5 As used herein, "recombinant construct" is defined either by its method of production or its structure. In reference to its method of production, e.g., a product made by a process, the process is use of recombinant nucleic acid techniques, e.g., involving human intervention in the nucleotide sequence, typically selection or production. Alternatively, in terms of structure, it can be a sequence comprising fusion of two or more  
10 nucleic acid sequences which are not naturally contiguous or operatively linked to each other

As used herein, "regulatory sequence" means a sequence of DNA concerned with controlling expression of a gene; e.g. promoters, operators and attenuators. A "heterologous regulatory sequence" is one which differs from the regulatory sequence  
15 naturally associated with a gene.

As used herein, "polynucleotide" and "oligonucleotide" are used interchangeably and mean a polymer of at least two nucleotides joined together by a phosphodiester bond and may consist of either ribonucleotides or deoxynucleotides.

As used herein, "sequence" means the linear order in which monomers appear in a  
20 polymer, for example, the order of amino acids in a polypeptide or the order of nucleotides in a polynucleotide.

As used herein, "polypeptide", "peptide", and "protein" are used interchangeably and mean a compound that consist of two or more amino acids that are linked by means of peptide bonds.

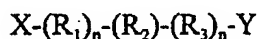
25 As used herein, the terms "complementary" or "complementarity" refer to the pairing of bases, purines and pyrimidines, that associate through hydrogen bonding in double stranded nucleic acids. For example, the following base pairs are complementary: guanine and cytosine; adenine and thymine; and adenine and uracil. The terms, as used herein, include complete and partial complementarity.

### Isolated proteins, Polypeptides and Polynucleotides

A first aspect of the present invention relates to isolated LCAT polynucleotides. The polynucleotide sequences of the present invention include isolated polynucleotides that encode the polypeptides of the invention having a deduced amino acid sequence  
5 selected from the group of sequences set forth in the Sequence Listing and to other polynucleotide sequences closely related to such sequences and variants thereof.

The invention provides a polynucleotide sequence identical over its entire length to each coding sequence as set forth in the Sequence Listing. The invention also provides the coding sequence for the mature polypeptide or a fragment thereof, as well as the coding  
10 sequence for the mature polypeptide or a fragment thereof in a reading frame with other coding sequences, such as those encoding a leader or secretory sequence, a pre-, pro-, or prepro- protein sequence. The polynucleotide can also include non-coding sequences, including for example, but not limited to, non-coding 5' and 3' sequences, such as the transcribed, untranslated sequences, termination signals, ribosome binding sites, sequences  
15 that stabilize mRNA, introns, polyadenylation signals, and additional coding sequence that encodes additional amino acids. For example, a marker sequence can be included to facilitate the purification of the fused polypeptide. Polynucleotides of the present invention also include polynucleotides comprising a structural gene and the naturally associated sequences that control gene expression.

20 The invention also includes polynucleotides of the formula:



wherein, at the 5' end, X is hydrogen, and at the 3' end, Y is hydrogen or a metal,  $R_1$  and  $R_3$  are any nucleic acid residue,  $n$  is an integer between 0 and 3000, preferably between 1 and 1000 and  $R_2$  is a nucleic acid sequence of the invention, particularly a nucleic acid  
25 sequence selected from the group set forth in the Sequence Listing and preferably SEQ ID NOs: 2, 4, 6, 8, 10-29, 33, 42-51, 73 and 75. In the formula,  $R_2$  is oriented so that its 5' end residue is at the left, bound to  $R_1$ , and its 3' end residue is at the right, bound to  $R_3$ . Any stretch of nucleic acid residues denoted by either R group, where R is greater than 1, may be either a heteropolymer or a homopolymer, preferably a heteropolymer.

30 The invention also relates to variants of the polynucleotides described herein that encode for variants of the polypeptides of the invention. Variants that are fragments of the polynucleotides of the invention can be used to synthesize full-length polynucleotides of the invention. Preferred embodiments are polynucleotides encoding polypeptide variants wherein 5 to 10, 1 to 5, 1 to 3, 2, 1 or no amino acid residues of a polypeptide sequence of

the invention are substituted, added or deleted, in any combination. Particularly preferred are substitutions, additions, and deletions that are silent such that they do not alter the properties or activities of the polynucleotide or polypeptide.

Further preferred embodiments of the invention that are at least 50%, 60%, or 70% identical over their entire length to a polynucleotide encoding a polypeptide of the invention, and polynucleotides that are complementary to such polynucleotides. More preferable are polynucleotides that comprise a region that is at least 80% identical over its entire length to a polynucleotide encoding a polypeptide of the invention and polynucleotides that are complementary thereto. In this regard, polynucleotides at least 90% identical over their entire length are particularly preferred, those at least 95% identical are especially preferred. Further, those with at least 97% identity are highly preferred and those with at least 98% and 99% identity are particularly highly preferred, with those at least 99% being the most highly preferred.

Preferred embodiments are polynucleotides that encode polypeptides that retain substantially the same biological function or activity as determined by the methods described herein as the mature polypeptides encoded by the polynucleotides set forth in the Sequence Listing.

The invention further relates to polynucleotides that hybridize to the above-described sequences. In particular, the invention relates to polynucleotides that hybridize under stringent conditions to the above-described polynucleotides. An example of stringent hybridization conditions is overnight incubation at 42°C in a solution comprising 50% formamide, 5x SSC (150 mM NaCl, 15 mM trisodium citrate), 50 mM sodium phosphate (pH 7.6), 5x Denhardt's solution, 10% dextran sulfate, and 20 micrograms/milliliter denatured, sheared salmon sperm DNA, followed by washing the hybridization support in 0.1x SSC at approximately 65°C. Also included are polynucleotides that hybridize under a wash stringency of 0.1X SSC or 0.1X SSPE (at 50°C. Other hybridization and wash conditions are well known and are exemplified in Sambrook, *et al.*, Molecular Cloning: A Laboratory Manual, Second Edition, Cold Spring Harbor, NY (1989), particularly Chapter 11.

The invention also provides a polynucleotide consisting essentially of a polynucleotide sequence obtainable by screening an appropriate library containing the complete gene for a polynucleotide sequence set for in the Sequence Listing under stringent hybridization conditions with a probe having the sequence of said polynucleotide

sequence or a fragment thereof; and isolating said polynucleotide sequence. Methods for screening libraries are well known in the art and can be found for example in Sambrook, *et al.*, *Molecular Cloning: A Laboratory Manual*, Second Edition, Cold Spring Harbor, NY (1989), particularly Chapter 8 and Ausubel *et al.*, *Short Protocols in Molecular Biology*, 3<sup>rd</sup> ed, Wiley and Sons, 1995, chapter 6. Nucleic acid sequences useful for obtaining such a polynucleotide include, for example, probes and primers as described herein and in particular SEQ ID NO: 2, 4, 6, 8, 10-29, 33, 42-51, 73 and 75. These sequences are particularly useful in screening libraries obtained from *Arabidopsis*, soybean and corn for sequences encoding lecithin:cholesterol acyltransferase and lecithin:cholesterol acyltransferase-like polypeptides and for screening libraries for sequences encoding acyl CoA:cholesterol acyl transferase and acyl CoA:cholesterol acyl transferase-like polypeptides.

As discussed herein regarding polynucleotide assays of the invention, for example, polynucleotides of the invention can be used as a hybridization probe for RNA, cDNA, or genomic DNA to isolate full length cDNAs or genomic clones encoding a polypeptide and to isolate cDNA or genomic clones of other genes that have a high sequence similarity to a polynucleotide set forth in the Sequence Listing and in particular SEQ ID NO: 2, 4, 6, 8, 10-29, 33, 42-51, 73 and 75. Such probes will generally comprise at least 15 bases. Preferably such probes will have at least 30 bases and can have at least 50 bases. Particularly preferred probes will have between 30 bases and 50 bases, inclusive.

The coding region of each gene that comprises or is comprised by a polynucleotide sequence set forth in the Sequence Listing may be isolated by screening using a DNA sequence provided in the Sequence Listing to synthesize an oligonucleotide probe. A labeled oligonucleotide having a sequence complementary to that of a gene of the invention is then used to screen a library of cDNA, genomic DNA or mRNA to identify members of the library which hybridize to the probe. For example, synthetic oligonucleotides are prepared which correspond to the LCAT EST sequences. The oligonucleotides are used as primers in polymerase chain reaction (PCR) techniques to obtain 5' and 3' terminal sequence of LCAT genes. Alternatively, where oligonucleotides of low degeneracy can be prepared from particular LCAT peptides, such probes may be used directly to screen gene libraries for LCAT gene sequences. In particular, screening of cDNA libraries in phage vectors is useful in such methods due to lower levels of background hybridization.

Typically, a LCAT sequence obtainable from the use of nucleic acid probes will show 60-70% sequence identity between the target LCAT sequence and the encoding sequence used as a probe. However, lengthy sequences with as little as 50-60% sequence identity may also be obtained. The nucleic acid probes may be a lengthy fragment of the nucleic acid sequence, or may also be a shorter, oligonucleotide probe. When longer nucleic acid fragments are employed as probes (greater than about 100 bp), one may screen at lower stringencies in order to obtain sequences from the target sample which have 20-50% deviation (i.e., 50-80% sequence homology) from the sequences used as probe. Oligonucleotide probes can be considerably shorter than the entire nucleic acid sequence encoding an LCAT enzyme, but should be at least about 10, preferably at least about 15, and more preferably at least about 20 nucleotides. A higher degree of sequence identity is desired when shorter regions are used as opposed to longer regions. It may thus be desirable to identify regions of highly conserved amino acid sequence to design oligonucleotide probes for detecting and recovering other related LCAT genes. Shorter probes are often particularly useful for polymerase chain reactions (PCR), especially when highly conserved sequences can be identified. (See, Gould, *et al.*, *PNAS USA* (1989) 86:1934-1938.).

Another aspect of the present invention relates to LCAT polypeptides. Such polypeptides include isolated polypeptides set forth in the Sequence Listing, as well as polypeptides and fragments thereof, particularly those polypeptides which exhibit LCAT activity and also those polypeptides which have at least 50%, 60% or 70% identity, preferably at least 80% identity, more preferably at least 90% identity, and most preferably at least 95% identity to a polypeptide sequence selected from the group of sequences set forth in the Sequence Listing, and also include portions of such polypeptides, wherein such portion of the polypeptide preferably includes at least 30 amino acids and more preferably includes at least 50 amino acids.

"Identity", as is well understood in the art, is a relationship between two or more polypeptide sequences or two or more polynucleotide sequences, as determined by comparing the sequences. In the art, "identity" also means the degree of sequence relatedness between polypeptide or polynucleotide sequences, as determined by the match between strings of such sequences. "Identity" can be readily calculated by known methods including, but not limited to, those described in *Computational Molecular Biology*, Lesk, A.M., ed., Oxford University Press, New York (1988); *Biocomputing: Informatics and Genome Projects*, Smith, D.W., ed., Academic Press, New York, 1993; *Computer Analysis*

of Sequence Data, Part I, Griffin, A.M. and Griffin, H.G., eds., Humana Press, New Jersey (1994); *Sequence Analysis in Molecular Biology*, von Heinje, G., Academic Press (1987); *Sequence Analysis Primer*, Gribskov, M. and Devereux, J., eds., Stockton Press, New York (1991); and Carillo, H., and Lipman, D., *SIAM J Applied Math*, 48:1073 (1988).

- 5 Methods to determine identity are designed to give the largest match between the sequences tested. Moreover, methods to determine identity are codified in publicly available programs. Computer programs which can be used to determine identity between two sequences include, but are not limited to, GCG (Devereux, J., et al., *Nucleic Acids Research* 12(1):387 (1984); suite of five BLAST programs, three designed for nucleotide  
10 sequences queries (BLASTN, BLASTX, and TBLASTX) and two designed for protein sequence queries (BLASTP and TBLASTN) (Coulson, *Trends in Biotechnology*, 12: 76-80 (1994); Birren, et al., *Genome Analysis*, 1: 543-559 (1997)). The BLAST X program is publicly available from NCBI and other sources (*BLAST Manual*, Altschul, S., et al., NCBI NLM NIH, Bethesda, MD 20894; Altschul, S., et al., *J. Mol. Biol.*, 215:403-410  
15 (1990)). The well known Smith Waterman algorithm can also be used to determine identity.

Parameters for polypeptide sequence comparison typically include the following:

Algorithm: Needleman and Wunsch, *J. Mol. Biol.* 48:443-453 (1970)

Comparison matrix: BLOSSUM62 from Hentikoff and Hentikoff, *Proc. Natl.*

- 20 *Acad. Sci USA* 89:10915-10919 (1992)

Gap Penalty: 12

Gap Length Penalty: 4

- A program which can be used with these parameters is publicly available as the  
"gap" program from Genetics Computer Group, Madison Wisconsin. The above  
25 parameters along with no penalty for end gap are the default parameters for peptide comparisons.

Parameters for polynucleotide sequence comparison include the following:

Algorithm: Needleman and Wunsch, *J. Mol. Biol.* 48:443-453 (1970)

Comparison matrix: matches = +10; mismatches = 0

- 30 Gap Penalty: 50

Gap Length Penalty: 3

A program which can be used with these parameters is publicly available as the  
"gap" program from Genetics Computer Group, Madison Wisconsin. The above  
parameters are the default parameters for nucleic acid comparisons.

The invention also includes polypeptides of the formula:



wherein, at the amino terminus, X is hydrogen, and at the carboxyl terminus, Y is hydrogen or a metal,  $R_1$  and  $R_3$  are any amino acid residue, n is an integer between 0 and 1000, and  $R_2$  is an amino acid sequence of the invention, particularly an amino acid sequence selected from the group set forth in the Sequence Listing and preferably SEQ ID NOs: 3, 5, 7, 9, 74 and 76. In the formula,  $R_2$  is oriented so that its amino terminal residue is at the left, bound to  $R_1$ , and its carboxy terminal residue is at the right, bound to  $R_3$ . Any stretch of amino acid residues denoted by either R group, where R is greater than 1, may be either a heteropolymer or a homopolymer, preferably a heteropolymer.

Polypeptides of the present invention include isolated polypeptides encoded by a polynucleotide comprising a sequence selected from the group of a sequence contained in SEQ ID NOs: 2, 4, 6, 8, 73 and 75.

The polypeptides of the present invention can be mature protein or can be part of a fusion protein.

Fragments and variants of the polypeptides are also considered to be a part of the invention. A fragment is a variant polypeptide which has an amino acid sequence that is entirely the same as part but not all of the amino acid sequence of the previously described polypeptides. The fragments can be "free-standing" or comprised within a larger polypeptide of which the fragment forms a part or a region, most preferably as a single continuous region. Preferred fragments are biologically active fragments which are those fragments that mediate activities of the polypeptides of the invention, including those with similar activity or improved activity or with a decreased activity. Also included are those polypeptides and polypeptide fragments that are antigenic or immunogenic in an animal, particularly a human and antibodies, either polyclonal or monoclonal that specifically bind the antigenic fragments. In one preferred embodiment, such antigenic or immunogenic fragments comprise at least 10 consecutive amino acids from the amino acid sequences disclosed herein or such sequences with at least one conservative amino acid substitution. In additional embodiments, such antigenic or immunogenic fragments comprise at least 15, at least 25, at least 50 or at least 100 consecutive amino acids from the amino acid sequences disclosed herein or such sequences with at least one conservative amino acid substitution. Methods for the production of antibodies from polypeptides and polypeptides conjugated to carrier molecules are well known in the art and can be found

for example in Ausubel et al., *Short Protocols in Molecular Biology*, 3<sup>rd</sup> ed., Wiley & Sons, 1995, particularly chapter 11.

5 Variants of the polypeptide also include polypeptides that vary from the sequences set forth in the Sequence Listing by conservative amino acid substitutions, substitution of a residue by another with like characteristics. Those of ordinary skill in the art are aware that modifications in the amino acid sequence of a peptide, polypeptide, or protein can result in equivalent, or possibly improved, second generation peptides, etc., that display equivalent or superior functional characteristics when compared to the original amino-acid  
10 sequences. Alterations can include amino acid insertions, deletions, substitutions, truncations, fusions, shuffling of subunit sequences, and the like, provided that the peptide sequences produced by such modifications have substantially the same functional properties as the naturally occurring counterpart sequences disclosed herein.

One factor that can be considered in making such changes is the hydropathic index  
15 of amino acids. The importance of the hydropathic amino acid index in conferring interactive biological function on a protein has been discussed by Kyte and Doolittle (*J. Mol. Biol.*, 157: 105-132, 1982). It is accepted that the relative hydropathic character of amino acids contributes to the secondary structure of the resultant protein. This, in turn, affects the interaction of the protein with molecules such as enzymes, substrates, receptors,  
20 DNA, antibodies, antigens, etc.

Based on its hydrophobicity and charge characteristics, each amino acid has been assigned a hydropathic index as follows: isoleucine (+4.5); valine (+4.2); leucine (+3.8); phenylalanine (+2.8); cysteine/cystine (+2.5); methionine (+1.9); alanine (+1.8); glycine (-0.4); threonine (-0.7); serine (-0.8); tryptophan (-0.9); tyrosine (-1.3); proline (-1.6);  
25 histidine (-3.2); glutamate/glutamine/aspartate/asparagine (-3.5); lysine (-3.9); and arginine (-4.5).

As is known in the art, certain amino acids in a peptide or protein can be substituted for other amino acids having a similar hydropathic index or score and produce a resultant peptide or protein having similar biological activity, i.e., which still retains  
30 biological functionality. In making such changes, it is preferable that amino acids having hydropathic indices within  $\pm 2$  are substituted for one another. More preferred substitutions are those wherein the amino acids have hydropathic indices within  $\pm 1$ . Most preferred substitutions are those wherein the amino acids have hydropathic indices within  $\pm 0.5$ .



Like amino acids can also be substituted on the basis of hydrophilicity. U.S. Patent No. 4,554,101 discloses that the greatest local average hydrophilicity of a protein, as governed by the hydrophilicity of its adjacent amino acids, correlates with a biological property of the protein. The following hydrophilicity values have been assigned to amino acids: arginine/lysine (+3.0); aspartate/glutamate (+3.0  $\pm$  1); serine (+0.3); asparagine/glutamine (+0.2); glycine (0); threonine (-0.4); proline (-0.5  $\pm$  1); alanine/histidine (-0.5); cysteine (-1.0); methionine (-1.3); valine (-1.5); leucine/isoleucine (-1.8); tyrosine (-2.3); phenylalanine (-2.5); and tryptophan (-3.4). Thus, one amino acid in a peptide, polypeptide, or protein can be substituted by another amino acid having a similar hydrophilicity score and still produce a resultant protein having similar biological activity, i.e., still retaining correct biological function. In making such changes, amino acids having hydropathic indices within  $\pm$ 2 are preferably substituted for one another, those within  $\pm$ 1 are more preferred, and those within  $\pm$ 0.5 are most preferred.

As outlined above, amino acid substitutions in the peptides of the present invention can be based on the relative similarity of the amino acid side-chain substituents, for example, their hydrophobicity, hydrophilicity, charge, size, etc. Exemplary substitutions that take various of the foregoing characteristics into consideration in order to produce conservative amino acid changes resulting in silent changes within the present peptides, etc., can be selected from other members of the class to which the naturally-occurring amino acid belongs. Amino acids can be divided into the following four groups: (1) acidic amino acids; (2) basic amino acids; (3) neutral polar amino acids; and (4) neutral non-polar amino acids. Representative amino acids within these various groups include, but are not limited to: (1) acidic (negatively charged) amino acids such as aspartic acid and glutamic acid; (2) basic (positively charged) amino acids such as arginine, histidine, and lysine; (3) neutral polar amino acids such as glycine, serine, threonine, cysteine, cystine, tyrosine, asparagine, and glutamine; and (4) neutral non-polar amino acids such as alanine, leucine, isoleucine, valine, proline, phenylalanine, tryptophan, and methionine. It should be noted that changes which are not expected to be advantageous can also be useful if these result in the production of functional sequences.

Variants that are fragments of the polypeptides of the invention can be used to produce the corresponding full length polypeptide by peptide synthesis. Therefore, these variants can be used as intermediates for producing the full-length polypeptides of the invention.

The polynucleotides and polypeptides of the invention can be used, for example, in the transformation of host cells, such as plant cells, animal cells, yeast cells, bacteria, bacteriophage, and viruses, as further discussed herein.

The invention also provides polynucleotides that encode a polypeptide that is a  
5 mature protein plus additional amino or carboxyl-terminal amino acids, or amino acids within the mature polypeptide (for example, when the mature form of the protein has more than one polypeptide chain). Such sequences can, for example, play a role in the processing of a protein from a precursor to a mature form, allow protein transport, shorten or lengthen protein half-life, or facilitate manipulation of the protein in assays or  
10 production. It is contemplated that cellular enzymes can be used to remove any additional amino acids from the mature protein.

A precursor protein, having the mature form of the polypeptide fused to one or more prosequences may be an inactive form of the polypeptide. The inactive precursors generally are activated when the prosequences are removed. Some or all of the  
15 prosequences may be removed prior to activation. Such precursor protein are generally called proproteins.

#### **Preparation of Expression Constructs and Methods of Use**

Of interest is the use of the nucleotide sequences in recombinant DNA constructs  
20 to direct the transcription or transcription and translation (expression) of the acyltransferase sequences of the present invention in a host cell. Of particular interest is the use of the polynucleotide sequences of the present invention in recombinant DNA constructs to direct the transcription or transcription and translation (expression) of the acyltransferase sequences of the present invention in a host plant cell.

The expression constructs generally comprise a regulatory sequence functional in a  
25 host cell operably linked to a nucleic acid sequence encoding a lecithin:cholesterol acyltransferase-like polypeptide or acyl CoA:cholesterol acyltransferase-like polypeptide of the present invention and a transcriptional termination region functional in a host plant cell. Of particular interest is the use of promoters (also referred to as transcriptional  
30 initiation regions) functional in plant host cells.

Those skilled in the art will recognize that there are a number of promoters which are functional in plant cells, and have been described in the literature including constitutive, inducible, tissue specific, organelle specific, developmentally regulated and environmentally regulated promoters. Chloroplast and plastid specific promoters,

chloroplast or plastid functional promoters, and chloroplast or plastid operable promoters are also envisioned.

One set of promoters are constitutive promoters such as the CaMV35S or FMV35S promoters that yield high levels of expression in most plant organs. Enhanced or  
5 duplicated versions of the CaMV35S and FMV35S promoters are useful in the practice of this invention (Odell, *et al.* (1985) *Nature* 313:810-812; Rogers, U.S. Patent Number 5,378, 619). Other useful constitutive promoters include, but are not limited to, the mannopine synthase (*mas*) promoter, the nopaline synthase (*nos*) promoter, and the octopine synthase (*ocs*) promoter.

10 Useful inducible promoters include heat-shock promoters (Ou-Lee *et al.* (1986) *Proc. Natl. Acad. Sci. USA* 83: 6815; Ainley *et al.* (1990) *Plant Mol. Biol.* 14: 949), a nitrate-inducible promoter derived from the spinach nitrite reductase gene (Back *et al.* (1991) *Plant Mol. Biol.* 17: 9), hormone-inducible promoters (Yamaguchi-Shinozaki *et al.* (1990) *Plant Mol. Biol.* 15: 905; Kares *et al.* (1990) *Plant Mol. Biol.* 15: 905), and  
15 light-inducible promoters associated with the small subunit of RuBP carboxylase and LHCP gene families (Kuhlemeier *et al.* (1989) *Plant Cell* 1: 471; Feinbaum *et al.* (1991) *Mol. Gen. Genet.* 226: 449; Weisshaar *et al.* (1991) *EMBO J.* 10: 1777; Lam and Chua (1990) *Science* 248: 471; Castresana *et al.* (1988) *EMBO J.* 7: 1929; Schulze-Lefert *et al.* (1989) *EMBO J.* 8: 651).

20 In addition, it may also be preferred to bring about expression of the acyltransferase gene in specific tissues of the plant, such as leaf, stem, root, tuber, seed, fruit, etc., and the promoter chosen should have the desired tissue and developmental specificity. Examples of useful tissue-specific, developmentally-regulated promoters include fruit-specific promoters such as the E4 promoter (Cordes *et al.* (1989) *Plant Cell*  
25 1:1025), the E8 promoter (Deikman *et al.* (1988) *EMBO J.* 7: 3315), the kiwifruit actinidin promoter (Lin *et al.* (1993) *PNAS* 90: 5939), the 2A11 promoter (Houck *et al.*, U.S. Patent 4,943,674), and the tomato pZ130 promoter (U.S. Patents 5,175, 095 and 5,530,185); the  $\beta$ -conglycinin 7S promoter (Doyle *et al.* (1986) *J. Biol. Chem.* 261: 9228; Slighon and Beachy (1987) *Planta* 172: 356), and seed-specific promoters (Knutzon *et al.*  
30 (1992) *Proc. Natl. Acad. Sci. USA* 89: 2624; Bustos *et al.* (1991) *EMBO J.* 10: 1469; Lam and Chua (1991) *J. Biol. Chem.* 266: 17131; Stayton *et al.* (1991) *Aust. J. Plant. Physiol.* 18: 507). Fruit-specific gene regulation is discussed in U.S. Patent 5,753,475. Other useful seed-specific promoters include, but are not limited to, the napin, phaseolin, zein, soybean trypsin inhibitor, 7S, ADR12, ACP, stearyl-ACP desaturase, oleosin,

*Lasquerella* hydroxylase, and barley aldose reductase promoters (Bartels (1995) *Plant J.* 7: 809-822), the EA9 promoter (U.S. Patent 5,420,034), and the Bce4 promoter (U.S. Patent 5,530,194). Useful embryo-specific promoters include the corn globulin 1 and oleosin promoters. Useful endosperm-specific promoters include the rice glutelin-1 promoter, the  
5 promoters for the low-pI  $\beta$  amylase gene (Amy32b) (Rogers et al. (1984) *J. Biol. Chem.* 259: 12234), the high-pI  $\beta$  amylase gene (Amy 64) (Khurseed et al. (1988) *J. Biol. Chem.* 263: 18953), and the promoter for a barley thiol protease gene ("Aleurain") (Whittier et al. (1987) *Nucleic Acids Res.* 15: 2515).

Of particular interest is the expression of the nucleic acid sequences of the present  
10 invention from transcription initiation regions which are preferentially expressed in a plant seed tissue. Examples of such seed preferential transcription initiation sequences include those sequences derived from sequences encoding plant storage protein genes or from genes involved in fatty acid biosynthesis in oilseeds. Examples of such promoters include the 5' regulatory regions from such genes as napin (Kridl et al., *Seed Sci. Res.* 1:209:219  
15 (1991)), phaseolin, zein, soybean trypsin inhibitor, ACP, stearyl-ACP desaturase, soybean  $\alpha'$  subunit of  $\beta$ -conglycinin (soy 7s, (Chen et al., *Proc. Natl. Acad. Sci.*, 83:8560-8564 (1986))) and oleosin. Seed-specific gene regulation is discussed in EP 0 255 378 B1 and U.S. Patents 5,420,034 and 5,608,152. Promoter hybrids can also be constructed to enhance transcriptional activity (Hoffman, U.S. Patent No. 5,106,739), or to combine  
20 desired transcriptional activity and tissue specificity.

It may be advantageous to direct the localization of proteins conferring LCAT to a particular subcellular compartment, for example, to the mitochondrion, endoplasmic reticulum, vacuoles, chloroplast or other plastidic compartment. For example, where the genes of interest of the present invention will be targeted to plastids, such as chloroplasts,  
25 for expression, the constructs will also employ the use of sequences to direct the gene to the plastid. Such sequences are referred to herein as chloroplast transit peptides (CTP) or plastid transit peptides (PTP). In this manner, where the gene of interest is not directly inserted into the plastid, the expression construct will additionally contain a gene encoding a transit peptide to direct the gene of interest to the plastid. The chloroplast transit  
30 peptides may be derived from the gene of interest, or may be derived from a heterologous sequence having a CTP. Such transit peptides are known in the art. See, for example, Von Heijne et al. (1991) *Plant Mol. Biol. Rep.* 9:104-126; Clark et al. (1989) *J. Biol. Chem.* 264:17544-17550; della-Cioppa et al. (1987) *Plant Physiol.* 84:965-968; Romer et al.

(1993) *Biochem. Biophys. Res Commun.* 196:1414-1421; and, Shah *et al.* (1986) *Science* 233:478-481.

Depending upon the intended use, the constructs may contain the nucleic acid sequence which encodes the entire LCAT protein, a portion of the LCAT protein, the entire ACAT protein, or a portion of the ACAT protein. For example, where antisense inhibition of a given LCAT or ACAT protein is desired, the entire sequence is not required. Furthermore, where LCAT or ACAT sequences used in constructs are intended for use as probes, it may be advantageous to prepare constructs containing only a particular portion of a LCAT or ACAT encoding sequence, for example a sequence which is discovered to encode a highly conserved region.

The skilled artisan will recognize that there are various methods for the inhibition of expression of endogenous sequences in a host cell. Such methods include, but are not limited to antisense suppression (Smith, *et al.* (1988) *Nature* 334:724-726), co-suppression (Napoli, *et al.* (1989) *Plant Cell* 2:279-289), ribozymes (PCT Publication WO 97/10328), and combinations of sense and antisense Waterhouse, *et al.* (1998) *Proc. Natl. Acad. Sci. USA* 95:13959-13964. Methods for the suppression of endogenous sequences in a host cell typically employ the transcription or transcription and translation of at least a portion of the sequence to be suppressed. Such sequences may be homologous to coding as well as non-coding regions of the endogenous sequence.

Regulatory transcript termination regions may be provided in plant expression constructs of this invention as well. Transcript termination regions may be provided by the DNA sequence encoding the diacylglycerol acyltransferase or a convenient transcription termination region derived from a different gene source, for example, the transcript termination region which is naturally associated with the transcript initiation region. The skilled artisan will recognize that any convenient transcript termination region which is capable of terminating transcription in a plant cell may be employed in the constructs of the present invention.

Alternatively, constructs may be prepared to direct the expression of the LCAT or ACAT sequences directly from the host plant cell plastid. Such constructs and methods are known in the art and are generally described, for example, in Svab, *et al.* (1990) *Proc. Natl. Acad. Sci. USA* 87:8526-8530 and Svab and Maliga (1993) *Proc. Natl. Acad. Sci. USA* 90:913-917 and in U.S. Patent Number 5,693,507.

A plant cell, tissue, organ, or plant into which the recombinant DNA constructs containing the expression constructs have been introduced is considered transformed,

transfected, or transgenic. A transgenic or transformed cell or plant also includes progeny of the cell or plant and progeny produced from a breeding program employing such a transgenic plant as a parent in a cross and exhibiting an altered phenotype resulting from the presence of a LCAT nucleic acid sequence.

5 Plant expression or transcription constructs having a plant LCAT as the DNA sequence of interest for increased or decreased expression thereof may be employed with a wide variety of plant life, particularly, plant life involved in the production of vegetable oils for edible and industrial uses. Most especially preferred are temperate oilseed crops. Plants of interest include, but are not limited to, rapeseed (Canola and High Erucic Acid  
10 varieties), sunflower, safflower, cotton, soybean, peanut, coconut and oil palms, and corn. Depending on the method for introducing the recombinant constructs into the host cell, other DNA sequences may be required. Importantly, this invention is applicable to dicotyledons and monocotyledons species alike and will be readily applicable to new and/or improved transformation and regulation techniques.

15 Of particular interest, is the use of plant LCAT and ACAT constructs in plants to produce plants or plant parts, including, but not limited to leaves, stems, roots, reproductive, and seed, with a modified content of lipid and/or sterol esters and to alter the oil production by such plants.

Of particular interest in the present invention, is the use of ACAT genes in  
20 conjunction with the LCAT sequences to increase the sterol content of seeds. Thus, overexpression of a nucleic acid sequence encoding an ACAT and LCAT in an oilseed crop may find use in the present invention to increase sterol levels in plant tissues and/or increase oil production.

It is contemplated that the gene sequences may be synthesized, either completely or  
25 in part, especially where it is desirable to provide plant-preferred sequences. Thus, all or a portion of the desired structural gene (that portion of the gene which encodes the LCAT or ACAT protein) may be synthesized using codons preferred by a selected host. Host-preferred codons may be determined, for example, from the codons used most frequently in the proteins expressed in a desired host species.

30 One skilled in the art will readily recognize that antibody preparations, nucleic acid probes (DNA and RNA) and the like may be prepared and used to screen and recover "homologous" or "related" sequences from a variety of plant sources. Homologous sequences are found when there is an identity of sequence, which may be determined upon comparison of sequence information, nucleic acid or amino acid, or through hybridization

reactions between a known LCAT and a candidate source. Conservative changes, such as Glu/Asp, Val/Ile, Ser/Thr, Arg/Lys and Gln/Asn may also be considered in determining sequence homology. Amino acid sequences are considered homologous by as little as 25% sequence identity between the two complete mature proteins. (See generally, 5 Doolittle, R.F., *OF URFS and ORFS* (University Science Books, CA, 1986.)

Thus, other LCATs may be obtained from the specific sequences provided herein. Furthermore, it will be apparent that one can obtain natural and synthetic sequences, including modified amino acid sequences and starting materials for synthetic-protein modeling from the exemplified LCAT and ACAT sequences and from sequences which 10 are obtained through the use of such exemplified sequences. Modified amino acid sequences include sequences which have been mutated, truncated, increased and the like, whether such sequences were partially or wholly synthesized. Sequences which are actually purified from plant preparations or are identical or encode identical proteins thereto, regardless of the method used to obtain the protein or sequence, are equally 15 considered naturally derived.

For immunological screening, antibodies to the protein can be prepared by injecting rabbits or mice with the purified protein or portion thereof, such methods of preparing antibodies being well known to those in the art. Either monoclonal or polyclonal antibodies can be produced, although typically polyclonal antibodies are more 20 useful for gene isolation. Western analysis may be conducted to determine that a related protein is present in a crude extract of the desired plant species, as determined by cross-reaction with the antibodies to the encoded proteins. When cross-reactivity is observed, genes encoding the related proteins are isolated by screening expression libraries representing the desired plant species. Expression libraries can be constructed in a variety 25 of commercially available vectors, including lambda gt11, as described in Sambrook, *et al.* (*Molecular Cloning: A Laboratory Manual*, Second Edition (1989) Cold Spring Harbor Laboratory, Cold Spring Harbor, New York).

To confirm the activity and specificity of the proteins encoded by the identified nucleic acid sequences as acyltransferase enzymes, *in vitro* assays are performed in insect 30 cell cultures using baculovirus expression systems. Such baculovirus expression systems are known in the art and are described by Lee, *et al.* U.S. Patent Number 5,348,886, the entirety of which is herein incorporated by reference.

In addition, other expression constructs may be prepared to assay for protein activity utilizing different expression systems. Such expression constructs are transformed

into yeast or prokaryotic host and assayed for acyltransferase activity. Such expression systems are known in the art and are readily available through commercial sources.

The method of transformation in obtaining such transgenic plants is not critical to the instant invention, and various methods of plant transformation are currently available.

5 Furthermore, as newer methods become available to transform crops, they may also be directly applied hereunder. For example, many plant species naturally susceptible to *Agrobacterium* infection may be successfully transformed via tripartite or binary vector methods of *Agrobacterium* mediated transformation. In many instances, it will be desirable to have the construct bordered on one or both sides by T-DNA, particularly  
10 having the left and right borders, more particularly the right border. This is particularly useful when the construct uses *A. tumefaciens* or *A. rhizogenes* as a mode for transformation, although the T-DNA borders may find use with other modes of transformation. In addition, techniques of microinjection, DNA particle bombardment, and electroporation have been developed which allow for the transformation of various  
15 monocot and dicot plant species.

Normally, included with the DNA construct will be a structural gene having the necessary regulatory regions for expression in a host and providing for selection of transformant cells. The gene may provide for resistance to a cytotoxic agent, e.g. antibiotic, heavy metal, toxin, etc., complementation providing prototrophy to an  
20 auxotrophic host, viral immunity or the like. Depending upon the number of different host species the expression construct or components thereof are introduced, one or more markers may be employed, where different conditions for selection are used for the different hosts.

Non-limiting examples of suitable selection markers include genes that confer  
25 resistance to bleomycin, gentamycin, glyphosate, hygromycin, kanamycin, methotrexate, phleomycin, phosphinotricin, spectinomycin, streptomycin, sulfonamide and sulfonylureas. Maliga et al., *Methods in Plant Molecular Biology*, Cold Spring Harbor Laboratory Press, 1995, p. 39. Examples of markers include, but are not limited to, alkaline phosphatase (AP), myc, hemagglutinin (HA),  $\beta$  glucuronidase (GUS), luciferase,  
30 and green fluorescent protein (GFP).

Where *Agrobacterium* is used for plant cell transformation, a vector may be used which may be introduced into the *Agrobacterium* host for homologous recombination with T-DNA or the Ti- or Ri-plasmid present in the *Agrobacterium* host. The Ti- or Ri-plasmid containing the T-DNA for recombination may be armed (capable of causing gall



formation) or disarmed (incapable of causing gall formation), the latter being permissible, so long as the *vir* genes are present in the transformed *Agrobacterium* host. The armed plasmid can give a mixture of normal plant cells and gall.

In some instances where *Agrobacterium* is used as the vehicle for transforming  
5 host plant cells, the expression or transcription construct bordered by the T-DNA border region(s) will be inserted into a broad host range vector capable of replication in *E. coli* and *Agrobacterium*, there being broad host range vectors described in the literature. Commonly used is pRK2 or derivatives thereof. See, for example, Ditta, *et al.*, (*Proc. Nat. Acad. Sci., U.S.A.* (1980) 77:7347-7351) and EPA 0 120 515, which are incorporated  
10 herein by reference. Alternatively, one may insert the sequences to be expressed in plant cells into a vector containing separate replication sequences, one of which stabilizes the vector in *E. coli*, and the other in *Agrobacterium*. See, for example, McBride and Summerfelt (*Plant Mol. Biol.* (1990) 14:269-276), wherein the pRiHRI (Jouanin, *et al.*, *Mol. Gen. Genet.* (1985) 201:370-374) origin of replication is utilized and provides for  
15 added stability of the plant expression vectors in host *Agrobacterium* cells.

Included with the expression construct and the T-DNA can be one or more markers, which allow for selection of transformed *Agrobacterium* and transformed plant cells. A number of markers have been developed for use with plant cells, such as resistance to  
20 chloramphenicol, kanamycin, the aminoglycoside G418, hygromycin, or the like. The particular marker employed is not essential to this invention, one or another marker being preferred depending on the particular host and the manner of construction.

For transformation of plant cells using *Agrobacterium*, explants may be combined and incubated with the transformed *Agrobacterium* for sufficient time for transformation,  
25 the bacteria killed, and the plant cells cultured in an appropriate selective medium. Once callus forms, shoot formation can be encouraged by employing the appropriate plant hormones in accordance with known methods and the shoots transferred to rooting medium for regeneration of plants. The plants may then be grown to seed and the seed used to establish repetitive generations and for isolation of vegetable oils.

30 Thus, in another aspect of the present invention, methods for modifying the sterol and/or stanol composition of a host cell. Of particular interest are methods for modifying the sterol and/or stanol composition of a host plant cell. In general the methods involve either increasing the levels of sterol ester compounds as a proportion of the total sterol

compounds. The method generally comprises the use of expression constructs to direct the expression of the polynucleotides of the present invention in a host cell.

Also provided are methods for reducing the proportion of sterol ester compounds as a percentage of total sterol compounds in a host plant cell. The method generally  
5 comprises the use of expression constructs to direct the suppression of endogenous acyltransferase proteins in a host cell.

Of particular interest is the use of expression constructs to modify the levels of sterol compounds in a host plant cell. Most particular, the methods find use in modifying the levels of sterol compounds in seed oils obtained from plant seeds.

10 Also of interest is the use of expression constructs of the present invention to alter oil production in a host cell and in particular to increase oil production. Of particular interest is the use of expression constructs containing nucleic acid sequences encoding LCAT and/or ACAT polypeptides to transform host plant cells and to use these host cells to regenerate whole plants having increase oil production as compared to the same plant  
15 not containing the expression construct.

The oils obtained from transgenic plants having modified sterol compound content find use in a wide variety of applications. Of particular interest in the present invention is the use of the oils containing modified levels of sterol compounds in applications involved in improving human nutrition and cardiovascular health. For example, phytosterols are  
20 beneficial for lowering serum cholesterol (Ling, *et al.* (1995) *Life Sciences* 57:195-206).

Cholesterol-lowering compositions comprise the oils and sterol ester compound compositions obtained using the methods of the present invention. Such cholesterol lowering compositions include, but are not limited to foods, food products, processed foods, food ingredients, food additive compositions, or dietary/nutritional supplements  
25 that contain oils and/or fats. Non-limiting examples include margarines; butters; shortenings; cooking oils; frying oils; dressings, such as salad dressings; spreads; mayonnaises; and vitamin/mineral supplements. Patent documents relating to such compositions include, U.S. Patents 4,588,717 and 5,244,887, and PCT International Publication Nos. WO 96/38047, WO 97/42830, WO 98/06405, and WO 98/06714.  
30 Additional non-limiting examples include toppings; dairy products such as cheese and processed cheese; processed meat; pastas; sauces; cereals; desserts, including frozen and shelf-stable desserts; dips; chips; baked goods; pastries; cookies; snack bars; confections; chocolates; beverages; unextracted seed; and unextracted seed that has been

ground, cracked, milled, rolled, extruded, pelleted, defatted, dehydrated, or otherwise processed, but which still contains the oils, etc., disclosed herein.

The cholesterol-lowering compositions can also take the form of pharmaceutical compositions comprising a cholesterol-lowering effective amount of the oils or sterol compound compositions obtained using the methods of the present invention, along with a pharmaceutically acceptable carrier, excipient, or diluent. These pharmaceutical compositions can be in the form of a liquid or a solid. Liquids can be solutions or suspensions; solids can be in the form of a powder, a granule, a pill, a tablet, a gel, or an extrudate. U.S. Patent 5,270,041 relates to sterol-containing pharmaceutical compositions.

Thus, by expression of the nucleic acid sequences encoding acyltransferase-like sequences of the present invention in a host cell, it is possible to modify the lipid content and/or composition as well as the sterol content and/or composition of the host cell.

The invention now being generally described, it will be more readily understood by reference to the following examples which are included for purposes of illustration only and are not intended to limit the present invention.

## EXAMPLES

### Example 1: RNA Isolations

Total RNA from the inflorescence and developing seeds of *Arabidopsis thaliana* was isolated for use in construction of complementary (cDNA) libraries. The procedure was an adaptation of the DNA isolation protocol of Webb and Knapp (D.M. Webb and S.J. Knapp, (1990) Plant Molec. Reporter, 8, 180-185). The following description assumes the use of 1g fresh weight of tissue. Frozen seed tissue was powdered by grinding under liquid nitrogen. The powder was added to 10ml REC buffer (50mM Tris-HCl, pH 9, 0.8M NaCl, 10mM EDTA, 0.5% w/v CTAB (cetyltrimethyl-ammonium bromide)) along with 0.2g insoluble polyvinylpolypyrrolidone, and ground at room temperature. The homogenate was centrifuged for 5 minutes at 12,000 xg to pellet insoluble material. The resulting supernatant fraction was extracted with chloroform, and the top phase was recovered.

The RNA was then precipitated by addition of 1 volume RecP (50mM Tris-HCL pH9, 10mM EDTA and 0.5% (w/v) CTAB) and collected by brief centrifugation as before. The RNA pellet was redissolved in 0.4 ml of 1M NaCl. The RNA pellet was redissolved in water and extracted with phenol/chloroform. Sufficient 3M potassium acetate (pH 5) was added to make the mixture 0.3M in acetate, followed by addition of two volumes of

ethanol to precipitate the RNA. After washing with ethanol, this final RNA precipitate was dissolved in water and stored frozen.

Alternatively, total RNA may be obtained using TRIzol reagent (BRL-Lifetechnologies, Gaithersburg, MD) following the manufacturer's protocol. The RNA  
5 precipitate was dissolved in water and stored frozen.

### Example 2: Identification of LCAT Sequences

Searches were performed on a Silicon Graphics Unix computer using additional Bioaccelerator hardware and GenWeb software supplied by Compugen Ltd. This  
10 software and hardware enabled the use of the Smith-Waterman algorithm in searching DNA and protein databases using profiles as queries. The program used to query protein databases was profilesearch. This is a search where the query is not a single sequence but a profile based on a multiple alignment of amino acid or nucleic acid sequences. The profile was used to query a sequence data set, i.e., a sequence database. The profile  
15 contained all the pertinent information for scoring each position in a sequence, in effect replacing the "scoring matrix" used for the standard query searches. The program used to query nucleotide databases with a protein profile was tprofilesearch. Tprofilesearch searches nucleic acid databases using an amino acid profile query. As the search is running, sequences in the database are translated to amino acid sequences in six reading  
20 frames. The output file for tprofilesearch is identical to the output file for profilesearch except for an additional column that indicates the frame in which the best alignment occurred.

The Smith-Waterman algorithm, (Smith and Waterman (1981) *J. Molec. Biol.* 147:195-197), was used to search for similarities between one sequence from the query  
25 and a group of sequences contained in the database.

A protein sequence of Lecithin: cholesterol acyltransferase from human (McLean J, et al. (1986) *Nucleic Acids Res.* 14(23):9397-406 SEQ ID NO:1)) was used to search the NCBI non-redundant protein database using BLAST. Three sequences were identified from *Arabidopsis*, GenBank accessions AC004557 (referred to herein as LCAT1, SEQ ID  
30 NO:2), AC003027 (referred to herein as LCAT2, SEQ ID NO:4), and AL024486 (referred to herein as LCAT3, SEQ ID NO:6). The deduced amino acid sequences are provided in SEQ ID NOs: 3, 5, and 7, respectively.

The profile generated from the queries using PSI-BLAST was excised from the hyper text markup language (html) file. The worldwide web (www)/html interface to

psiblast at ncbi stores the current generated profile matrix in a hidden field in the html file that is returned after each iteration of psiblast. However, this matrix has been encoded into string62 (s62) format for ease of transport through html. String62 format is a simple conversion of the values of the matrix into html legal ascii characters.

- 5           The encoded matrix width (x axis) is 26 characters, and comprise the consensus characters, the probabilities of each amino acid in the order A,B,C,D,E,F,G,H,I,K,L,M,N, P,Q,R,S,T,V,W,X,Y,Z (where B represents D and N, and Z represents Q and E, and X represents any amino acid), gap creation value, and gap extension value.

- 10           The length (y axis) of the matrix corresponds to the length of the sequences identified by PSI-BLAST. The order of the amino acids corresponds to the conserved amino acid sequence of the sequences identified using PSI-BLAST, with the N-terminal end at the top of the matrix. The probabilities of other amino acids at that position are represented for each amino acid along the x axis, below the respective single letter amino acid abbreviation.

- 15           Thus, each row of the profile consists of the highest scoring (consensus) amino acid, followed by the scores for each possible amino acid at that position in sequence matrix, the score for opening a gap at that position, and the score for continuing a gap at that position.

- 20           The string62 file is converted back into a profile for use in subsequent searches. The gap open field is set to 11 and the gap extension field is set to 1 along the x axis. The gap creation and gap extension values are known, based on the settings given to the PSI-BLAST algorithm. The matrix is exported to the standard GCG profile form. This format can be read by GenWeb.

- 25           The algorithm used to convert the string62 formatted file to the matrix is outlined in Table 1.

Table 1

1. if encoded character z then the value is blast score min
2. if encoded character Z then the value is blast score max
- 5 3. else if the encoded character is uppercase then its value is (64-(ascii # of char))
4. else if the encoded character is a digit the value is ((ascii # of char)-48)
5. else if the encoded character is not uppercase then the value is ((ascii # of char) - 87)
6. ALL B positions are set to min of D and N amino acids at that row in sequence matrix
7. ALL Z positions are set to min of Q and E amino acids at that row in sequence matrix
- 10 8. ALL X positions are set to min of all amino acids at that row in sequence matrix
9. kBLAST\_SCORE\_MAX=999;
10. kBLAST\_SCORE\_MIN=-999;
11. all gap opens are set to 11
12. all gap lens are set to 1

15

The protein sequences of LCAT1, LCAT2, and LCAT3 as well as the PSI-BLAST profile were used to search public and proprietary databases for additional LCAT sequences. Two EST sequences were identified which appear to be identical to LCAT1 and LCAT3, respectively. One additional *Arabidopsis* sequence was identified from the proprietary databases, LCAT4 (SEQ ID NO:8). The deduced protein sequence of LCAT4 is provided in SEQ ID NO:9. Two additional genomic sequences were identified using the PSI-BLAST profile from libraries of *Arabidopsis* ecotypes Columbia and Landsberg, LCAT7 (SEQ ID NO:10) and LCAT8 (SEQ ID NO:11). The LCAT7 sequence was present in both the Columbia and Landsberg genomic libraries, while the LCAT8 sequence was only present in the Columbia library.

25

An open reading frame was predicted from the genomic sequence of LCAT7 in the *Arabidopsis* public database and this sequence was called MSH12 (referred to herein as LCAT5, SEQ ID NO: 73). The deduced protein sequence of LCAT5 is provided in SEQ ID NO: 74.

30

The PSI-BLAST profile and the LCAT sequences were used to query the public yeast database and proprietary libraries containing corn and soy EST sequences. The yeast genome contains only one gene, *LRO1* (LCAT Related Open reading frame, YNR008W, Figure 1) with distinct similarity to the human LCAT. The DNA sequence of *LRO1* is

provided in SEQ ID NO: 75 and the protein sequence is provided in SEQ ID NO: 76. Seven EST sequences were identified from soybean libraries as being LCAT sequences. Two sequences from soy (SEQ ID NOS: 12 and 13) are most closely related to the *Arabidopsis* LCAT1 sequence, a single sequence was identified as being most closely  
5 related to LCAT2 (SEQ ID NO:14), three were closely related to LCAT3 (SEQ ID NOS: 15-17), and an additional single sequence was identified (SEQ ID NO:18). A total of 11 corn EST sequences were identified as being related to the *Arabidopsis* LCAT sequences. Two corn EST sequences (SEQ ID NOS: 19 and 20) were most closely related to LCAT1, two sequences were identified as closely related to LCAT2 (SEQ ID NOS: 21 and 22), four  
10 corn EST sequences were identified as closely related to LCAT3 (SEQ ID NOS: 23-26), and an additional three corn EST sequences were also identified (SEQ ID NOS: 27-29).

### Example 3: Identification of ACAT Sequences

Since plant ACATs are unknown in the art, searches were performed to identify  
15 known and related ACAT sequences from mammalian sources from public databases. These sequences were then used to search public and proprietary EST databases to identify plant ACAT-like sequences.

A public database containing mouse Expressed Sequence Tag (EST) sequences (dBEST) was searched for ACAT-like sequences. The search identified two sequences  
20 (SEQ ID 30 and 31) which were related (approximately 20% identical), but divergent, to known ACAT sequences.

In order to identify ACAT-like sequences from other organisms, the two mouse ACAT sequences were used to search public and proprietary databases containing EST sequences from human and rat tissues. Results of the search identified several sequences  
25 from the human database and from the rat database which were closely related to the mouse sequences. The human and rat ACAT-like EST sequences were assembled, using the GCG assembly program, to construct a complete inferred cDNA sequence by identifying overlapping sequences (SEQ ID NOS: 32 and 33, respectively).

The protein sequence of the human ACAT-like sequence was aligned with known  
30 ACAT sequences from human (Chang, *et al.* (1993) *J. Biol. Chem.* 268:20747-20755, SEQ ID NO:34), mouse (Uelmen, *et al.* (1995) *J. Biol. Chem.* 270:26192-26201 SEQ ID NO:35) and yeast (Yu, *et al.* (1996) *J. Biol. Chem.* 271:24157-24163, SEQ ID NO:36 and Yang, *et al.* (1996) *Science* 272:1353-1356, SEQ ID NO:37) using MacVector (Oxford Molecular, Inc.). Results of the alignment demonstrated that the sequence was related to

the known sequences, however the related sequence was only about 25% similar to the known sequences.

The protein sequence of the human sterol O-acyltransferase (ACAT, Acyl CoA:Cholesterol acyltransferase, Accession number A48026) related sequence was used  
5 to search protein and nucleic acid Genbank databases. A single plant homologue was identified in the public *Arabidopsis* EST database (Accession A042298, SEQ ID NO:38). The protein sequence (SEQ ID NO:39) was translated from the EST sequence, and was found to contain a peptide sequence conserved in both mammalian and yeast ACATs (Chang *et al.*, (1997) *Ann. Rev. Biochem.*, 66:613-638).

10 To obtain the entire coding region corresponding to the *Arabidopsis* ACAT-like EST, synthetic oligo-nucleotide primers were designed to amplify the 5' and 3' ends of partial cDNA clones containing ACAT-like sequences. Primers were designed according to the *Arabidopsis* ACAT-like EST sequence and were used in Rapid Amplification of cDNA Ends (RACE) reactions (Frohman *et al.* (1988) *Proc. Natl. Acad. Sci. USA*  
15 85:8998-9002).

Primers were designed (5'-TGCAAATTGACGAGCACACCAACCCCTTC-3' (SEQ ID NO:40) and 5'-AAGGATGCTTTGAGTTCCTGACAATAGG-3' (SEQ ID NO:41)) to amplify the 5' end from the *Arabidopsis* ACAT EST sequence. Amplification of flanking sequences from cDNA clones were performed using the Marathon cDNA  
20 Amplification kit (Clontech, CA).

The sequence derived from the 5'-RACE amplification was used to search proprietary *Arabidopsis* EST libraries. A single EST accession, LIB25-088-C7 (SEQ ID NO:42), was identified which contained a sequence identical to the 5'-RACE sequence. Furthermore, LIB25-088-C7 was found to contain the complete putative coding sequence  
25 for the *Arabidopsis* ACAT-like product.

The nucleic acid as well as the putative translation product sequences of A042298 were used to search public and proprietary databases. Four EST sequences were identified in both soybean (SEQ ID NOs:43-46) and maize (SEQ ID NOs:47-50) proprietary  
30 databases, and a single ACAT-like sequence was identified from *Mortierrella alpina* EST sequences (SEQ ID NO:51).

Sequence alignments between ACAT sequences from several different sources were compared to identify the similarity between the sequences. Nucleotide sequences from known human and mouse ACATs, as well as nucleotide sequences from known yeast ACATs were compared to the ACAT-like EST sequences from human and *Arabidopsis*.



Analysis of the sequence alignments revealed several classes of ACATs based on sequence similarity. The known human and mouse ACATs, being 88% similar in the nucleotide sequence, formed one class of ACATs. Another class of ACATs included the yeast ACATs which are less than 20% similar to the known human and mouse class

5 ACATs.

The final class of ACATs included the Arabidopsis and human sequences disclosed in the present invention. This class is approximately 22% similar to the known human and mouse ACAT class and approximately 23% similar to the yeast class of ACATs. Thus, the ACAT sequences disclosed in the present invention represent a novel class of ACAT  
10 enzymes. Partial mouse sequences of this class are also provided.

#### Example 4: Expression Construct Preparation

Constructs were prepared to direct expression of the LCAT1, LCAT2, LCAT3, LCAT4, LCAT5 and the yeast LRO1 sequences in plants and cultured insect cells. The  
15 entire coding region of each LCAT was amplified from the appropriate EST clone or an Arabidopsis genomic cDNA library using the following oligonucleotide primers in a polymerase chain reactions (PCR). The LCAT1 coding sequence was amplified from the EST clone Lib25-082-Q1-E1-G4 using the primers  
5'-GGATCCGCGGCCCGCACAATGAAAAAATATCTTCACATTATTTCGG-3' (SEQ  
20 ID NO:52) and 5'-GGATCCCCTGCAGGTCATTTCATTGACGGCATTAAACATTGG-3'  
(SEQ ID NO:53). The LCAT2 coding sequence was amplified from an Arabidopsis genomic cDNA library using the synthetic oligo nucleotide primers  
5'-GGATCCGCGGCCCGCACAATGGGAGCGAATTCGAAATCAGTAACG-3' (SEQ  
ID NO:54) and 5'-GGATCCCCTGCAGGTTAATACCCACTTTTATCAAGCTCCC-3'  
25 (SEQ ID NO:55). The LCAT3 coding sequence was amplified from the EST clone LIB22-004-Q1-E1-B4 using the synthetic oligo nucleotide primers  
5'-GGATCCGCGGCCCGCACAATGTCTCTATTACTGGAA GAGATC-3' (SEQ ID  
NO:56) and 5'-GGATCCCCTGCAGGTTATGCATC AACAGAGACACTTACAGC-3'  
(SEQ ID NO:57). The LCAT4 coding sequence was amplified from the EST clone  
30 LIB23-007-Q1-E1-B5 using the synthetic oligo nucleotide primers  
5'-GGATCCGCGGCCCGCACAATGGGCTGGATTCCGTGTCCGTGC-3' (SEQ ID  
NO:58) and 5'-GGATCCCCTGCAGGTTAACCAGAATCAACTACTTTGTG-3' (SEQ  
ID NO:59). The LCAT5 coding sequence was amplified from LIB23-053-Q1-E1-E3 using  
the synthetic oligo nucleotide primers

5'-GGATCCGCGGCCGCGCACAATGCCCCTTATTCATCGG-3' (SEQ ID NO:77) and 5'-GGATCCCCTGCAGGTCACAGCTTCAGGTCAATACG-3' (SEQ ID NO:78).

The yeast LROI coding sequence was amplified from genomic yeast DNA using the synthetic oligo nucleotide primers

- 5 5'GGATCCGCGGCCGCGCACAATGGGCACACTGTTTCGAAG3' (SEQ ID NO:79)  
and 5'GGATCCCCTGCAGGTTACATTGGGAAGGGCATCTGAG3' (SEQ ID NO:80).

The entire coding region of the *Arabidopsis* ACAT sequence (SEQ ID NO: 42) was amplified from the EST clone LIB25-088-C7 using oligonucleotide primers

- 5'-TCGACCTGCAGGAAGCTTAGAAATGGCGATTTTGGATTC-3' (SEQ ID NO: 60)  
10 and 5'-GGATCCGCGGCCGCTCATGACATCGATCCTTTTCGG-3' (SEQ ID NO: 61)  
in a polymerase chain reaction (PCR).

Each resulting PCR product was subcloned into pCR2.1Topo (Invitrogen) and labeled pCGN9964 (LCAT1), pCGN9985 (LCAT2), pCGN9965 (LCAT3), pCGN9995 (LCAT4), pCGN10964 (LCAT5), pCGN10963 (*LROI*), and pCGN8626 (ACAT).

- 15 Double stranded DNA sequence was obtained to verify that no errors were introduced by the PCR amplification.

#### 4A. Baculovirus Expression Constructs

- Constructs are prepared to direct the expression of the *Arabidopsis* LCAT and  
20 yeast LCAT sequences in cultured insect cells. The entire coding region of the LCAT proteins was removed from the respective constructs by digestion with *NotI* and *Sse8387I*, followed by gel electrophoresis and gel purification. The fragments containing the LCAT coding sequences were cloned into *NotI* and *PstI* digested baculovirus expression vector pFastBac1 (Gibco-BRL, Gaithersburg, MD). The resulting baculovirus expression  
25 constructs were referred to as pCGN9992 (LCAT1), pCGN9993 (LCAT2), pCGN9994 (LCAT3), pCGN10900 (LCAT4), pCGN10967 (LCAT5), and pCGN10962 (*LROI*).

#### 4B. Plant Expression Construct Preparation

- A plasmid containing the napin cassette derived from pCGN3223 (described in  
30 U.S. Patent No. 5,639,790, the entirety of which is incorporated herein by reference) was modified to make it more useful for cloning large DNA fragments containing multiple restriction sites, and to allow the cloning of multiple napin fusion genes into plant binary transformation vectors. An adapter comprised of the self annealed oligonucleotide of sequence 5'-

CGCGATTTAAATGGCGCGCCCTGCAGGCGGCCGCCTGCAGGGCGCGCCATTTA  
AAT-3' (SEQ ID NO:62) was ligated into the cloning vector pBC SK+ (Stratagene) after  
digestion with the restriction endonuclease BssHII to construct vector pCGN7765.

Plamids pCGN3223 and pCGN7765 were digested with NotI and ligated together. The  
5 resultant vector, pCGN7770, contained the pCGN7765 backbone with the napin seed  
specific expression cassette from pCGN3223.

The cloning cassette, pCGN7787, contained essentially the same regulatory  
elements as pCGN7770, with the exception of the napin regulatory regions of pCGN7770  
have been replaced with the double CAMV 35S promoter and the tml polyadenylation and  
10 transcriptional termination region.

A binary vector for plant transformation, pCGN5139, was constructed from  
pCGN1558 (McBride and Summerfelt, (1990) Plant Molecular Biology, 14:269-276). In  
pCGN5139, the polylinker of pCGN1558 was replaced as a HindIII/Asp718 fragment with  
a polylinker containing unique restriction endonuclease sites, AscI, PacI, XbaI, SmaI,  
15 BamHI, and NotI. The Asp718 and HindIII restriction endonuclease sites are retained in  
pCGN5139.

A series of turbo binary vectors was constructed to allow for the rapid cloning of  
DNA sequences into binary vectors containing transcriptional initiation regions  
(promoters) and transcriptional termination regions.

20 The plasmid pCGN8618 was constructed by ligating oligonucleotides  
5'-TCGAGGATCCGCGGCCGCAAGCTTCCTGCAGG-3' (SEQ ID NO:63) and  
5'-TCGACCTGCAGGAAGCTTGCGGCCGCGGATCC-3' (SEQ ID NO:64) into  
SalI/XhoI-digested pCGN7770. A fragment containing the napin promoter, polylinker and  
napin 3' region was excised from pCGN8618 by digestion with Asp718I; the fragment  
25 was blunt-ended by filling in the 5' overhangs with Klenow fragment then ligated into  
pCGN5139 that had been digested with Asp718I and HindIII and blunt-ended by filling in  
the 5' overhangs with Klenow fragment. A plasmid containing the insert oriented so that  
the napin promoter was closest to the blunted Asp718I site of pCGN5139 and the napin 3'  
was closest to the blunted HindIII site was subjected to sequence analysis to confirm both  
30 the insert orientation and the integrity of cloning junctions. The resulting plasmid was  
designated pCGN8622.

The plasmid pCGN8619 was constructed by ligating oligonucleotides  
5'-TCGACCTGCAGGAAGCTTGCGGCCGCGGATCC-3' (SEQ ID NO:65) and

5'-TCGAGGATCCGCGGCCGCAAGCTTCCTGCAGG-3' (SEQ ID NO:66) into SalI/XhoI-digested pCGN7770. A fragment containing the napin promoter, polylinker and napin 3' region was removed from pCGN8619 by digestion with Asp718I; the fragment was blunt-ended by filling in the 5' overhangs with Klenow fragment then ligated into pCGN5139 that had been digested with Asp718I and HindIII and blunt-ended by filling in the 5' overhangs with Klenow fragment. A plasmid containing the insert oriented so that the napin promoter was closest to the blunted Asp718I site of pCGN5139 and the napin 3' was closest to the blunted HindIII site was subjected to sequence analysis to confirm both the insert orientation and the integrity of cloning junctions. The resulting plasmid was designated pCGN8623.

The plasmid pCGN8620 was constructed by ligating oligonucleotides 5'-TCGAGGATCCGCGGCCGCAAGCTTCCTGCAGGAGCT -3' (SEQ ID NO:67) and 5'-CCTGCAGGAAGCTTGCGGCCGCGGATCC-3' (SEQ ID NO:68) into SalI/SacI-digested pCGN7787. A fragment containing the d35S promoter, polylinker and tml 3' region was removed from pCGN8620 by complete digestion with Asp718I and partial digestion with NotI. The fragment was blunt-ended by filling in the 5' overhangs with Klenow fragment then ligated into pCGN5139 that had been digested with Asp718I and HindIII and blunt-ended by filling in the 5' overhangs with Klenow fragment. A plasmid containing the insert oriented so that the d35S promoter was closest to the blunted Asp718I site of pCGN5139 and the tml 3' was closest to the blunted HindIII site was subjected to sequence analysis to confirm both the insert orientation and the integrity of cloning junctions. The resulting plasmid was designated pCGN8624.

The plasmid pCGN8621 was constructed by ligating oligonucleotides 5'-TCGACCTGCAGGAAGCTTGCGGCCGCGGATCCAGCT -3' (SEQ ID NO:69) and 5'-GGATCCGCGGCCGCAAGCTTCCTGCAGG-3' (SEQ ID NO:70) into SalI/SacI-digested pCGN7787. A fragment containing the d35S promoter, polylinker and tml 3' region was removed from pCGN8621 by complete digestion with Asp718I and partial digestion with NotI. The fragment was blunt-ended by filling in the 5' overhangs with Klenow fragment then ligated into pCGN5139 that had been digested with Asp718I and HindIII and blunt-ended by filling in the 5' overhangs with Klenow fragment. A plasmid containing the insert oriented so that the d35S promoter was closest to the blunted Asp718I site of pCGN5139 and the tml 3' was closest to the blunted HindIII site was subjected to sequence analysis to confirm both the insert orientation and the integrity of cloning junctions. The resulting plasmid was designated pCGN8625.

The plasmid construct pCGN8640 is a modification of pCGN8624 described above. A 938bp PstI fragment isolated from transposon Tn7 which encodes bacterial spectinomycin and streptomycin resistance (Fling et al. (1985), *Nucleic Acids Research* 13(19):7095-7106), a determinant for *E. coli* and *Agrobacterium* selection, was blunt ended with Pfu polymerase. The blunt ended fragment was ligated into pCGN8624 that had been digested with SpeI and blunt ended with Pfu polymerase. The region containing the PstI fragment was sequenced to confirm both the insert orientation and the integrity of cloning junctions.

The spectinomycin resistance marker was introduced into pCGN8622 and pCGN8623 as follows. A 7.7 Kbp AvrII-SnaBI fragment from pCGN8640 was ligated to a 10.9 Kbp AvrII-SnaBI fragment from pCGN8623 or pCGN8622, described above. The resulting plasmids were pCGN8641 and pCGN8643, respectively.

The plasmid pCGN8644 was constructed by ligating oligonucleotides 5'-GATCACCTGCAGGAAGCTTGCGGCCGCGGATCCAATGCA-3' (SEQ ID NO:71) and 5'-TTGGATCCGCGGCCGCAAGCTTCCTGCAGGT-3' (SEQ ID NO:72) into BamHI-PstI digested pCGN8640.

#### 4C. Plant LCAT Expression Construct Preparation

The coding sequence of LCAT1 was cloned from pCGN9964 as a *NotI*/ *Sse8387I* fragment into pCGN8640, pCGN8641, pCGN8643, and pCGN8644 to create the expression constructs pCGN9960, pCGN9961, pCGN9962, and pCGN9963, respectively. The construct pCGN9960 was designed to express the LCAT1 coding sequence in the sense orientation from the constitutive promoter CaMV 35S. The construct pCGN9961 was designed to express the LCAT1 coding sequence in the antisense orientation from the napin promoter. The construct pCGN9962 was designed to express the LCAT1 coding sequence in the sense orientation from the napin promoter. The construct pCGN9963 was designed to express the LCAT1 coding sequence in the antisense orientation from the constitutive promoter CaMV 35S.

The coding sequence of LCAT2 was cloned from pCGN9985 as a *NotI*/ *Sse8387I* fragment into pCGN8640, pCGN8641, pCGN8643, and pCGN8644 to create the expression constructs pCGN9981, pCGN9982, pCGN9983, and pCGN9984, respectively. The construct pCGN9981 was designed to express the LCAT2 coding sequence in the sense orientation from the constitutive promoter CaMV 35S. The construct pCGN9982 was designed to express the LCAT2 coding sequence in the antisense orientation from the

napin promoter. The construct pCGN9983 was designed to express the LCAT2 coding sequence in the sense orientation from the napin promoter. The construct pCGN9984 was designed to express the LCAT2 coding sequence in the antisense orientation from the constitutive promoter CaMV 35S.

5           The coding sequence of LCAT3 was cloned from pCGN9965 as a *NotI*/*Sse8387I* fragment into pCGN8640, pCGN8641, pCGN8643, and pCGN8644 to create the expression constructs pCGN9966, pCGN9967, pCGN9968, and pCGN9969, respectively. The construct pCGN9966 was designed to express the LCAT3 coding sequence in the sense orientation from the constitutive promoter CaMV 35S. The construct pCGN9967  
10       was designed to express the LCAT3 coding sequence in the antisense orientation from the napin promoter. The construct pCGN9968 was designed to express the LCAT3 coding sequence in the sense orientation from the napin promoter. The construct pCGN9969 was designed to express the LCAT3 coding sequence in the antisense orientation from the constitutive promoter CaMV 35S.

15           The coding sequence of LCAT4 was cloned from pCGN9995 as a *NotI*/*Sse8387I* fragment into pCGN8640, pCGN8641, pCGN8643, and pCGN8644 to create the expression constructs pCGN9996, pCGN9997, pCGN9998, and pCGN9999, respectively. The construct pCGN9996 was designed to express the LCAT4 coding sequence in the sense orientation from the constitutive promoter CaMV 35S. The construct pCGN9997  
20       was designed to express the LCAT4 coding sequence in the antisense orientation from the napin promoter. The construct pCGN9998 was designed to express the LCAT4 coding sequence in the sense orientation from the napin promoter. The construct pCGN9999 was designed to express the LCAT4 coding sequence in the antisense orientation from the constitutive promoter CaMV 35S.

25           The coding sequence of LCAT5 was cloned from pCGN10964 as a *NotI*/*Sse8387I* fragment into pCGN9977 and pCGN9979, to create the expression constructs pCGN10965, and pCGN10966, respectively. The construct pCGN10965 was designed to express the LCAT5 coding sequence in the sense orientation from the constitutive promoter CaMV 35S. The construct pCGN10966 was designed to express the LCAT5  
30       coding sequence in the sense orientation from the napin promoter.

          The coding sequence of *LROI* was cloned from pCGN10963 as a *NotI*/*Sse8387I* fragment into pCGN9977 and pCGN9979, to create the expression constructs pCGN10960, and pCGN10961, respectively. The construct pCGN10960 was designed to express the *LROI* coding sequence in the sense orientation from the constitutive promoter

CaMV 35S. The construct pCGN10961 was designed to express the *LRO1* coding sequence in the sense orientation from the napin promoter.

#### 4D. Plant ACAT Expression Construct Preparation

5 A fragment containing the *Arabidopsis* ACAT-like coding region was removed from pCGN8626 by digestion with Sse8387I and Not I. The fragment containing the ACAT-like sequence was ligated into PstI-Not I digested pCGN8622. The resulting plasmid was designated pCGN8627. DNA sequence analysis confirmed the integrity of the cloning junctions.

10 A fragment containing the *Arabidopsis* ACAT-like coding region (SEQ ID NO: 42) was removed from pCGN8626 by digestion with Sse8387I and Not I. The fragment was ligated into PstI-Not I digested pCGN8623. The resulting plasmid was designated pCGN8628. DNA sequence analysis confirmed the integrity of the cloning junctions.

15 A fragment containing the *Arabidopsis* ACAT-like coding region was removed from pCGN8626 by digestion with Sse8387 and Not I. The fragment was ligated into PstI-Not I digested pCGN8624. The resulting plasmid was designated pCGN8629. DNA sequence analysis confirmed the integrity of the cloning junctions.

20 A fragment containing the *Arabidopsis* ACAT-like coding region was removed from pCGN8626 by digestion with Sse8387 and Not I. The fragment was ligated into PstI-Not I digested pCGN8625. The resulting plasmid was designated pCGN8630. DNA sequence analysis confirmed the integrity of the cloning junctions.

25 An additional expression construct for the suppression of endogenous ACAT-like activity was also prepared. The construct pCGN8660 was constructed by cloning approximately 1 Kb of the *Arabidopsis* ACAT-like coding region from pCGN8626 in the sense orientation, and the full-length *Arabidopsis* ACAT-like coding region in the antisense orientation under the regulatory control of the napin transcription initiation sequence.

For expression of the rat ACAT-like sequence in plants, the NotI-Sse8387I fragment of pCGN8592 was cloned into NotI-PstI digested binary vectors pCGN8621, 30 pCGN8622, and pCGN8624 to yield plasmids, pCGN 9700, pCGN9701, and pCGN9702, respectively. Plasmid pCGN9700 expresses a sense transcript of the rat ACAT-like cDNA under control of a napin promoter, plasmid pCGN9701 expresses an antisense transcript of the rat ACAT-like cDNA under control of a napin promoter, and plasmid pCGN9702 expresses a sense transcript of the rat ACAT-like cDNA under control of a double 35S

promoter. Plasmids pCGN 9700, pCGN9701, and pCGN9702 were introduced in *Agrobacterium tumefaciens* EHA101.

Constructs were prepared to direct the expression of the rat ACAT-like sequence in the seed embryo of soybean and the endosperm of corn. For expression of the rat ACAT-like DNA sequence in soybean, a 1.5 kb *NotI/Sse8387I* fragment from pCGN8592 containing the coding sequence of the rat ACAT-like sequence was blunt ended using Mung bean nuclease, and ligated into the *SmaI* site of the turbo 7S binary/cloning vector pCGN8809 to create the vector pCGN8817 for transformation into soybean by particle bombardment. The vector pCGN8817 contained the operably linked components of the promoter region of the soybean  $\alpha'$  subunit of  $\beta$ -conglycinin (7S promoter, (Chen *et al.*, (1986), *Proc. Natl. Acad. Sci.*, 83:8560-8564), the DNA sequence coding for the entire rat ACAT-like protein, and the transcriptional termination region of pea RuBisCo small subunit, referred to as E9 3' (Coruzzi, *et al.* (1984) *EMBO J.* 3:1671-1679 and Morelli, *et al.* (1985) *Nature* 315:200-204). This construct further contained sequences for the selection of positive transformed plants by screening for resistance to glyphosate using the CP4 EPSPS (U.S. Patent 5,633,435) expressed under the control of the figwort mosaic virus (FMV) promoter (U.S. Patent Number 5,378,619) and the transcriptional termination region of E9.

For expression of the rat ACAT-like sequence in the corn endosperm, a 1.5 kb *NotI/Sse8387I* fragment from pCGN8592 containing the coding sequence of the rat ACAT-like sequence was blunt ended using Mung bean nuclease, and ligated into the *BamHI* site of the rice pGt1 expression cassette pCGN8592 for expression from the pGt1 promoter (Leisy, D.J. *et al.*, *Plant Mol. Biol.* 14 (1989) 41-50) and the HSP70 intron sequence (U.S. Patent Number 5,593,874). This cassette also included the transcriptional termination region downstream of the cloning site of nopaline synthase, *nos* 3' (Depicker *et al.*, *J. Molec. Appl. Genet.* (1982) 1: 562-573). A 7.5 kb fragment containing the pGt1 promoter, the DNA sequence encoding the rat ACAT-like protein, and the *nos* transcriptional termination sequence was cloned into the binary vector pCGN8816 to create the vector pCGN8818 for transformation into corn. This construct also contained sequences for the selection of positive transformants with kanamycin using the kanamycin resistance gene from Tn5 bacteria under the control of the CAMV 35S promoter and tml transcriptional termination regions.



**Example 5: Expression in Insect Cell Culture**

A baculovirus expression system was used to express the LCAT cDNAs in cultured insect cells.

The baculovirus expression constructs pCGN9992, pCGN9993, pCGN9994, pCGN10900, pCGN10962, and pCGN10967 were transformed and expressed using the BAC-to-BAC Baculovirus Expression System (Gibco-BRL, Gaithersburg, MD) according to the manufacturer's directions.

The transformed insect cells were used to assay for acyltransferase activities using methods known in the art (see Example 8).

10

**Example 6: Plant Transformation**

A variety of methods have been developed to insert a DNA sequence of interest into the genome of a plant host to obtain the transcription or transcription and translation of the sequence to effect phenotypic changes. Transgenic plants were obtained by *Agrobacterium*-mediated transformation as described by Radke *et al.* (*Theor. Appl. Genet.* (1988) 75:685-694; *Plant Cell Reports* (1992) 11:499-505). Alternatively, microprojectile bombardment methods, such as described by Klein *et al.* (*Bio/Technology* 10:286-291) may also be used to obtain nuclear transformed plants. Other plant species may be similarly transformed using related techniques.

The plant binary constructs described above were used in plant transformation to direct the expression of the sterol acyltransferases in plant tissues. Binary vector constructs were transformed into strain EHA101 *Agrobacterium* cells (Hood *et al.*, *J. Bacteriol* (1986) 168:1291-1301), by the method of Holsters *et al.* (*Mol. Gen. Genet.* (1978) 163:181-187). Transgenic *Arabidopsis thaliana* plants were obtained by *Agrobacterium*-mediated transformation as described by Valverkens *et al.*, (*Proc. Nat. Acad. Sci.* (1988) 85:5536-5540), Bent *et al.* ((1994), *Science* 265:1856-1860), and Bechtold *et al.* ((1993), *C. R. Acad. Sci., Life Sciences* 316:1194-1199).

**Example 7: Plant Assays for Modified Sterol Content/Profile****7A: NMR of T2 seed**

Seed from plants expressing LCAT 1 through 4 under the control of the napin promoter were analyzed by NMR. Arabidopsis seeds from transgenic plants were placed directly into wide-mouth MAS NMR sample tubes.

High-resolution spectra were measured at 11.7 T ( $^1\text{H}$ =500 MHz,  $^{13}\text{C}$ =125 MHz) using Varian NMR Instruments (Palo Alto, CA) Inova<sup>TM</sup> NMR spectrometers equipped with carbon-observe MAS Nanoprobes<sup>TM</sup>. The  $^{13}\text{C}$  spectra were acquired without a field-frequency lock at ambient temperature (approx. 21-22°C) for 14 hours using the following conditions: spectral width = 29.996 kHz, acquisition time = 2.185 seconds, p/2 pulse (3.8 ms) with no relaxation delay,  $^1\text{H}$  g B2 = 2.5 kHz with Waltz decoupling. Data processing conditions were typically: digital resolution = 0.11 Hz, 0.3 to 1.5 Hz line broadening and time-reversed linear prediction of the first three data points. Chemical shifts were referenced by adding neat tetramethylsilane (TMS) to Arabidopsis seeds and using the resulting referencing parameters for subsequent spectra. The  $^{13}\text{C}$  resolution was 2-3 Hz for the most narrow seed resonances. Spectral resolution was independent of MAS spinning speeds (1.5-3.5 kHz) and data were typically obtained with 1.5 kHz spinning speeds. Spinning sidebands were approx. 1% of the main resonance. Phytosterol  $^{13}\text{C}$  assignments were based on model samples composed of triolein,  $\beta$ -sitosterol and cholesterol oleate. Triacylglycerol  $^{13}\text{C}$  assignments were made from comparison with literature assignments or with shifts computed from a  $^{13}\text{C}$  NMR database (Advanced Chemical Development, Inc., version 3.50, Toronto Canada).

The results of these analyses are displayed in Figure 2 and show that there was a trend of an approximately 2 fold increase of phytosterols in the seeds derived from plant line 5 expressing the LCAT 4 gene (pCGN9998) under the control of the napin promoter. During the course of this analysis it was also noted that the average oil content of seed from plants expressing the LCAT2 construct (pCGN9983) under the control of the napin promoter was higher than that of controls. This is the first *in planta* evidence supporting the concept that overexpression of a nucleotide sequence encoding a lecithin:cholesterol acyltransferase-like polypeptide can increase oil content.

#### 7B: HPLC/MS of T2 seed

Seed oil from T2 plants expressing LCAT1 through 4 under the control of the napin promoter was extracted using an accelerated solvent extractor (ASE) method. Seed samples were ground, using a mortar and pestle, to achieve a fine homogeneous meal. Oil was obtained using a Dionex Accelerated Solvent Extractor (ASE). Clean ground seed was added to an equal amount of diatomaceous earth. The ground seed sample and the diatomaceous earth were thoroughly mixed until a homogeneous texture was achieved.

The sample was then loaded into the instrument and oil extraction was achieved using hexane under validated laboratory protocols.

Oil from these seed samples was then analyzed for sterol ester analysis using HPLC/MS for free campesterol, stigmasterol, and sitosterol and their fatty acid esters. To  
5 the autosampler vial containing approximately 0.1 grams oil was added 0.3 mLs  $\text{CDCl}_3$ . One-hundred microliters of this solution was added to 900 microliters  $\text{CHCl}_3$ . Five microliters of this diluted sample was subsequently injected into an HPLC/MS with positive ion atmospheric pressure ionization. The individual components in the oils were separated using two 4.6 x 50 mm  $\text{C}_8$  Zorbax columns in series and a gradient using  
10 acetonitrile and acetonitrile with 40%  $\text{CHCl}_3$ . The sterol concentrations were calculated assuming each sterol and its fatty acids have the same molar responses. This was observed to be the case with cholesterol and its esters and was assumed to be the case for campesterol, stigmasterol, and sitosterol. In the present study, the sterol identified as stigmasterol was actually an isomer of this compound.

15 The results of these analyses are displayed in Figures 3 and 4 and show that there were sterol ester enhancements on the order of 50% in the seeds derived from six out of seven T2 plant lines expressing LCAT3 (pCGN9968) under the control of the napin promoter.

#### 20 **Example 8: Baculovirus Insect Cell Culture for Sterol Esterification Activity**

Baculovirus expression construct pCGN9992, pCGN9993, pCGN9994 and pCGN10900 (see Example 4) were transformed and expressed using the BAC-TOBAC  
Baculovirus Expression System (Gibco-BRL, Gaithersburg, MD) according to the manufacturer's instructions except harvesting of recombinant viruses was done 5 days  
25 post-transfection. The supernatant from the transfection mixture was used for generating virus stock which in turn was used for infecting Sf9 cells used in the assay.

The transformed cells were assayed for lecithin:sterol acyltransferase activities using the method described herein. Insect cells were centrifuged and the resulting cell pellet was either used immediately or stored at  $-70^\circ\text{C}$  for later analysis. Cells were  
30 resuspended in Medium A (100 mM Tricine/NaOH, pH 7.8, 10% (w/v) glycerol, 280 mM NaCl with : 0.1  $\mu\text{M}$  Aprotinin, 1  $\mu\text{M}$  Leupeptin, and 100  $\mu\text{M}$  Pefabloc (all from Boehringer Mannheim, Germany) and lysed by sonication (2 x 10 sec). Cell walls and other debris were pelleted by centrifugation (14,000 x g, 10 min,  $4^\circ\text{C}$ ). The supernatant

was transferred to a new vial and membranes pelleted by centrifugation (100,000 x g, Ti 70.1 rotor, 46,000 rpm for 1 hour at 4°C). Total membranes were resuspended in Medium A. Lecithin:sterol acyltransferase activity was assayed in a 0.1 ml reaction mixture containing 100 mM Tris/HCl, pH 7, 28 mM NaCl, 0.03% Triton X-100, 0.1 mM sitosterol, 20 µM 1,2-[<sup>14</sup>C]-palmitoyl-phosphatidyl choline (246420 dpm/nmole), and 0.05-20 mg of membrane protein. After 15 minutes at 30 °C, the reaction was terminated by addition of a 0.5 ml solution of methylene chloride:methanol (4:1, v/v ) containing 100 µg cholesterol and cholesterol ester as cold carriers. A portion (0.1 ml) of the bottom organic layer was removed and evaporated under nitrogen gas. The concentrated extract was resuspended in 30 µl of hexane and spotted onto a silica gel-G thin layer chromatographic plate. The plate was migrated in hexane:diethyl ether:acetic acid (80:20:1) to the top, then air dried. Radioactivity was determined by exposure to a Low Energy Phosphor-imaging Screen. Following exposure, the screen was read on a phosphorimager.

The LCAT 4 protein from pCGN10900 in baculovirus membranes showed a radioactive spot in the region of the TLC plate where cholesterol ester migrates indicating that LCAT 4 has the ability to catalyze the transfer of an acyl group from lecithin (PC) to sitosterol to make a sitosterol ester.

#### Example 9: Plant Assay for Modified Lipid Content

Nir (near infrared spectroscopy spectral scanning) can be used to determine the total oil content of Arabidopsis seeds in a non-destructive way provided that a spectral calibration curve has been developed and validated for seed oil content. A seed oil spectral calibration curve was developed using seed samples from 85 Arabidopsis plants. Seed was cleaned and scanned using a Foss NIR model 6500 (Foss-Nirs Systems, Inc.). Approximately 50 to 100 milligrams of whole seeds, per sample, were packed in a mini sample ring cup with quartz lens [ IH-0307 ] consisting a mini-insert [ IH-0337 ] and scanned in reflectance mode to obtain the spectral data. The seed samples were then ground, using a mortar and pestle, to achieve a fine homogeneous meal. The ground samples were measured for oil using an accelerated solvent extractor (ASE).

Measurement for the total oil content was performed on the Dionex Accelerated Solvent Extractor (ASE). Approximately 500 mg of clean ground seed was weighed to the nearest 0.1 mg onto a 9 x 9 cm weigh boat. An equal amount of diatomaceous earth was added using a top-loading balance accurate to the nearest 0.01 g. The ground seed sample

and the diatomaceous earth were thoroughly mixed until a homogeneous texture was achieved. The sample was loaded on to the instrument and oil extraction was achieved using hexane under validated laboratory protocols. Standard Rapeseed samples were obtained from the Community Bureau of Reference (BCR). The ASE extraction method  
5 was validated using the BCR reference standards. A total percent oil recovery of 99% to 100% was achieved. "As-is" oil content was calculated to the nearest 0.01 mass percentage using the formula:

$$\text{Oil Content} = 100\% \times (\text{vial plus extracted oil wt} - \text{initial vial wt}) / (\text{sample wt})$$

10

The analytical data generated by ASE were used to perform spectral calibrations. Nir calibration equations were generated using the built-in statistical package within the NirSystems winisi software. The spectral calibration portion of the software is capable of calibration and self-validation. From a total of 85 samples, 57 samples were used to  
15 generate the total percent oil calibration. The remaining samples were used to validate the oil calibrations. Optimized smoothing, derivative size, and mathematical treatment (modified partial least square) was utilized to generate the calibration. The samples that were not used in building respective calibrations were used as a validation set. Statistical tools such as correlation coefficient ( R ), coefficient of determination ( R<sup>2</sup> ), standard error  
20 of prediction ( SEP ), and the standard error of prediction corrected for bias (SEPC) were used to evaluate the calibration equations.

T2 seeds from plants that had been transformed with the LCAT genes were cleaned and scanned using a Foss NIR model 6500 (Foss-Nirs Systems, Inc.). Approximately 50 to 100 milligrams of whole seeds, per sample, were packed in a mini sample ring cup with  
25 quartz lens [ IH-0307 ] consisting a mini-insert [ IH-0337 ] and scanned in reflectance mode to obtain the spectral data. Oil percentage in each seed sample was determined using the seed oil spectral calibration curve detailed above.

The results of these analyses are found in Figure 5 and Table 2 and show that there was a significant increase in the oil level in seed from T2 plants expressing the LCAT2  
30 gene. This increase in oil was seen in plants when LCAT2 was driven by either the 35S constitutive promoter or the seed-specific napin promoter. These results show that overexpression of a nucleic acid sequence encoding a lecithin:cholesterol acyltransferase-like polypeptide can increase seed oil production in plants.

Table 2

Construct number		Seed Oil Percentage (%)
CONTROL		24.7
CONTROL		28.0
CONTROL		31.8
CONTROL		32.4
NAPIN LCAT1	PCGN9962	28.5
NAPIN LCAT1	PCGN9962	28.9
NAPIN LCAT1	PCGN9962	29.6
NAPIN LCAT1	PCGN9962	30.1
NAPIN LCAT1	PCGN9962	30.1
NAPIN LCAT1	PCGN9962	30.1
NAPIN LCAT1	PCGN9962	30.8
NAPIN LCAT1	PCGN9962	31.0
NAPIN LCAT1	pCGN9962	32.1
NAPIN LCAT1	pCGN9962	34.2
NAPIN LCAT3	pCGN9968	26.8
NAPIN LCAT3	pCGN9968	27.4
NAPIN LCAT3	pCGN9968	29.0
NAPIN LCAT3	pCGN9968	29.0
NAPIN LCAT3	pCGN9968	32.6
NAPIN LCAT2	pCGN9983	26.5
NAPIN LCAT2	pCGN9983	34.7
NAPIN LCAT2	pCGN9983	34.8
NAPIN LCAT2	pCGN9983	35.7
NAPIN LCAT2	pCGN9983	35.8
NAPIN LCAT2	pCGN9983	36.3
NAPIN LCAT2	pCGN9983	36.7
NAPIN LCAT2	pCGN9983	37.0
NAPIN LCAT2	pCGN9983	37.2
NAPIN LCAT2	pCGN9983	37.3
NAPIN LCAT2	pCGN9983	37.3
NAPIN LCAT2	pCGN9983	37.4
NAPIN LCAT2	pCGN9983	37.8
NAPIN LCAT2	pCGN9983	38.0
NAPIN LCAT2	pCGN9983	38.0
35S LCAT2	pCGN9981	27.3
35S LCAT2	pCGN9981	28.1
35S LCAT2	pCGN9981	28.2
35S LCAT2	pCGN9981	28.6
35S LCAT2	pCGN9981	29.8
35S LCAT2	pCGN9981	30.3
35S LCAT2	pCGN9981	32.4
35S LCAT2	pCGN9981	32.5
35S LCAT2	pCGN9981	33.6
35S LCAT2	pCGN9981	34.1
35S LCAT2	pCGN9981	35.5
35S LCAT2	pCGN9981	36.4
35S LCAT2	pCGN9981	37.1
35S LCAT2	pCGN9981	38.3

35S LCAT2	pCGN9981	38.5
35S LCAT2	pCGN9981	39.1

In light of the detailed description of the invention and the examples presented above, it can be appreciated that the several aspects of the invention are achieved.

5 It is to be understood that the present invention has been described in detail by way of illustration and example in order to acquaint others skilled in the art with the invention, its principles, and its practical application. Particular formulations and processes of the present invention are not limited to the descriptions of the specific embodiments presented, but rather the descriptions and examples should be viewed in terms of the  
10 claims that follow and their equivalents. While some of the examples and descriptions above include some conclusions about the way the invention may function, the inventors do not intend to be bound by those conclusions and functions, but put them forth only as possible explanations.

It is to be further understood that the specific embodiments of the present invention  
15 as set forth are not intended as being exhaustive or limiting of the invention, and that many alternatives, modifications, and variations will be apparent to those of ordinary skill in the art in light of the foregoing examples and detailed description. Accordingly, this invention is intended to embrace all such alternatives, modifications, and variations that fall within the spirit and scope of the following claims.

What is claimed is:

1. An isolated nucleic acid sequence comprising a polynucleotide encoding a plant lecithin:cholesterol acyltransferase-like polypeptide or fragment thereof.
2. The isolated nucleic acid sequence of claim 1, wherein said plant lecithin:cholesterol acyltransferase-like polypeptide is selected from the group consisting of *Arabidopsis*, soybean and corn.
3. An isolated nucleic acid sequence comprising a polynucleotide encoding a plant acyl CoA:cholesterol acyltransferase-like polypeptide.
4. The isolated nucleic acid sequence of claim 3, wherein said polynucleotide is SEQ ID NO: 42 or degenerate variants thereof.
5. The isolated nucleic acid sequence of claim 1, wherein said polynucleotide is selected from the group consisting of SEQ ID NO: 2, 4, 6, 8, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 43, 44, 45, 46, 47, 48, 49, 50, 51, 73 and 75 or degenerate variants thereof.
6. An isolated nucleic acid sequence consisting essentially of SEQ ID NO: 2, 4, 6, 8, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 73 or 75.
7. An isolated nucleic acid sequence consisting of SEQ ID NO: 2, 4, 6, 8, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 73 or 75.
8. An isolated nucleic acid sequence comprising a polynucleotide selected from the group consisting of:
  - a) an isolated polynucleotide encoding a polypeptide of SEQ ID NO 3 or SEQ ID NO 3 with at least one conservative amino acid substitution;
  - b) SEQ ID NO: 2;
  - c) an isolated polynucleotide that has at least 70% sequence identity to SEQ ID NO: 2;



- d) an isolated polynucleotide that has at least 80% sequence identity to SEQ ID NO: 2;
- 10 e) an isolated polynucleotide that has at least 90% sequence identity to SEQ ID NO: 2;
- f) an isolated polynucleotide that has at least 95% sequence identity to SEQ ID NO: 2;
- 15 g) an isolated polynucleotide of at least 10 nucleic acids that hybridizes under stringent conditions to SEQ ID NO: 2;
- h) an isolated polynucleotide complementary to a polynucleotide of (a), (b), (c), (d), (e), (f) or (g); and
- i) an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 2 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.
- 20
9. An isolated nucleic acid sequence consisting essentially of a polynucleotide of the formula 5' X-(R<sub>1</sub>)<sub>n</sub>-(R<sub>2</sub>)<sub>n</sub>-(R<sub>3</sub>)<sub>n</sub>-Y 3', where X is hydrogen, Y is hydrogen or a metal, R<sub>1</sub> and R<sub>3</sub> are any nucleic acid, n is an integer between 0-3000, and R<sub>2</sub> is selected from the group consisting of:
- 5 a) an isolated polynucleotide encoding a polypeptide of SEQ ID NO: 3 or SEQ ID NO: 3 with at least one conservative amino acid substitution;
- b) SEQ ID NO: 2;
- c) an isolated polynucleotide that has at least 70% sequence identity to SEQ ID NO: 2;
- 10 d) an isolated polynucleotide that has at least 80% sequence identity to SEQ ID NO: 2;
- e) an isolated polynucleotide that has at least 90% sequence identity to SEQ ID NO: 2;
- f) an isolated polynucleotide that has at least 95% sequence identity to SEQ ID NO: 2;
- 15 g) an isolated polynucleotide of at least 10 nucleic acids that hybridizes under stringent conditions to SEQ ID NO: 2;
- h) an isolated polynucleotide complementary to a polynucleotide of (a), (b), (c), (d), (e), (f) or (g); and

- 20 i) an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 2 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.
10. An isolated nucleic acid sequence comprising a polynucleotide selected from the group consisting of:
- 5 a) an isolated polynucleotide encoding a polypeptide of SEQ ID NO: 5 or SEQ ID NO: 5 with at least one conservative amino acid substitution;
- b) SEQ ID NO: 4;
- c) an isolated polynucleotide having at least 70% sequence identity with SEQ ID NO: 4;
- d) an isolated polynucleotide having at least 80% sequence identity with SEQ ID NO: 4;
- 10 e) an isolated polynucleotide having at least 90% sequence identity with SEQ ID NO: 4;
- f) an isolated polynucleotide having at least 95% sequence identity with SEQ ID NO: 4;
- 15 g) an isolated polynucleotide of at least 10 nucleic acids that hybridizes under stringent conditions to SEQ ID NO: 4;
- h) an isolated polynucleotide complementary to a polynucleotide of (a), (b), (c), (d), (e), (f) or (g); and
- 20 i) an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 4 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.
11. An isolated nucleic acid sequence consisting essentially of a polynucleotide of the formula  $5' X-(R_1)_n-(R_2)_n-(R_3)_n-Y 3'$ , where X is hydrogen, Y is hydrogen or a metal,  $R_1$  and  $R_3$  are any nucleic acid, n is an integer between 0-3000, and  $R_2$  is selected from the group consisting of:
- 5 a) an isolated polynucleotide encoding a polypeptide of SEQ ID NO: 5 or SEQ ID NO: 5 with at least one conservative amino acid substitution;
- b) SEQ ID NO: 4;

- 10 c) an isolated polynucleotide having at least 70% sequence identity with SEQ ID NO: 4;
- d) an isolated polynucleotide having at least 80% sequence identity with SEQ ID NO: 4;
- e) an isolated polynucleotide having at least 90% sequence identity with SEQ ID NO: 4;
- 15 f) an isolated polynucleotide having at least 95% sequence identity with SEQ ID NO: 4;
- g) an isolated polynucleotide of at least 10 nucleic acids that hybridizes under stringent conditions to SEQ ID NO: 4;
- h) an isolated polynucleotide complementary to a polynucleotide of (a), (b), (c), (d), (e), (f) or (g); and
- 20 i) an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 4 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.

12. An isolated nucleic acid sequence comprising a polynucleotide selected from the group consisting of:

- a) an isolated polynucleotide encoding a polypeptide of SEQ ID NO: 7 or SEQ ID NO: 7 with at least one conservative amino acid substitution;
- 5 b) SEQ ID NO: 6;
- c) an isolated polynucleotide having at least 70% sequence identity with SEQ ID NO: 6;
- d) an isolated polynucleotide having at least 80% sequence identity with SEQ ID NO: 6;
- 10 e) an isolated polynucleotide having at least 90% sequence identity with SEQ ID NO: 6;
- f) an isolated polynucleotide having at least 95% sequence identity with SEQ ID NO: 6;
- 15 g) an isolated polynucleotide of at least 10 nucleic acids that hybridizes under stringent conditions to SEQ ID NO: 6;
- h) an isolated polynucleotide complementary to a polynucleotide of (a), (b), (c), (d), (e), (f) or (g); and

- 20 i) an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 6 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.
13. An isolated nucleic acid sequence consisting essentially of a polynucleotide of the formula  $5' X-(R_1)_n-(R_2)_n-(R_3)_n-Y 3'$ , where X is hydrogen, Y is hydrogen or a metal,  $R_1$  and  $R_3$  are any nucleic acid, n is an integer between 0-3000, and  $R_2$  is selected from the group consisting of:
- 5 a) an isolated polynucleotide encoding a polypeptide of SEQ ID NO: 7 or SEQ ID NO: 7 with at least one conservative amino acid substitution;
- b) SEQ ID NO: 6;
- c) an isolated polynucleotide having at least 70% sequence identity with SEQ ID NO: 6;
- 10 d) an isolated polynucleotide having at least 80% sequence identity with SEQ ID NO: 6;
- e) an isolated polynucleotide having at least 90% sequence identity with SEQ ID NO: 6;
- f) an isolated polynucleotide having at least 95% sequence identity with SEQ ID NO: 6;
- 15 g) an isolated polynucleotide of at least 10 nucleic acids that hybridizes under stringent conditions to SEQ ID NO: 6;
- h) an isolated polynucleotide complementary to a polynucleotide of (a), (b), (c), (d), (e), (f) or (g); and
- 20 i) an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 6 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.
14. An isolated nucleic acid sequence comprising a polynucleotide selected from the group consisting of
- a) an isolated polynucleotide encoding a polypeptide of SEQ ID NO: 9 or SEQ ID NO: 9 with at least one conservative amino acid substitution;
- 5 b) SEQ ID NO 8;

- c) an isolated polynucleotide having at least 70% sequence identity with SEQ ID NO: 8;
- d) an isolated polynucleotide having at least 80% sequence identity with SEQ ID NO: 8;
- 10 e) an isolated polynucleotide having at least 90% sequence identity with SEQ ID NO: 8;
- f) an isolated polynucleotide having at least 95% sequence identity with SEQ ID NO: 8;
- 15 g) an isolated polynucleotide of at least 10 nucleic acids that hybridizes under stringent conditions to SEQ ID NO: 8;
- h) an isolated polynucleotide complementary to a polynucleotide of (a), (b), (c), (d), (e), (f) or (g); and
- i) an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 8 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.

15. An isolated nucleic acid sequence consisting essentially of a polynucleotide of the formula 5' X-(R<sub>1</sub>)<sub>n</sub>-(R<sub>2</sub>)<sub>n</sub>-(R<sub>3</sub>)<sub>n</sub>-Y 3', where X is hydrogen, Y is hydrogen or a metal, R<sub>1</sub> and R<sub>3</sub> are any nucleic acid, n is an integer between 0-3000, and R<sub>2</sub> is selected from the group consisting of:

- 5 a) an isolated polynucleotide encoding a polypeptide of SEQ ID NO: 9 or SEQ ID NO: 9 with at least one conservative amino acid substitution;
- b) SEQ ID NO: 8;
- c) an isolated polynucleotide having at least 70% sequence identity with SEQ ID NO: 8;
- 10 d) an isolated polynucleotide having at least 80% sequence identity with SEQ ID NO: 8;
- e) an isolated polynucleotide having at least 90% sequence identity with SEQ ID NO: 8;
- f) an isolated polynucleotide having at least 95% sequence identity with SEQ ID NO: 8;
- 15 g) an isolated polynucleotide of at least 10 nucleic acids that hybridizes under stringent conditions to SEQ ID NO: 8;

- h) an isolated polynucleotide complementary to a polynucleotide of (a), (b), (c), (d), (e), (f) or (g); and
- 20 i) an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 8 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.
16. An isolated nucleic acid sequence comprising a polynucleotide selected from the group consisting of:
- a) an isolated polynucleotide encoding a polypeptide of SEQ ID NO: 74 or SEQ ID NO: 74 with at least one conservative amino acid substitution;
- 5 b) SEQ ID NO: 73;
- c) an isolated polynucleotide having at least 70% sequence identity with SEQ ID NO: 73;
- d) an isolated polynucleotide having at least 80% sequence identity with SEQ ID NO: 73;
- 10 e) an isolated polynucleotide having at least 90% sequence identity with SEQ ID NO: 73;
- f) an isolated polynucleotide having at least 95% sequence identity with SEQ ID NO: 73;
- g) an isolated polynucleotide of at least 10 nucleic acids that hybridizes under  
15 stringent conditions to SEQ ID NO: 73;
- h) an isolated polynucleotide complementary to a polynucleotide of (a), (b), (c), (d), (e), (f) or (g); and
- i) an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 73 and encodes a plant lecithin:cholesterol acyltransferase-like  
20 polypeptide.
17. An isolated nucleic acid sequence consisting essentially of a polynucleotide of the formula 5' X-(R<sub>1</sub>)<sub>n</sub>-(R<sub>2</sub>)<sub>n</sub>-(R<sub>3</sub>)<sub>n</sub>-Y 3', where X is hydrogen, Y is hydrogen or a metal, R<sub>1</sub> and R<sub>3</sub> are any nucleic acid, n is an integer between 0-3000, and R<sub>2</sub> is selected from the group consisting of:
- 5 a) an isolated polynucleotide encoding a polypeptide of SEQ ID NO: 74 or SEQ ID NO: 74 with at least one conservative amino acid substitution;

- b) SEQ ID NO: 73;
- c) an isolated polynucleotide having at least 70% sequence identity with SEQ ID NO: 73;
- 10 d) an isolated polynucleotide having at least 80% sequence identity with SEQ ID NO: 73;
- e) an isolated polynucleotide having at least 90% sequence identity with SEQ ID NO: 73;
- f) an isolated polynucleotide having at least 95% sequence identity with SEQ ID  
15 NO: 73;
- g) an isolated polynucleotide of at least 10 nucleic acids that hybridizes under stringent conditions to SEQ ID NO: 73;
- ) h) an isolated polynucleotide complementary to a polynucleotide of (a), (b), (c), (d), (e), (f) or (g); and
- 20 i) an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 73 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.

18. A isolated nucleic acid sequence comprising a polynucleotide selected from the group consisting of:

- a) an isolated polynucleotide encoding a polypeptide of SEQ ID NO: 76 or SEQ ID NO: 76 with at least one conservative amino acid substitution;
- 5 b) SEQ ID NO: 75;
- ) c) an isolated polynucleotide having at least 70% sequence identity with SEQ ID NO: 75;
- d) an isolated polynucleotide having at least 80% sequence identity with SEQ ID NO: 75;
- 10 e) an isolated polynucleotide having at least 90% sequence identity with SEQ ID NO: 75;
- f) an isolated polynucleotide having at least 95% sequence identity with SEQ ID NO: 75;
- 15 g) an isolated polynucleotide of at least 10 nucleic acids that hybridizes under stringent conditions to SEQ ID NO: 75;

- h) an isolated polynucleotide complementary to a polynucleotide of (a), (b), (c), (d), (e), (f) or (g); and
- i) an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 75 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.

19. An isolated nucleic acid sequence consisting essentially of a polynucleotide of the formula 5' X-(R<sub>1</sub>)<sub>n</sub>-(R<sub>2</sub>)<sub>n</sub>-(R<sub>3</sub>)<sub>n</sub>-Y 3', where X is hydrogen, Y is hydrogen or a metal, R<sub>1</sub> and R<sub>3</sub> are any nucleic acid, n is an integer between 0-3000, and R<sub>2</sub> is selected from the group consisting of:

- a) an isolated polynucleotide encoding a polypeptide of SEQ ID NO: 76 or SEQ ID NO: 76 with at least one conservative amino acid substitution;
- b) SEQ ID NO: 75;
- c) an isolated polynucleotide having at least 70% sequence identity with SEQ ID NO: 75;
- d) an isolated polynucleotide having at least 80% sequence identity with SEQ ID NO: 75;
- e) an isolated polynucleotide having at least 90% sequence identity with SEQ ID NO: 75;
- f) an isolated polynucleotide having at least 95% sequence identity with SEQ ID NO: 75;
- g) an isolated polynucleotide of at least 10 nucleic acids that hybridizes under stringent conditions to SEQ ID NO: 75;
- h) an isolated polynucleotide complementary to a polynucleotide of (a), (b), (c), (d), (e), (f) or (g); and
- i) an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 75 and encodes a plant lecithin:cholesterol acyltransferase-like polypeptide.

20. An isolated nucleic acid sequence comprising a polynucleotide selected from the group consisting of:

- a) SEQ ID NO: 42 or a degenerate variant thereof;



- b) an isolated polynucleotide having at least 70% sequence identity with SEQ ID NO: 42;
- c) an isolated polynucleotide having at least 80% sequence identity with SEQ ID NO: 42;
- d) an isolated polynucleotide having at least 90% sequence identity with SEQ ID NO: 42;
- e) an isolated polynucleotide having at least 95% sequence identity with SEQ ID NO: 42;
- f) an isolated polynucleotide of at least 10 nucleic acids that hybridizes under stringent conditions to SEQ ID NO: 42;
- g) an isolated polynucleotide complementary to a polynucleotide of (a), (b), (c), (d), (e), or (f); and
- h) an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 42 and encodes an acyl CoA:cholesterol acyltransferase-like polypeptide.

21. An isolated nucleic acid sequence consisting essentially of a polynucleotide of the formula 5' X-(R<sub>1</sub>)<sub>n</sub>-(R<sub>2</sub>)<sub>n</sub>-(R<sub>3</sub>)<sub>n</sub>-Y 3', where X is hydrogen, Y is hydrogen or a metal R<sub>1</sub> and R<sub>3</sub> are any nucleic acid, n is an integer between 0 and 3000, and R<sub>2</sub> is selected from the group consisting of:

- a) SEQ ID NO: 42 or degenerate variants thereof;
- b) an isolated polynucleotide having at least 70% sequence identity to SEQ ID NO: 42;
- c) an isolated polynucleotide having at least 80% sequence identity to SEQ ID NO: 42;
- d) an isolated polynucleotide having at least 90% sequence identity to SEQ ID NO: 42;
- e) an isolated polynucleotide having at least 95% sequence identity to SEQ ID NO: 42;
- f) an isolated polynucleotide of at least 10 nucleic acids that hybridizes under stringent conditions to SEQ ID NO: 42;
- g) an isolated polynucleotide complementary to a polynucleotide of (a), (b), (c), (d), (e), or (f); and

h) an isolated polynucleotide that hybridizes under stringent conditions to SEQ ID NO: 42 and encodes an acyl CoA:cholesterol acyltransferase-like polypeptide.

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22. A recombinant nucleic acid construct comprising a regulatory sequence operably linked to polynucleotide encoding a lecithin:cholesterol acyltransferase-like polypeptide or a fragment thereof.
23. The recombinant nucleic acid construct of claim 22, wherein said lecithin:cholesterol acyltransferase-like polypeptide is a plant lecithin:cholesterol acyltransferase-like polypeptide.
24. A recombinant nucleic acid construct comprising a regulatory sequence operably linked to a polynucleotide encoding an acyl CoA:cholesterol acyltransferase-like polypeptide.
25. The recombinant nucleic acid construct of claim 24, wherein said acyl CoA:cholesterol acyltransferase-like polypeptide is a plant acyl CoA:cholesterol acyltransferase-like polypeptide.
26. The recombinant construct of claim 22, wherein said regulatory sequence comprises a heterologous regulatory sequence.
27. The recombinant construct of claim 24, wherein said regulatory sequence comprises a heterologous regulatory sequence.
28. The recombinant construct of claim 22, wherein said regulatory sequence is functional in a plant cell.
29. The recombinant construct of claim 24, wherein said regulatory sequence is functional in a plant cell.
30. The recombinant construct of claim 22, further comprising a termination sequence.

31. The recombinant construct of claim 24 further comprising a termination sequence.
32. The recombinant construct of claim 22 wherein said polynucleotide is selected from the group consisting of SEQ ID NO: 2, 4, 6, 8, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 43, 44, 45, 46, 47, 48, 49, 50, 51, 73 and 75.
33. The recombinant construct of claim 24, wherein said polynucleotide is selected from the group consisting of SEQ ID NO: 33 and 42.
34. The recombinant construct of claim 22, wherein said regulatory sequence comprises a constitutive promoter.
35. The recombinant construct of claim 24, wherein said regulatory sequence comprises a constitutive promoter.
36. The recombinant construct of claim 22, wherein said regulatory sequence comprises an inducible promoter.
37. The recombinant construct of claim 24, wherein said regulatory sequence comprises an inducible promoter.
38. The recombinant construct of claim 22, wherein said regulatory sequence is selected from the group consisting of a tissue specific promoter, a developmentally regulated promoter, an organelle specific promoter, and a seed specific promoter.
39. The recombinant construct of claim 24, wherein said regulatory sequence is selected from the group consisting of a tissue specific promoter, a developmentally regulated promoter, an organelle specific promoter, and a seed specific promoter.
40. A host cell containing the recombinant construct of claim 22 or 24.

41. The host cell of claim 40, wherein said host cell is selected from the group consisting of plant cells, animal cells, insect cells, yeast, bacteria, bacteriophage and viruses.
42. The host cell of claim 40, wherein said host cell is a plant cell.
43. The host cell of claim 40, wherein said host cell expresses a lecithin:cholesterol acyltransferase-like polypeptide or an acyl CoA:cholesterol acyltransferase-like polypeptide.
44. The host cell of claim 43, wherein said cholesterol acyltransferase-like polypeptide is a plant acyltransferase-like polypeptide.
45. A plant comprising at least one host cell of claim 40.
46. The progeny of a plant of claim 45.
47. A seed from the plant of claim 45.
48. A plant comprising the recombinant construct of claim 22 or 24.
49. The progeny of a plant of claim 48.
50. A seed from the plant of claim 48.
51. A purified polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO: 3, SEQ ID NO: 3 with at least one conservative amino acid substitution, SEQ ID NO: 5, SEQ ID NO: 5 with at least one conservative amino acid substitution, SEQ ID NO: 7, SEQ ID NO: 7 with at least one conservative amino acid substitution, SEQ ID NO: 9, SEQ ID NO: 9 with at least one conservative amino acid substitution, SEQ ID NO: 74, SEQ ID NO: 74 with at least one conservative amino acid substitution, SEQ ID NO: 76 and SEQ ID NO: 76 with at least one conservative amino acid substitution.

52. A purified immunogenic polypeptide comprising at least 10 consecutive amino acids from an amino acid sequence selected from the group consisting of SEQ ID NO: 3, SEQ ID NO: 3 with at least one conservative amino acid substitution, SEQ ID NO: 5, SEQ ID NO: 5 with at least one conservative amino acid substitution, SEQ ID NO: 7, SEQ ID NO: 7 with at least one conservative amino acid substitution, SEQ ID NO: 9, SEQ ID NO: 9 with at least one conservative amino acid substitution, SEQ ID NO: 74, SEQ ID NO: 74 with at least one conservative amino acid substitution, SEQ ID NO: 76 and SEQ ID NO: 76 with at least one conservative amino acid substitution.
53. An antibody which specifically binds to an immunogenic polypeptide of claim 52.
54. A method for producing a lecithin:cholesterol acyltransferase-like polypeptide or an acyl CoA:cholesterol acyltransferase-like polypeptide comprising culturing a host cell of claim 40 under conditions permitting expression of said lecithin:cholesterol acyltransferase-like polypeptide or acyl CoA:cholesterol acyltransferase-like polypeptide.
55. The method of claim 54, further comprising isolating the cholesterol acyltransferase-like polypeptide from the host cell or from the medium in which the host cell is cultured.
56. A method for modifying the sterol content of a host cell, comprising transforming a host cell with a recombinant construct containing a regulatory sequence operably linked to a polynucleotide encoding a lecithin:cholesterol acyltransferase-like polypeptide and culturing said host cell under conditions wherein said host cell expresses a lecithin:cholesterol acyltransferase-like polypeptide such that said host cell has a modified sterol composition as compared to host cells without the recombinant construct.
57. The method of claim 56, wherein said lecithin:cholesterol acyltransferase-like polypeptide is a plant lecithin:cholesterol acyltransferase-like polypeptide.

58. A method for modifying the sterol content of a host cell, comprising transforming a host cell with a recombinant construct containing a regulatory sequence operably linked to a polynucleotide encoding an acyl CoA:cholesterol acyltransferase-like polypeptide and culturing said host cell under conditions wherein said host cell expresses an acyl CoA:cholesterol acyltransferase-like polypeptide such that said host cell has a modified sterol composition as compared to host cells without the recombinant construct.
59. The method of claim 58, wherein said acyl CoA:cholesterol acyltransferase-like polypeptide is a plant acyl CoA:cholesterol acyltransferase-like polypeptide.
60. The method of claim 56, wherein said modified sterol composition is an increase in sterol esters.
61. The method of claim 58, wherein said modified sterol composition is an increase in sterol esters.
62. The method of claim 56, wherein said polynucleotide encoding a lecithin:cholesterol acyltransferase-like polypeptide is selected from the group consisting of SEQ ID NO: 2, 4, 6, 8, 10, 11, 73 and 75.
63. The method of claim 58 wherein said polynucleotide encoding a acyl CoA:cholesterol acyltransferase-like polypeptide is SEQ ID NO 33 OR 42.
64. The method of claim 56, wherein said regulatory sequence comprises a constitutive promoter.
65. The method of claim 58, wherein said regulatory sequence comprises a constitutive promoter.
66. The method of claim 56, wherein said regulatory sequence is an inducible promoter.
67. The method of claim 58, wherein said regulatory sequence is an inducible promoter.

68. The method of claim 56, wherein said regulatory sequence is a tissue specific promoter.
69. The method of claim 58, wherein said regulatory sequence is a tissue specific promoter.
70. The method of claim 56, wherein said regulatory sequence is a seed specific promoter.
71. The method of claim 58, wherein said regulatory sequence is a seed specific promoter.
72. The method of claim 56, wherein said polynucleotide encoding a lecithin:cholesterol acyltransferase-like polypeptide is in the antisense orientation.
73. The method of claim 58, wherein said polynucleotide encoding an acyl CoA:cholesterol acyltransferase-like polypeptide is in the antisense orientation.
74. The method of claim 72, wherein said modified sterol composition is a decrease in sterol esters.
75. The method of claim 73, wherein said modified sterol composition is a decrease in sterol esters.
76. A plant comprising a recombinant construct containing a regulatory sequence operably linked to a polynucleotide encoding a lecithin:cholesterol acyltransferase-like polypeptide wherein expression of said recombinant construct results in modified sterol composition of said plant as compared to the same plant without said recombinant construct.
77. The plant of claim 76, wherein said lecithin:cholesterol acyltransferase-like polypeptide is a plant lecithin:cholesterol acyltransferase-like polypeptide.

78. The plant of claim 76, wherein said polynucleotide encoding a lecithin:cholesterol acyltransferase-like polypeptide is selected from the group consisting of SEQ ID NO: 2, 4, 6, 8, 10, 11, 73 and 75.
79. A plant comprising a recombinant construct containing a regulatory sequence operably linked to a polynucleotide encoding an acyl CoA:cholesterol acyltransferase-like polypeptide wherein expression of said recombinant construct results in modified sterol composition of said plant as compared to the same plant without said recombinant construct.
80. The plant of claim 79, wherein said acyl CoA:cholesterol acyltransferase-like polypeptide is a plant acyl CoA:cholesterol acyltransferase-like polypeptide.
81. The plant of claim 79, wherein said polynucleotide encoding an acyl CoA:cholesterol acyltransferase-like polypeptide is SEQ ID NO: 33 or 42.
82. The plant of claim 76, wherein said regulatory sequence comprises a tissue specific promoter.
83. The plant of claim 79, wherein said regulatory sequence comprises a tissue specific promoter.
84. The plant of claim 76, wherein said regulatory sequence comprises a seed specific promoter.
85. The plant of claim 79, wherein said regulatory sequence comprises a seed specific promoter.
86. The plant of claim 76, wherein said modified sterol composition is an increase in sterol esters.
87. The plant of claim 79, wherein said modified sterol composition is an increase in sterol esters.



88. The plant of claim 76, wherein the polynucleotide encoding a lecithin:cholesterol acyltransferase-like polypeptide is in the antisense orientation.
89. The plant of claim 79, wherein the polynucleotide encoding an acyl CoA:cholesterol acyltransferase-like polypeptide is in the antisense orientation.
90. An oil obtained from the plant of claim 76 or 79.
91. A method for producing an oil with a modified sterol composition comprising, providing a plant of claim 76 or 79 and extracting the oil from said plant.
92. An oil produced by the method of claim 91.
93. A method for altering oil production by a host cell comprising, transforming a host cell with a recombinant construct containing a regulatory sequence operably linked to a polynucleotide encoding a lecithin:cholesterol acyltransferase-like polypeptide and culturing said host cell under conditions wherein said host cell expresses a  
5 lecithin:cholesterol acyltransferase-like polypeptide such that said host cell has an altered oil production as compared to host cells without the recombinant construct.
94. The method of claim 93, wherein said lecithin:cholesterol acyltransferase-like polypeptide is a plant lecithin:cholesterol acyltransferase-like polypeptide.
95. A method for altering oil production by a host cell comprising, transforming a host cell with a recombinant construct containing a regulatory sequence operably linked to a polynucleotide encoding an acyl CoA:cholesterol acyltransferase-like polypeptide and culturing said host cell under conditions wherein said host cell expresses an acyl  
5 CoA:cholesterol acyltransferase-like polypeptide such that said host cell has an altered oil production as compared to host cells without the recombinant construct.
96. The method of claim 95, wherein said acyl CoA:cholesterol acyltransferase-like polypeptide is a plant acyl CoA:cholesterol acyltransferase-like polypeptide.

97. The method of claim 93, wherein said oil production is increased.
98. The method of claim 95, wherein said oil production is increased.
99. The method of claim 93, wherein said host cell is a plant cell.
100. The method of claim 95, wherein said host cell is a plant cell.
101. The method of claim 93, wherein said polynucleotide encoding a lecithin:cholesterol acyltransferase-like polypeptide is selected from the group consisting of SEQ ID NO: 2, 4, 6, 8, 10, 11, 73 and 75.
102. The method of claim 95, wherein said polynucleotide encoding an acyl CoA:cholesterol acyltransferase-like polypeptide is SEQ ID NO: 33 or 42.
103. The method of claim 93, wherein said regulatory sequence is a tissue specific promoter.
104. The method of claim 95, wherein said regulatory sequence is a tissue specific promoter.
105. The method of claim 93, wherein said regulatory sequence is a seed specific promoter.
106. The method of claim 95, wherein said regulatory sequence is a seed specific promoter.
107. A plant comprising a recombinant construct containing a regulatory sequence operably linked to a polynucleotide encoding a lecithin:cholesterol acyltransferase-like polypeptide wherein expression of said recombinant construct results in an altered production of oil by said plant as compared to the same plant without said recombinant construct.

108. The plant of claim 107, wherein said lecithin:cholesterol acyltransferase-like polypeptide is a plant lecithin:cholesterol acyltransferase-like polypeptide.
109. A plant comprising a recombinant construct containing a regulatory sequence operably linked to a polynucleotide encoding an acyl CoA:cholesterol acyltransferase-like polypeptide wherein expression of said recombinant construct results in an altered production of oil by said plant as compared to the same plant without said recombinant construct.
- 5 110. The plant of claim 109, wherein said acyl CoA:cholesterol acyltransferase-like polypeptide is a plant acyl CoA:cholesterol acyltransferase-like polypeptide.
111. The plant of claim 107, wherein said oil production is increased.
112. The plant of claim 109, wherein said oil production is increased.
113. The plant of claim 107, wherein said polynucleotide encoding a lecithin:cholesterol acyltransferase-like polypeptide is selected from the group consisting of SEQ ID NO: 2, 4, 6, 8, 10, 11, 73 and 75.
114. The plant of claim 109, wherein said polynucleotide encoding an acyl CoA:cholesterol acyltransferase-like polypeptide is SEQ ID NO: 33 or 42.
115. The plant of claim 107, wherein said regulatory sequence is a tissue specific promoter.
116. The plant of claim 109, wherein said regulatory sequence is a tissue specific promoter.
117. The plant of claim 107, wherein said regulatory sequence is a seed specific promoter.
118. The plant of claim 109, wherein said regulatory sequence is a seed specific promoter.
119. A food product comprising the oil of claim 90 or 92.

120. A food product comprising the plant of claim 107 or 109.

## SEQUENCE LISTING

&lt;110&gt; Monsanto Company

&lt;120&gt; PLANT STEROL ACYLTRANSFERASES

&lt;130&gt; MTC6718

5 &lt;140&gt;

&lt;141&gt;

&lt;150&gt; 60/152,493

&lt;151&gt; 1999-08-30

&lt;160&gt; 80

10 &lt;170&gt; PatentIn Ver. 2.1

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       65                    70                    75                    80

10 Asp Asn Ile Glu Val Leu Val Pro Asp Asp Asp His Gly Leu Tyr Ala  
                     85                    90                    95

Ile Asp Ile Leu Asp Pro Ser Trp Phe Val Lys Leu Cys His Leu Thr  
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Glu Val Tyr His Phe His Asp Met Ile Glu Met Leu Val Gly Cys Gly  
                     115                    120                    125

15 Tyr Lys Lys Gly Thr Thr Leu Phe Gly Tyr Gly Tyr Asp Phe Arg Gln  
                     130                    135                    140

Ser Asn Arg Ile Asp Leu Leu Ile Leu Gly Leu Lys Lys Lys Leu Glu  
       145                    150                    155                    160

20 Thr Ala Tyr Lys Arg Ser Gly Gly Arg Lys Val Thr Ile Ile Ser His  
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Ser Met Gly Gly Leu Met Val Ser Cys Phe Met Tyr Leu His Pro Glu  
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Ala Phe Ser Lys Tyr Val Asn Lys Trp Ile Thr Ile Ala Thr Pro Phe  
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25 Gln Gly Ala Pro Gly Cys Ile Asn Asp Ser Ile Leu Thr Gly Val Gln  
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Phe Val Glu Gly Leu Glu Ser Phe Phe Phe Val Ser Arg Trp Thr Met  
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30 His Gln Leu Leu Val Glu Cys Pro Ser Ile Tyr Glu Met Met Ala Asn  
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Pro Asp Phe Lys Trp Lys Lys Gln Pro Glu Ile Arg Val Trp Arg Lys  
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Lys Ser Glu Asn Asp Val Asp Thr Ser Val Glu Leu Glu Ser Phe Gly  
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35 Leu Ile Glu Ser Ile Asp Leu Phe Asn Asp Ala Leu Lys Asn Asn Glu  
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Leu Ser Tyr Gly Gly Asn Lys Ile Ala Leu Pro Phe Asn Phe Ala Ile  
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 5 Pro Phe Asp Val Cys Tyr Gly Thr Glu Thr Ser Pro Ile Asp Asp Leu  
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 Ser Glu Ile Cys Gln Thr Met Pro Glu Tyr Thr Tyr Val Asp Gly Asp  
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35 <220>  
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<220>  
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 <222> (418)  
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 gccaaaggata ttgctgttgc caggacgata gctccaggat ttttanataa cnatctgttt 360  
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 <211> 311  
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 <213> Zea mays

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 <221> unsure  
 <222> (1)..(311)  
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 <212> DNA  
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&lt;210&gt; 24

&lt;211&gt; 227

&lt;212&gt; DNA

25 &lt;213&gt; Zea mays

&lt;220&gt;

&lt;221&gt; unsure

&lt;222&gt; (1) .. (227)

&lt;223&gt; n=unknown

30 &lt;400&gt; 24

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    tgggggggatt acttgtgaaa tgtntcatct cactgcacag tgatatatnt gaaaaatatg 180
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35 &lt;210&gt; 25

&lt;211&gt; 1587

&lt;212&gt; DNA

&lt;213&gt; Zea mays

&lt;220&gt;

40 &lt;221&gt; unsure

&lt;222&gt; (1) .. (1587)

&lt;223&gt; n=unknown

&lt;400&gt; 25

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    catctgtcta tgggtgatca cttccatgat atgattgata tgctcataaa ctgtggatat 240
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    <213> Zea mays

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    <221> unsure
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30 <223> n=unknown

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atgaatcatg atgtgagttt tcatgttttc tgtgtttttt ttgcttttgc ataaatatcc 180
35 atgtcaattt cccccatttt ctaggtatc actangtatg tcaacaaatg gatttgcatt 240
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<210> 27
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<212> DNA
40 <213> Zea mays

    <220>
    <221> unsure
    <222> (1)..(1240)
    <223> n=unknown

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aagascaata ttgaactcat agwagsgaca aatgggtggaa atagggtggt ggkmgatccc 180

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acnactccat ggggtcnttn atttntgtcn ttttacgnaa tggntcgaag ccctcctccg 240  
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 atctttctta ggagttccta aggctgttgc tgggcttttt ttcttctaag caaaagatgt 360  
 tgccggttgc taggtataag taatgattca tttattttaa gcaaaaggga atagcaaaag 420  
 5 aatgaatatt attggatgct cgacaagctt gcgagcttt tgctcccaag ccctctctg 480  
 gacctcacia gtccaggag tgcctgcctc tgatcctcat catcaggaac aggtcaagt 540  
 atgcaccgac ggtaccgtga ggtcatttct atcctgatgc aacaccatgt acttgttgat 600  
 ggcaagggtca ggactgacaa gacctaccct gctgggttca tggatgtcat ttccatccct 660  
 aagacaaacg agaactacag gctgctttcg tcttcaccca atcagggatg aggatgccaa 720  
 10 gttcaagctc tacaagggtga ggtctgttca gtttggccag aaagacatcc cctatctgaa 780  
 cacctacgac gaccgcacca tccgctaccc cgaccgcctc atcaaggcca acgacaccat 840  
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 ggtcatggtg atcggcagga ggaataccgg gcgtgttaga gtgatcaara taaggagaa 960  
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 15 ttctcctatt tgtgtacag gaaaacatag aatgaaattc aaatttggtg gccacaaaag 1080  
 tgtggagact tgatttcata taaagttagg cttaacatta gtgcaaacag ttgtatttta 1140  
 gtttagattt agagtacact atgtatgctg tgtttgacaa tgcttattta tgatatattg 1200  
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 <213> Zea mays

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 cggctgaagg cggggaggaa aatagctgct tgaaaggggg tgtttactta gccgatggtg 180  
 atgaaactgt tccagttctt agtgcgggct acatgtgtgc aaaaggatgg cgtggcaaaa 240  
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 ctcccatggg ggggtggcggg ggtccagact ggtgtgagaa gcatattaaa gctgtaatga 180  
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aagtggggct gatccaacag tggatgggcc ctactatcca gaactccatg gaagcccttt 420  
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<211> 299

<212> DNA

10 <213> Mus musculus

<400> 31

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ccacgactac tacgtgctca actacgatgc cccagtgggt catgagctac tgccaaaggc 180  
15 agccctccct aacctgggcc tggagttctg gaggggttcc tggtgctg cactctctc 240  
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<210> 32

<211> 1895

<212> DNA

20 <213> Artificial Sequence

<220>

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<220>

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<223> n=unknown

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35 cggccaatgt ctttgcctg gctgcattcc aggttgagaa gcgctggcg gtgggtgccc 420  
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cagcggtgt ggtcttactg gttgagtcta tctctcaggt gggctccctg ctggcgctga 540  
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ccaccttggtg ctacgagctc aactttcccc gctctcccc catccggaag cgctttctgc 780  
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&lt;210&gt; 33

&lt;211&gt; 1766

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&lt;213&gt; Artificial Sequence

&lt;220&gt;

<223> Description of Artificial Sequence: Inferred cDNA  
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				20					25						30				
10	Leu	Val	Val	Asp	Tyr	Ile	Asp	Glu	Gly	Arg	Leu	Val	Leu	Glu	Phe	Ser			
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	Leu	Leu	Ser	Tyr	Ala	Phe	Gly	Lys	Phe	Pro	Thr	Val	Val	Trp	Thr	Trp			
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	His	Trp	Arg	Thr	Gly	Tyr	Ser	Lys	Ser	Ser	His	Pro	Leu	Ile	Arg	Ser			
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				165						170					175				
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	Cys	Phe	Phe	Tyr	Val	Tyr	Tyr	Ile	Phe	Glu	Arg	Leu	Cys	Ala	Pro	Leu			
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35	225					230					235					240			
	Cys	Val	Phe	Asn	Ser	Ile	Leu	Pro	Gly	Val	Leu	Ile	Leu	Phe	Leu	Thr			
				245						250					255				
	Phe	Phe	Ala	Phe	Leu	His	Cys	Trp	Leu	Asn	Ala	Phe	Ala	Glu	Met	Leu			
				260					265					270					

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 305 310 315 320  
 Lys Ser Ala Ala Met Leu Ala Val Phe Ala Val Ser Ala Val Val His  
 325 330 335  
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 Val Leu Phe Met Phe Phe Gly Met Ala Phe Asn Phe Ile Val Asn Asp  
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 Ser Arg Lys Lys Pro Ile Trp Asn Val Leu Met Trp Thr Ser Leu Phe  
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 20 25 30  
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 Trp Ala Met Phe Leu Ser Thr Leu Ser Ile Pro Tyr Phe Leu Phe Gln  
 65 70 75 80  
 Pro Trp Ala His Gly Tyr Ser Lys Ser Ser His Pro Leu Ile Tyr Ser  
 35 85 90 95  
 Leu Val His Gly Leu Leu Phe Leu Val Phe Gln Leu Gly Val Leu Gly  
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Phe Val Pro Thr Tyr Val Val Leu Ala Tyr Thr Leu Pro Pro Ala Ser  
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 130 135 140  
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 30 Ala Leu Ile Asp Tyr Tyr Tyr Gln His Asn Gly Ser Phe Lys Asp Ser  
 35 40 45  
 Glu Ile Leu Lys Phe Met Thr Thr Asn Leu Phe Thr Val Ala Ser Val  
 50 55 60  
 35 Asp Leu Leu Met Tyr Leu Ser Thr Tyr Phe Val Val Gly Ile Gln Tyr  
 65 70 75 80  
 Leu Cys Lys Trp Gly Val Leu Lys Trp Gly Thr Thr Gly Trp Ile Phe  
 85 90 95



Thr Ser Ile Tyr Glu Phe Leu Phe Val Ile Phe Tyr Met Tyr Leu Thr  
 100 105 110  
 Glu Asn Ile Leu Lys Leu His Trp Leu Ser Lys Ile Phe Leu Phe Leu  
 115 120 125  
 5 His Ser Leu Val Leu Leu Met Lys Met His Ser Phe Ala Phe Tyr Asn  
 130 135 140  
 Gly Tyr Leu Trp Gly Ile Lys Glu Glu Leu Gln Phe Ser Lys Ser Ala  
 145 150 155 160  
 10 Leu Ala Lys Tyr Lys Asp Ser Ile Asn Asp Pro Lys Val Ile Gly Ala  
 165 170 175  
 Leu Glu Lys Ser Cys Glu Phe Cys Ser Phe Glu Leu Ser Ser Gln Ser  
 180 185 190  
 Leu Ser Asp Gln Thr Gln Lys Phe Pro Asn Asn Ile Ser Ala Lys Ser  
 195 200 205  
 15 Phe Phe Trp Phe Thr Met Phe Pro Thr Leu Ile Tyr Gln Ile Glu Tyr  
 210 215 220  
 Pro Arg Thr Lys Glu Ile Arg Trp Ser Tyr Val Leu Glu Lys Ile Cys  
 225 230 235 240  
 20 Ala Ile Phe Gly Thr Ile Phe Leu Met Met Ile Asp Ala Gln Ile Leu  
 245 250 255  
 Met Tyr Pro Val Ala Met Arg Ala Leu Ala Val Arg Asn Ser Glu Trp  
 260 265 270  
 Thr Gly Ile Leu Asp Arg Leu Leu Lys Trp Val Gly Leu Leu Val Asp  
 275 280 285  
 25 Ile Val Pro Gly Phe Ile Val Met Tyr Ile Leu Asp Phe Tyr Leu Ile  
 290 295 300  
 Trp Asp Ala Ile Leu Asn Cys Val Ala Glu Leu Thr Arg Phe Gly Asp  
 305 310 315 320  
 30 Arg Tyr Phe Tyr Gly Asp Trp Trp Asn Cys Val Ser Trp Ala Asp Phe  
 325 330 335  
 Ser Arg Ile Trp Asn Ile Pro Val His Lys Phe Leu Leu Arg His Val  
 340 345 350  
 Tyr His Ser Ser Met Ser Ser Phe Lys Leu Asn Lys Ser Gln Ala Thr  
 355 360 365  
 35 Leu Met Thr Phe Phe Leu Ser Ser Val Val His Glu Leu Ala Met Tyr  
 370 375 380  
 Val Ile Phe Lys Lys Leu Arg Phe Tyr Leu Phe Phe Phe Gln Met Leu  
 385 390 395 400

Gln Met Pro Leu Val Ala Leu Thr Asn Thr Lys Phe Met Arg Asn Arg  
 405 410 415

Thr Ile Ile Gly Asn Val Ile Phe Trp Leu Gly Ile Cys Met Gly Pro  
 420 425 430

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<210> 38  
 <211> 1942  
 <212> DNA  
 <213> Arabidopsis thaliana

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 tccgctcttt cctctccat tagattctgt ttctctttc aatttcttct gcatgcttct 180  
 cgattctctc tgacgcctct tttctccga cgtgtttcg tcaaacgctt ttcgaaatgg 240  
 15 cgattttgga ttctgctggc gttactacgg tgacggagaa cgggtggcga gagttcgtcg 300  
 atcttgatag gcttcgtcga cggaaatcga gatcggattc ttctaacgga cttcttctct 360  
 ctggttccga taataattct ccttcggatg atgttgagc tcccgcgcac gttagggatc 420  
 ggattgattc cgttgtaaac gatgacgctc agggaaacagc caatttggcc ggagataata 480  
 acggtggtgg cgataataac ggtggtggaa gaggcggcgg agaaggaaga ggaaacgccg 540  
 20 atgctacgtt tacgtatcga ccgtcggttc cagctcatcg gagggcgaga gagagtccac 600  
 ttagctccga cgcaatcttc aaacagagcc atgccggatt attcaacctc tgtgtagtag 660  
 ttcttattgc tgtaaacagt agactcatca tcgaaaatct tatgaagtat ggttggttga 720  
 tcagaacgga tttctggtt agttcaagat cgctcgcaga ttggccgctt ttcattgtgt 780  
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 tgtatccagt ttacgtcacc ctaagggtgtg attctgcttt tttatcaggt gtcactttga 960  
 tgctctcac ttgcattgtg tggctaaagt tggtttctta tgctcact agctatgaca 1020  
 taagacgctt agccaatgca gctgataagg ccaatcctga agtctcctac tacgttagct 1080  
 tgaagacctt ggcatatttc atggtcgtct ccacattgtg ttatcagcca agttatccac 1140  
 30 gttctgcatg tatacggag ggttgggtgg ctctcaatt tgcaaaactg gtcatttca 1200  
 ccggattcat gggatttata atagaacaat atataaatcc tattgtcagg aactcaaagc 1260  
 atcctttgaa aggcatctt ctatatgcta ttgaaagagt gttgaagctt tcagttccaa 1320  
 atttatatgt gtggtctctc atgttctact gcttcttcca cctttggtta aacatattgg 1380  
 cagagcttct ctgcttcggg gatcgtgaat tctacaaaga ttggtggaat gcaaaaagtg 1440  
 35 tgggagatta ctggagaatg tggaatatgc ctgttcataa atggatggtt cgacatatat 1500  
 acttcccgtg cttgcgcagc aagataccaa agacactcgc cattatcatt gctttcctag 1560  
 tctctgcagt ctttcatgag ctatgcatcg cagttccttg tegtctcttc aagctatggg 1620  
 cttttcttgg gattatgttt caggtgcctt tggcttctat cacaactat ctacaggaaa 1680  
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 40 cgatgtgtgt gcttctttat taccacgacc tgatgaaccg aaaaggatcg atgtcatgaa 1800  
 acaactgttc aaaaaatgac tttcttcaaa catctatggc ctctgttgat ctccgttgat 1860  
 gttgtggtgg ttctgatgct aaaacgacaa atagtgttat aaccattgaa gaagaaaaga 1920  
 caattagagt tggtgtatcg ca 1942

<210> 39  
 45 <211> 520  
 <212> PRT  
 <213> Arabidopsis thaliana

<400> 39  
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1 5 10 15  
5 Gly Gly Glu Phe Val Asp Leu Asp Arg Leu Arg Arg Arg Lys Ser Arg  
20 25 30  
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35 40 45  
Pro Ser Asp Asp Val Gly Ala Pro Ala Asp Val Arg Asp Arg Ile Asp  
50 55 60  
10 Ser Val Val Asn Asp Asp Ala Gln Gly Thr Ala Asn Leu Ala Gly Asp  
65 70 75 80  
Asn Asn Gly Gly Gly Asp Asn Asn Gly Gly Gly Arg Gly Gly Gly Glu  
85 90 95  
15 Gly Arg Gly Asn Ala Asp Ala Thr Phe Thr Tyr Arg Pro Ser Val Pro  
100 105 110  
Ala His Arg Arg Ala Arg Glu Ser Pro Leu Ser Ser Asp Ala Ile Phe  
115 120 125  
Lys Gln Ser His Ala Gly Leu Phe Asn Leu Cys Val Val Val Leu Ile  
130 135 140  
20 Ala Val Asn Ser Arg Leu Ile Ile Glu Asn Leu Met Lys Tyr Gly Trp  
145 150 155 160  
Leu Ile Arg Thr Asp Phe Trp Phe Ser Ser Arg Ser Leu Arg Asp Trp  
165 170 175  
25 Pro Leu Phe Met Cys Cys Ile Ser Leu Ser Ile Phe Pro Leu Ala Ala  
180 185 190  
Phe Thr Val Glu Lys Leu Val Leu Gln Lys Tyr Ile Ser Glu Pro Val  
195 200 205  
Val Ile Phe Leu His Ile Ile Ile Thr Met Thr Glu Val Leu Tyr Pro  
210 215 220  
30 Val Tyr Val Thr Leu Arg Cys Asp Ser Ala Phe Leu Ser Gly Val Thr  
225 230 235 240  
Leu Met Leu Leu Thr Cys Ile Val Trp Leu Lys Leu Val Ser Tyr Ala  
245 250 255  
35 His Thr Ser Tyr Asp Ile Arg Ser Leu Ala Asn Ala Ala Asp Lys Ala  
260 265 270  
Asn Pro Glu Val Ser Tyr Tyr Val Ser Leu Lys Ser Leu Ala Tyr Phe  
275 280 285

Met Val Ala Pro Thr Leu Cys Tyr Gln Pro Ser Tyr Pro Arg Ser Ala  
 290 295 300  
 Cys Ile Arg Lys Gly Trp Val Ala Arg Gln Phe Ala Lys Leu Val Ile  
 305 310 315 320  
 5 Phe Thr Gly Phe Met Gly Phe Ile Ile Glu Gln Tyr Ile Asn Pro Ile  
 325 330 335  
 Val Arg Asn Ser Lys His Pro Leu Lys Gly Asp Leu Leu Tyr Ala Ile  
 340 345 350  
 10 Glu Arg Val Leu Lys Leu Ser Val Pro Asn Leu Tyr Val Trp Leu Cys  
 355 360 365  
 Met Phe Tyr Cys Phe Phe His Leu Trp Leu Asn Ile Leu Ala Glu Leu  
 370 375 380  
 Leu Cys Phe Gly Asp Arg Glu Phe Tyr Lys Asp Trp Trp Asn Ala Lys  
 385 390 395 400  
 15 Ser Val Gly Asp Tyr Trp Arg Met Trp Asn Met Pro Val His Lys Trp  
 405 410 415  
 Met Val Arg His Ile Tyr Phe Pro Cys Leu Arg Ser Lys Ile Pro Lys  
 420 425 430  
 20 Thr Leu Ala Ile Ile Ile Ala Phe Leu Val Ser Ala Val Phe His Glu  
 435 440 445  
 Leu Cys Ile Ala Val Pro Cys Arg Leu Phe Lys Leu Trp Ala Phe Leu  
 450 455 460  
 Gly Ile Met Phe Gln Val Pro Leu Val Phe Ile Thr Asn Tyr Leu Gln  
 465 470 475 480  
 25 Glu Arg Phe Gly Ser Thr Val Gly Asn Met Ile Phe Trp Phe Ile Phe  
 485 490 495  
 Cys Ile Phe Gly Gln Pro Met Cys Val Leu Leu Tyr Tyr His Asp Leu  
 500 505 510  
 30 Met Asn Arg Lys Gly Ser Met Ser  
 515 520

&lt;210&gt; 40

&lt;211&gt; 29

&lt;212&gt; DNA

&lt;213&gt; Artificial Sequence

&lt;220&gt;

<223> Description of Artificial Sequence: Synthetic  
oligonucleotide primer

&lt;400&gt; 40

5 tgcaaattga cgagcacacc aaccccttc

29

&lt;210&gt; 41

&lt;211&gt; 28

&lt;212&gt; DNA

&lt;213&gt; Artificial Sequence

10 &lt;220&gt;

<223> Description of Artificial Sequence: Synthetic  
oligonucleotide primer

&lt;400&gt; 41

aaggatgctt tgagttcctg acaatagg

28

15 &lt;210&gt; 42

&lt;211&gt; 1942

&lt;212&gt; DNA

&lt;213&gt; Arabidopsis thaliana

&lt;400&gt; 42

20 ctctcgtgaa tcctttttcc tttctttctc ttcttctctt cagagaaaaac tttgcttctc 60  
 tttctataag gaaccagaca cgaatcccat tcccaccgat ttcttagctt ctctcttcaa 120  
 tccgctcttt cctcttccat tagattctgt ttctctttc aatttcttct gcatgcttct 180  
 cgattctctc tgacgcctct tttctcccga cgctgtttcg tcaaacgctt ttcgaaatgg 240  
 cgatttttga ttctgctggc gttactacgg tgacggagaa cgggtggcga gagttcgtcg 300  
 25 atcttgatag gttcgtcga cggaatcga gatcggatc ttctaaccga cttcttctct 360  
 ctgggtccga taataattct ccttcggatg atgttgagc tcccgcgcac gttagggatc 420  
 ggattgattc cggttgtaac gatgacgctc agggaaacagc caatttgccc ggagataata 480  
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 30 ttagctccga cgcaatcttc aaacagagcc atgccggatt attcaacctc tgtgtagtag 660  
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 tcagaacgga tttctgggtt agttcaagat cgctgcgaga ttggccgctt ttcattgtgt 780  
 gtatatccct ttcgatcttt cctttggctg cctttacggg tgagaaattg gtacttcaga 840  
 aatacatatc agaacctgtt gtcattcttc ttcatattat tatcaccatg acagagggtt 900  
 35 tgtatccagt ttacgtcacc ctaagggtgtg attctgcttt tttatcaggg gtcactttga 960  
 tgctectcac ttgcattgtg tggctaaagt tgggtttctta tgctcatact agctatgaca 1020  
 taagatccct agccaatgca gctgataagg ccaatcctga agtctctac tacgttagct 1080  
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 gttctgcatg tatacggaa ggttggtgtg ctgcgtcaatt tgcaaaactg gtcataattca 1200  
 40 ccggattcat gggatttata atagaacaat atataaatcc tattgtcagg aactcaaagc 1260  
 atccctttgaa aggcgatctt ctatatgcta ttgaaagagt gttgaagctt tcagttccaa 1320  
 atttatatgt gtggctctgc atgttctact gcttcttcca cctttgggta aacatattgg 1380  
 cagagcttct ctgcttcggg gatcgtgaat tctacaaaga ttggtggaat gcaaaaagt 1440  
 tgggagatta ctggagaatg tggaaatagc ctgttcataa atggatgggt cgacatatat 1500  
 45 acttcccgtg cttgcgcagc aagataccaa agacactcgc cattatcatt gctttcctag 1560  
 tctctgcagt ctttcatgag ctatgcacg cagttccttg tcgtctcttc aagctatggg 1620  
 cttttcttgg gattatgttt caggtgcctt tgggtcttcat cacaactat ctacaggaaa 1680  
 ggtttggtc aacggtgggg aacatgatct tctggttcat cttctgcatt ttcggacaac 1740

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cgatgtgtgt gcttctttat taccacgacc tgatgaaccg aaaaggatcg atgtcatgaa 1800
acaactgttc aaaaaatgac tttcttcaaa catctatggc ctcgttggat ctccgttgat 1860
gttgtgggtg ttctgatgct aaaacgacaa atagtgttat aaccattgaa gaagaaaaa 1920
caattagagt tggtgtatcg ca 1942

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5 <210> 43  
 <211> 234  
 <212> DNA  
 <213> Glycine max

<220>  
 10 <221> unsure  
 <222> (1) .. (234)  
 <223> n=unknown

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<400> 43
gtaagcttca agagcttagc atanttcctg gttgccccta ncattatggt accagccaan 60
15 ctatcctcgc acaccttata ttcgaaaggg ttggtgttt cgccaacttg tcaactgata 120
atatttacag gagttatggg atttataata gaacaatata ttaatcccat tgtacaaaat 180
tcacagcatc ctctcaaggg aaaccttctt tacgccatcg agagagttct gaag 234

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<210> 44  
 <211> 267  
 20 <212> DNA  
 <213> Glycine max

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<400> 44
ctgcttttgt atctgggtgc acgttgatgc tattaacttg catttgtgtg ttaaaattgg 60
tgatcatatgc acatacaaac tatgatatga gagcacttac tgtttcgaat gaaaagggag 120
25 aaacattacc caatactttg atatggagta tccgtacact gtgaccttca ggagtttggc 180
atacttcatg gttgctccta cattatgcta tcagacaagc tatcctcgca caccttcagt 240
tcgaaagggg tgggtgtttc gtcaact 267

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<210> 45  
 <211> 275  
 30 <212> DNA  
 <213> Glycine max

<220>  
 <221> unsure  
 <222> (1) .. (275)  
 35 <223> n=unknown

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<400> 45
gtggaatgcc aaaactgttg aagattattg gaggatgtgg aatatgcctg ttcacaaatg 60
gatgatccgc cacctatatt ttccatgttt aaggcacggg ataccaaagg ccgttgctct 120
tttaattgcc ttcttggttc tgctttattc catgagctgt gcacgctgt tccttgccca 180
40 catattcaag tngtgggttt cngnggaatt nagtttcagg tnccttgggt ttcnaccnna 240
atnntnnggc naaaaaatc cngaaccoc ggggg 275

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<210> 46  
<211> 257  
<212> DNA  
<213> Glycine max

5 <400> 46  
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ggcttttggg aaaactggca tgcttccttc aacaagtggc ttgtgaggta tatatacatt 120  
cctcttgggg gatctaagaa aaagctacta aatgtgtggg ttgttttcac atttgttgca 180  
atctggcatg atttagagtg gaagcttctt tcatgggcat gggtgacgtg tttattcttc 240  
10 atccctgagt tggtttt 257

<210> 47  
<211> 253  
<212> DNA  
<213> Zea mays

15 <400> 47  
agaaaatgga acatgcctgt gcataaatgg attgttcgtc atatatattt tccttgcattg 60  
cgaaatggta tatcaaagga agttgctgtt tttatatcgt tcttgtttct gctgtacttc 120  
atgagttatg tgttgcgtgt ccttgccaca tactcaagtt ctgggctttt tttaggaatc 180  
atgcttcaga ttccctcat catattgaca tcataacctca aaaataaatt cagtgcacaca 240  
20 atggttggca ata 253

<210> 48  
<211> 254  
<212> DNA  
<213> Zea mays

25 <400> 48  
tgaagtatgg cttattaata agatctggct tttggtttaa tgctacatca ttgcgagact 60  
ggccactgct aatgtgttgc cttagtctac ccatatttcc ccttggtgca tttgcagtcg 120  
aaaagtggc attcaacaat ctcattagtg atcctgctac tacctgtttt cacatccttt 180  
ttacaacatt tgaaattgta tatccagtgc tcgtgattct taagtgtgat tctgcagttt 240  
30 tatcaggctt tgtg 254

<210> 49  
<211> 262  
<212> DNA  
<213> Zea mays

35 <400> 49  
gaagtatggc ttattaataa gatctggctt ttggtttaat gctacatcat tgcgagactg 60  
gccactgcta atgtgttggc ttagtctacc catatttccc cttggtgcat ttgcagtcga 120  
aaagtggca ttcaacaatc tcattagtga tctgctact acctgtttc acatcctttt 180  
tacaacattt gaaattgtat atccagtgc cgtgattctt aagtgtgatt ctgcagtttt 240  
40 acaggctttg tgtgatgtt ta 262

<210> 50  
<211> 325  
<212> DNA  
<213> Zea mays

<220>  
 <221> unsure  
 <222> (1)..(325)  
 <223> n=unknown

5 <400> 50  
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 cacatantca natctnggca tnnncgggat catngttcag ataccgntgg nattcttgac 120  
 aagatatctc catgctacgt tcaagcatgt aatgggtggc aacatgatan tttgntctn 180  
 cagtatagtc ggacagccga tgnnnnnna tctatactac catgacgtca tgaacaggca 240  
 10 ggcccaggca agtagatagt ncggcagaga catgtacttc aacatcganc atcagnagca 300  
 nacngagcga gcggcangaa ncagc 325

<210> 51  
 <211> 519  
 <212> DNA  
 15 <213> Mortierrella alpina

<220>  
 <221> unsure  
 <222> (1)..(519)  
 <223> n=unknown

20 <400> 51  
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 ttanactnaa ttngaaaatn cnncccaac ttnagnnact tnnagncccc ccnacttgac 120  
 aacggagcac tatatttacc cgtggtngt tcaaccacgc catctcacc ttgcgagcat 180  
 tgggtgctgct cttgataccc ttcattgctta actatctcat gatcttttac atcattttcg 240  
 25 agtgcacctg caacgccttt gcggaactaa gttgctttgc ggatcgcaac ttttacgagg 300  
 attgggtgaa ctgcgtcage tttgatgagt gggcacgcaa atggaacaag cctgtgcaac 360  
 acttcttgct ccgccacgtg tacgactoga gcacccgagt ccttccactt gtccgaaatc 420  
 caatgccgcn aattgcaaac gttccttccc ggtcgtcaat gcgttcaacg aacctgggtg 480  
 aagaatgggt ggtgacaacg ttaaagtgcg cccggtatc 519

30 <210> 52  
 <211> 45  
 <212> DNA  
 <213> Artificial Sequence

<220>  
 35 <223> Description of Artificial Sequence:  
 Oligonucleotide primer

<400> 52  
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<210> 53  
 40 <211> 40  
 <212> DNA  
 <213> Artificial Sequence



<220>

<223> Description of Artificial Sequence:  
Oligonucleotide primer

<400> 53

5 ggatccccctg caggtcattc attgacggca ttaacattgg

40

<210> 54

<211> 44

<212> DNA

<213> Artificial Sequence

10 <220>

<223> Description of Artificial Sequence: Synthetic  
oligonucleotide primer

<400> 54

ggatccgcgg ccgcacaatg ggagcgaatt cgaaatcagt aacg

44

15 <210> 55

<211> 40

<212> DNA

<213> Artificial Sequence

<220>

20 <223> Description of Artificial Sequence: Synthetic  
oligonucleotide primer

<400> 55

ggatccccctg caggttaata cccactttta tcaagctccc

40

<210> 56

25 <211> 41

<212> DNA

<213> Artificial Sequence

<220>

30 <223> Description of Artificial Sequence: Synthetic  
oligonucleotide primer

<400> 56

ggatccgcgg ccgcacaatg tctctattac tggaagagat c

41

<210> 57

<211> 41

35 <212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Synthetic  
oligonucleotide primer

<400> 57  
ggatcccctg caggttatgc atcaacagag acacttacag c 41

<210> 58  
<211> 41  
5 <212> DNA  
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<220>  
<223> Description of Artificial Sequence: Synthetic  
oligonucleotide primer

10 <400> 58  
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<210> 59  
<211> 38  
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15 <213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: Synthetic  
oligonucleotide primer

20 <400> 59  
ggatcccctg caggttaacc agaatcaact actttgtg 38

<210> 60  
<211> 39  
<212> DNA  
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25 <220>  
<223> Description of Artificial Sequence:  
Oligonucleotide primer

<400> 60  
tcgacctgca ggaagcttag aaatggcgat tttggattc 39

30 <210> 61  
<211> 36  
<212> DNA  
<213> Artificial Sequence

<220>  
35 <223> Description of Artificial Sequence:  
Oligonucleotide primer

<400> 61  
ggatccgcgg ccgctcatga catcgatcct tttcgg 36

- <210> 62  
<211> 56  
<212> DNA  
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- 5 <220>  
<223> Description of Artificial Sequence: Annealed  
oligonucleotide adapter
- <400> 62  
cgcgatttaa atggcgcgcc ctgcaggcgg ccgcctgcag ggcgcgccat taaat 56
- 10 <210> 63  
<211> 32  
<212> DNA  
<213> Artificial Sequence
- <220>  
15 <223> Description of Artificial Sequence: Ligating  
oligonucleotide
- <400> 63  
tcgaggatcc gcggccgcaa gcttcctgca gg 32
- <210> 64  
20 <211> 32  
<212> DNA  
<213> Artificial Sequence
- <220>  
<223> Description of Artificial Sequence: Ligating  
25 oligonucleotide
- <400> 64  
tcgacctgca ggaagcttgc ggccgcggat cc 32
- <210> 65  
<211> 32  
30 <212> DNA  
<213> Artificial Sequence
- <220>  
<223> Description of Artificial Sequence: Ligating  
oligonucleotide
- 35 <400> 65  
tcgacctgca ggaagcttgc ggccgcggat cc 32

- <210> 66  
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<212> DNA  
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- 5 <220>  
<223> Description of Artificial Sequence: Ligating  
oligonucleotide
- <400> 66  
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- 10 <210> 67  
<211> 36  
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- <220>  
15 <223> Description of Artificial Sequence: Ligating  
oligonucleotide
- <400> 67  
tcgaggatcc gcggccgcaa gcttcctgca ggagct 36
- <210> 68  
20 <211> 28  
<212> DNA  
<213> Artificial Sequence
- <220>  
<223> Description of Artificial Sequence: Ligating  
25 oligonucleotide
- <400> 68  
cctgcaggaa gcttgccggcc gcggatcc 28
- <210> 69  
<211> 36  
30 <212> DNA  
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## ClustalW Formatted Alignments

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Yeast (YNR008W)	G S A K R N E R G K D F D R K R D G N G R K R W R D S R R L I F I L G A F L G V L P F S F G A Y H				
Human LCAT					
Rat LCAT					
AlLCAT1		M G L P G S P W Q W V L - - - L L L G L L L P P A T S F W L L N N V L			
AlLCAT2			M K K - - - I S S H Y S V V I A I L V V V T M T S		
AlLCAT3			M G A N S K - - - S V T A S F T V I A V F L I C N G R N		
AlLCAT4			M S L - - - L L E E I I R S V E A L L K L R N N		

	110	120	130	140	150
Yeast (YNR008W)	VHNSDSDLFD	NFVNFD	SLKVYLDDWKD	VLLPQG	ISSFIDDIQA
Human LCAT	FPPTHTPKAE	LSN	HTR	VLVPG	CL
Rat LCAT	FPPTHTPKAE	LSN	HTR	VLVPG	CM
A1LCAT1	MCQAVGSNVY		PLILVPG	NG	
A1LCAT2	TAVEDETFHGDY		SKLSGI	IPGFA	
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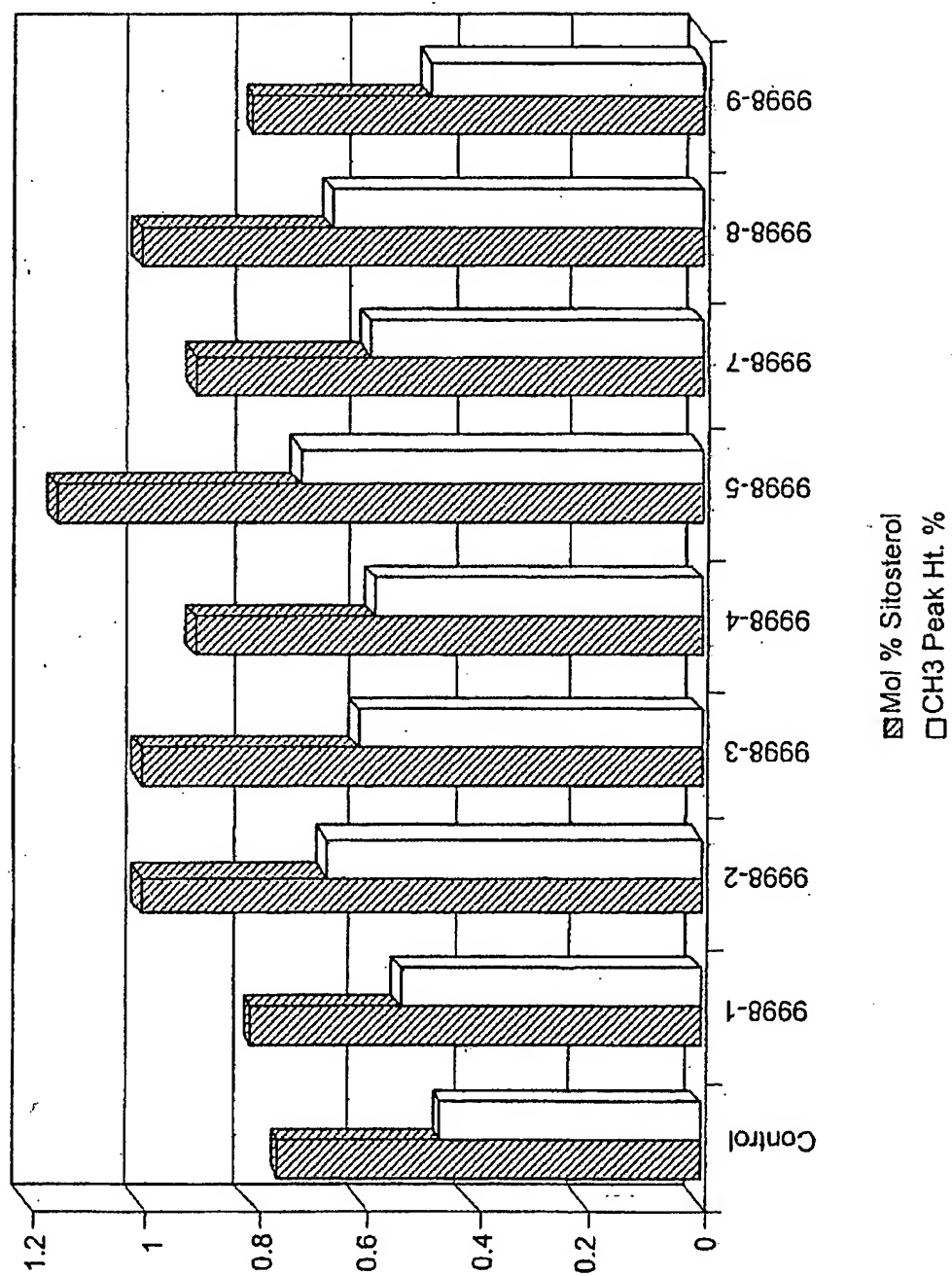
FIG. 1B

Yeast (YNR008W)	160	170	180	190	200
Human LCAT	DDLS	ENFA	VCKQL	LRDY	NI
Rat LCAT	DDLS	ENFA	VCKQL	LRDY	NI
AlLCAT1	DDLS	ENFA	VCKQL	LRDY	NI
AlLCAT2	DDLS	ENFA	VCKQL	LRDY	NI
AlLCAT3	DDLS	ENFA	VCKQL	LRDY	NI
AlLCAT4	DDLS	ENFA	VCKQL	LRDY	NI
Yeast (YNR008W)	210	220	230	240	250
Human LCAT	ECDS	ALNM	FFLP	LP	LP
Rat LCAT	ECDS	ALNM	FFLP	LP	LP
AlLCAT1	ECDS	ALNM	FFLP	LP	LP
AlLCAT2	ECDS	ALNM	FFLP	LP	LP
AlLCAT3	ECDS	ALNM	FFLP	LP	LP
AlLCAT4	ECDS	ALNM	FFLP	LP	LP
Yeast (YNR008W)	260	270	280	290	300
Human LCAT	AAQGF	ES	TD	YFI	AG
Rat LCAT	AAQGF	ES	TD	YFI	AG
AlLCAT1	AAQGF	ES	TD	YFI	AG
AlLCAT2	AAQGF	ES	TD	YFI	AG
AlLCAT3	AAQGF	ES	TD	YFI	AG
AlLCAT4	AAQGF	ES	TD	YFI	AG

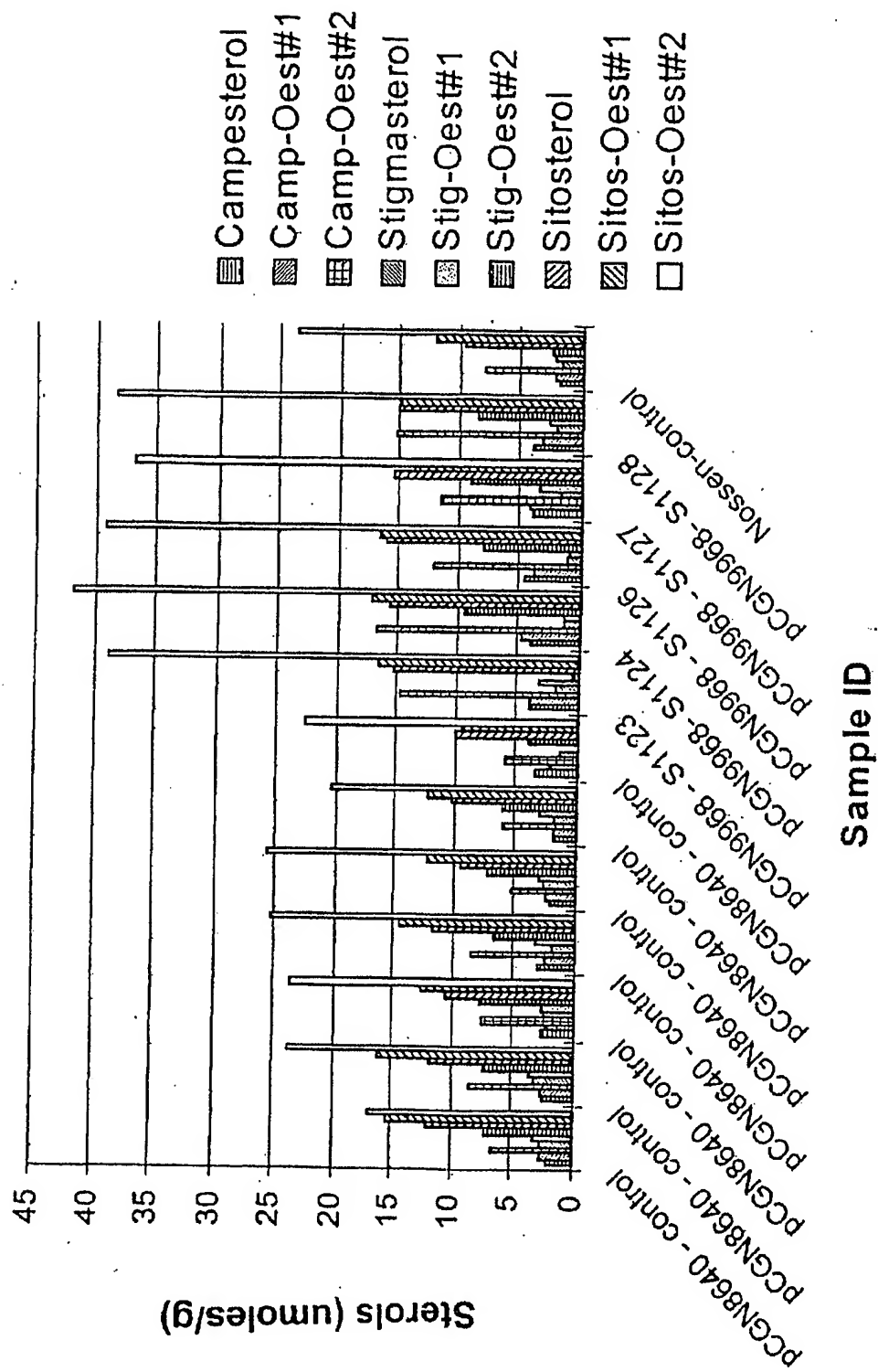
FIG. 1C

Yeast (YNR008W)	310	320	330	340	350
Human LCAT	A Y	L D L E R R D R Y F T K I K E Q I E L F H Q L S C G	C L I G H S M G S Q I I F Y F F F F F F F		
Rat LCAT	E P	G Q Q E E Y Y R K K L A G L V E E M H A A Y C G	C L I G H S M G S Q I I F Y F F F F F F F		
Ai LCAT1	A P	G Q Q E E Y Y R K K L A G L V E E M H A A Y C G	C L I G H S M G S Q I I F Y F F F F F F F		
Ai LCAT2	G L A A S G H P S R V A S Q F L Q Q D L K K L K E Q I E L F H Q L S C G	C L I G H S M G S Q I I F Y F F F F F F F			
Ai LCAT3	S P	T K L E E R D L Y F H K L K L T F E E M H A A Y C G	C L I G H S M G S Q I I F Y F F F F F F F		
Ai LCAT4	S S	R L O E T L D Q F A K K L E E E L F H Q L S C G	C L I G H S M G S Q I I F Y F F F F F F F		
Yeast (YNR008W)	360	370	380	390	400
Human LCAT	M K W V E A E G P L Y G N G G R G W V N E H T D S F T N A G T L L G A P K A V P A L I S S G E M K D				
Rat LCAT	T L R	Q P Q S W K D D R F F I D D G G F I S L G A P P M L V L A S S G D D N			
Ai LCAT1	L L N R	Q P Q S W K D D R F F I D D G G F I S L G A P P M L V L A S S G D D N			
Ai LCAT2	L E W L R L E I A P K	T T P S W L D Q H I H A Y F A V G A P P M L V L A S S G D D N			
Ai LCAT3	M G L	H S D I F E K Y V V Q N K W I T I A T P P M L V L A S S G D D N			
Ai LCAT4	M Y L	H P E A F S K Y V V Q N K W I T I A T P P M L V L A S S G D D N			
Yeast (YNR008W)	410	420	430	440	450
Human LCAT	T I Q L N T L A M Y G L E K F F S R I E R V K M L Q T W G G I P S M L P K G E E V I W G D M				
Rat LCAT	- - - - -	Q P Q S W K D D R F F I D D G G F I S L G A P P M L V L A S S G D D N			
Ai LCAT1	- - - - -	Q P Q S W K D D R F F I D D G G F I S L G A P P M L V L A S S G D D N			
Ai LCAT2	- - - - -	Q P Q S W K D D R F F I D D G G F I S L G A P P M L V L A S S G D D N			
Ai LCAT3	- - - - -	Q P Q S W K D D R F F I D D G G F I S L G A P P M L V L A S S G D D N			
Ai LCAT4	- - - - -	Q P Q S W K D D R F F I D D G G F I S L G A P P M L V L A S S G D D N			

FIG. 2



## Sterol Distribution





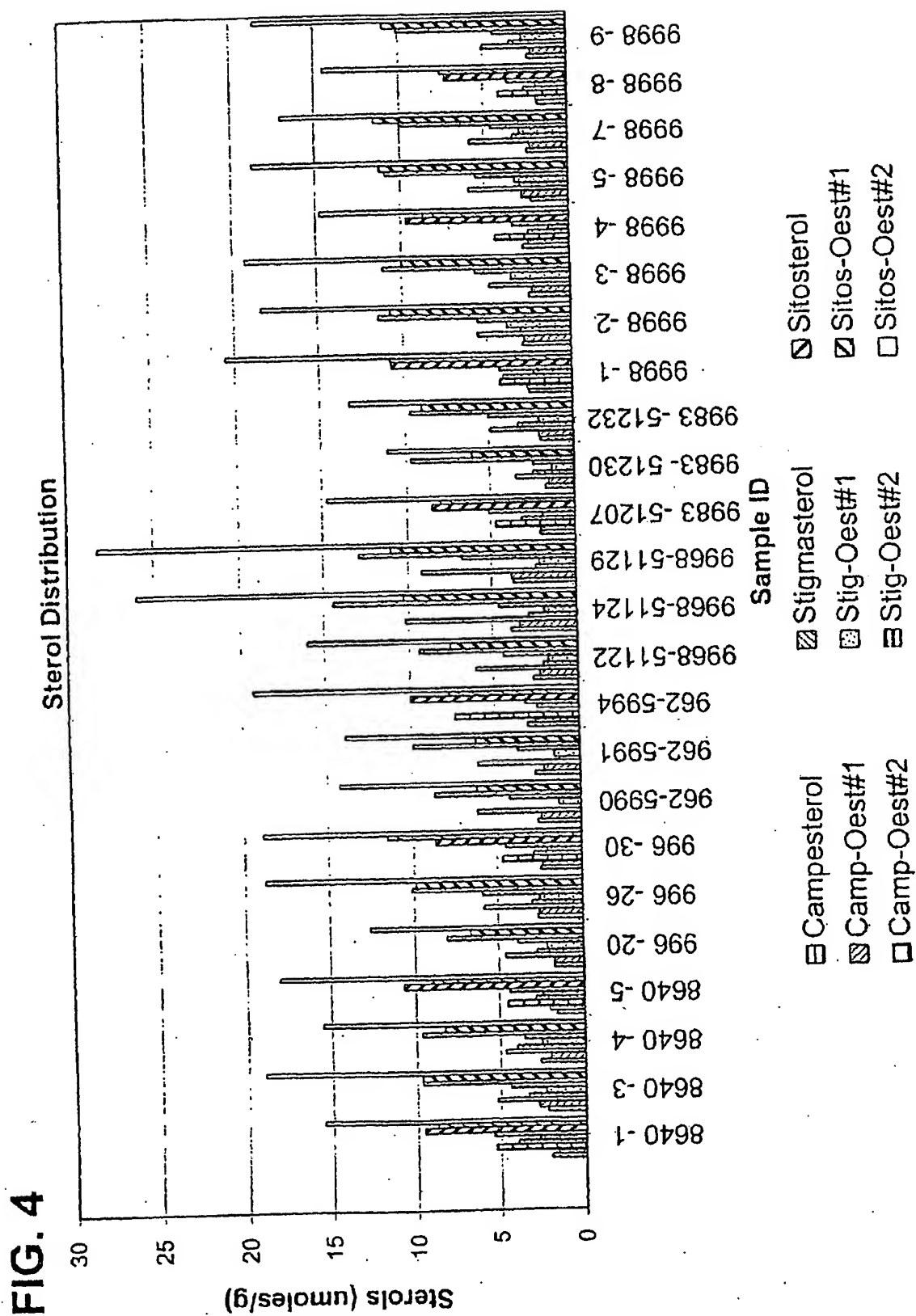
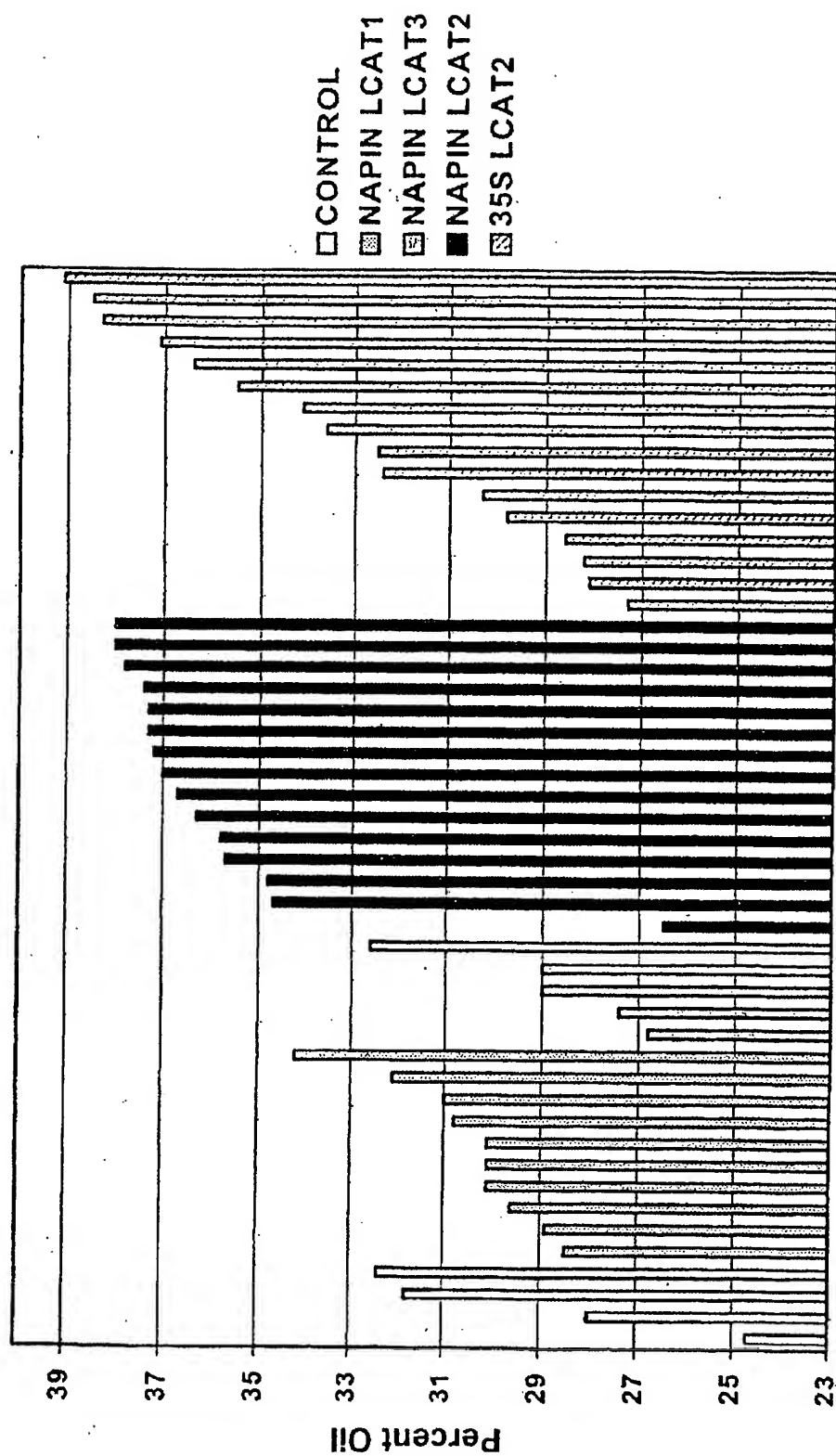
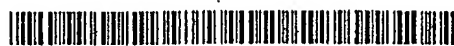


FIG. 5

NIR Analysis of LCAT



(19) World Intellectual Property Organization  
International Bureau



(43) International Publication Date  
8 March 2001 (08.03.2001)

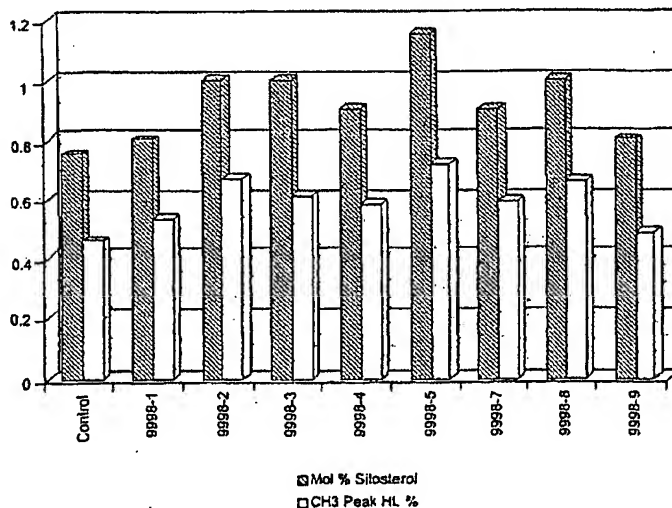
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(10) International Publication Number  
**WO 01/16308 A3**

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- (21) International Application Number: PCT/US00/23863
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(54) Title: PLANT STEROL ACYLTRANSFERASES



(57) Abstract: The present invention is directed to lecithin: cholesterol acyltransferase-like polypeptides (LCAT) and acyl CoA: cholesterol acyltransferases-like polypeptides (ACAT). The invention provides polynucleotides encoding such cholesterol: acyltransferases-like polypeptides, polypeptides encoded by such polynucleotides, and the use of such polynucleotides to alter sterol composition and oil production in plants and host cells. Also provided are oils produced by the plants and host cells containing the polynucleotides and food products, nutritional supplements, and pharmaceutical composition containing plants or oils of the present invention. The polynucleotides of the present invention include those derived from plant sources.

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# INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 00/23863

## A. CLASSIFICATION OF SUBJECT MATTER

IPC 7 C12N15/82

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	<p>HOBBS D H ET AL: "Cloning of a cDNA encoding diacylglycerol acyltransferase from Arabidopsis thaliana and its functional expression"</p> <p>FEBS LETTERS, NL, ELSEVIER SCIENCE PUBLISHERS, AMSTERDAM, vol. 452, no. 3, 11 June 1999 (1999-06-11), pages 145-149, XP002122747</p> <p>ISSN: 0014-5793 &amp; DATABASE EMBL [Online] EBI</p> <p>accession no. AJ131831.1, 10 June 1999 (1999-06-10) see sequence</p> <p style="text-align: center;">---</p> <p style="text-align: center;">-/--</p>	<p>1-4, 6, 7, 20-31, 33-50, 54-61, 63-77, 79-100, 102-112, 114-120</p>

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

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Date of the actual completion of the international search

27 February 2001

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# INTERNATIONAL SEARCH REPORT

International Application No  
PCT/US 00/23863

## C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	<p>DATABASE EMBL [Online] EBI accession no. AF164434, 26 July 1999 (1999-07-26) NYKIFORUK C.L. ET AL: "Brassica napus putative diacylglycerol acyltransferase (DGAT1) mRNA" XP002161573 see sequence</p>	<p>1-4,6,7, 20-31, 33-50, 54-61, 63-77, 79-100, 102-112, 114-120</p>
P,X	<p>WO 99 63096 A (LASSNER MICHAEL W ;RUEZINSKY DIANE M (US); CALGENE LLC (US)) 9 December 1999 (1999-12-09)</p> <p>claim 4; figure 1 &amp; DATABASE GENESEQ [Online] Derwent accession no. Z45371, 27 March 2000 (2000-03-27) see sequence</p>	<p>1-4,6,7, 20-31, 33-50, 54-61, 63-77, 79-100, 102-112, 114-120</p>
A	<p>FRENTZEN M (REPRINT): "Acyltransferases from basic science to modified seed oils" FETT - LIPID,WILEY-VCH VERLAG,WEINHEIM,DE, vol. 100, no. 4/05, May 1998 (1998-05), pages 161-166, XP002122744 ISSN: 0931-5985 the whole document</p>	
A	<p>TANIYAMA YOSHIO ET AL: "Cloning and expression of a novel lysophospholipase which structurally resembles lecithin cholesterol acyltransferase." BIOCHEMICAL AND BIOPHYSICAL RESEARCH COMMUNICATIONS, vol. 257, no. 1, 2 April 1999 (1999-04-02), pages 50-56, XP002161572 ISSN: 0006-291X abstract</p>	

# INTERNATIONAL SEARCH REPORT

International application No.  
PCT/US 00/23863

## Box I Observations where certain claims were found unsearchable (Continuation of Item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:  
because they relate to subject matter not required to be searched by this Authority, namely:
  
2. ☐ Claims Nos.:  
because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
  
3. ☐ Claims Nos.:  
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

## Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
  
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
  
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
  
4. ☒ No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

1-3(part)4(all), 6-7, 20-31, 33-50, 54-61, 63-77, 79-100, 102-112, 114-120(all part)

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
- ☐ No protest accompanied the payment of additional search fees.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

1. Claims: 1-3(part)4 (all),6-7,20-31,33-50,54-61,63-77,  
79-100,102-112,114-120(all part)

SEQ ID 42 and related subject matter.

2. Claims: 1-3,5-120

Groups 2 through 37 - SEQ Ids 2-75 as listed in claim 5 and  
related subject-matter.



Information on patent family members

PCT/US 00/23863

Form PCT/SA/210 (patent family annex) (July 1992)

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